

# Package ‘PharmacoGx’

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**Description** Contains a set of functions to perform large-scale analysis of pharmacogenomic data.

**License** Artistic-2.0

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amcc *Calculate an Adaptive Matthews Correlation Coefficient*

### Description

This function calculates an Adaptive Matthews Correlation Coefficient (AMCC) for two vectors of values identical length. It assumes the entries in the two vectors are paired. The Adaptive Matthews Correlation Coefficient for two vectors of values is defined as the Maximum Matthews Coefficient over all possible binary splits of the ranks of the two vectors. In this way, it calculates the best possible agreement of a binary classifier on the two vectors of data. #If the AMCC is low, then it is impossible to find any binary classification of the two vectors with a high degree of concordance.

### Usage

```
amcc(x, y, step.prct = 0, min.cat = 3, nperm = 1000, setseed = 12345,
      nthread = 1)
```

### Arguments

x, y	Two paired vectors of values. Could be replicates of observations for the same experiments for example.
step.prct	Instead of testing all possible splits of the data, it is possible to test steps of a percentage size of the total number of ranks in x/y. If this variable is 0, function defaults to testing all possible splits.
min.cat	The minimum number of members per category. Classifications with less members fitting into both categories will not be considered.
nperm	The number of perumutation to use for estimating significance. If 0, then no p-value is calculated.
setseed	Allows setting a consistant seed for reproducibility of permutation testing results. Defaults to 12345.
nthread	Number of threads to parallize over. Both the AMCC calculation and the permutation testing is done in parallel.

### Value

Returns a list with two elements. \$amcc contains the highest "mcc" value over all the splits, the p value, as well as the rank at which the split was done.

### Examples

```
x <- c(1,2,3,4,5,6,7)
y <- c(1,3,5,4,2,7,6)
amcc(x,y, min.cat=2)
```

availablePSets                      *Return a table of PharmacoSets available for download*

---

### Description

The function fetches a table of all PharmacoSets available for download from the PharmacoGx server. The table includes the names of the PharmacoSet, the types of data available in the object, and the date of last update.

### Usage

```
availablePSets(saveDir = file.path(".", "PSets"), myfn = "PSets.csv",  
              verbose = TRUE)
```

### Arguments

saveDir	character	Directory to save the table of PSets
myfn	character	The filename for the table of PSets
verbose	bool	Should status messages be printed during download.

### Value

A data.frame with details about the available PharmacoSet objects

### Examples

```
if (interactive()){  
  availablePSets()  
}
```

---

CCLEsmall                      *Cancer Cell Line Encyclopedia (CCLE) Example PharmacoSet*

---

### Description

A small example version of the CCLE PharmacoSet, used in the documentation examples. All credit for the data goes to the CCLE group at the Broad Institute. This is not a full version of the dataset, most of the dataset was removed to make runnable example code. For the full dataset, please download using the downloadPSet function.

### Usage

```
data(CCLEsmall)
```

### Format

PharmacoSet object

**References**

Barretina et al. The Cancer Cell Line Encyclopedia enables predictive modelling of anticancer drug sensitivity. Nature, 2012

---

cellInfo	<i>cellInfo Generic</i>
----------	-------------------------

---

**Description**

Generic for cellInfo method

**Usage**

```
cellInfo(pSet)
```

**Arguments**

pSet                    The PharmacoS<sub>e</sub>t to retrieve cell info from

**Value**

a data.frame with the cell annotations

**Examples**

```
data(CCLEsmall)
cellInfo(CCLEsmall)
```

---

cellInfo<-	<i>cellInfo&lt;- Generic</i>
------------	------------------------------

---

**Description**

Generic for cellInfo replace method

**Usage**

```
cellInfo(object) <- value
```

**Arguments**

object                    The PharmacoS<sub>e</sub>t to replace cell info in  
value                    A data.frame with the new cell annotations

**Value**

Updated PharmacoS<sub>e</sub>t

**Examples**

```
data(CCLEsmall)
cellInfo(CCLEsmall) <- cellInfo(CCLEsmall)
```

---

<code>cellNames</code>	<i>cellNames Generic</i>
------------------------	--------------------------

---

**Description**

A generic for the `cellNames` method

**Usage**

```
cellNames(pSet)
```

**Arguments**

`pSet`            The PharmacoSet to return cell names from

**Value**

A vector of the cell names used in the PharmacoSet

**Examples**

```
data(CCLEsmall)
cellNames(CCLEsmall)
```

---

<code>cellNames&lt;-</code>	<i>cellNames&lt;- Generic</i>
-----------------------------	-------------------------------

---

**Description**

A generic for the `cellNames` replacement method

**Usage**

```
cellNames(object) <- value
```

**Arguments**

`object`            The PharmacoSet to update  
`value`             A character vector of the new cell names

**Value**

Updated PharmacoSet

**Examples**

```
data(CCLEsmall)
cellNames(CCLEsmall) <- cellNames(CCLEsmall)
```

---

checkPSetStructure     *A function to verify the structure of a PharmacoSet*

---

**Description**

This function checks the structure of a PharmacoSet, ensuring that the correct annotations are in place and all the required slots are filled so that matching of cells and drugs can be properly done across different types of data and with other studies.

**Usage**

```
checkPSetStructure(pSet, plotDist = FALSE, result.dir = ".")
```

**Arguments**

pSet	A PharmacoSet to be verified
plotDist	Should the function also plot the distribution of molecular data?
result.dir	The path to the directory for saving the plots as a string

**Value**

Prints out messages whenever describing the errors found in the structure of the pset object passed in.

**Examples**

```
data(CCLEsmall)
checkPSetStructure(CCLEsmall)
```

---

CMAFsmall     *Connectivity Map Example PharmacoSet*

---

**Description**

A small example version of the Connectivity Map PharmacoSet, used in the documentation examples. All credit for the data goes to the Connectivity Map group at the Broad Institute. This is not a full version of the dataset, most of the dataset was removed to make runnable example code. For the full dataset, please download using the downloadPSet function.

**Usage**

```
data(CMAFsmall)
```

**Format**

PharmacoSeset object

**References**

Lamb et al. The Connectivity Map: using gene-expression signatures to connect small molecules, genes, and disease. Science, 2006.

---

computeABC	<i>Fits dose-response curves to data given by the user and returns the ABC of the fitted curves.</i>
------------	--

---

**Description**

Fits dose-response curves to data given by the user and returns the ABC of the fitted curves.

**Usage**

```
computeABC(conc1, conc2, viability1, viability2, Hill_fit1, Hill_fit2,
  conc_as_log = FALSE, viability_as_pct = TRUE, trunc = TRUE,
  verbose = TRUE)
```

**Arguments**

conc1	[vector] is a vector of drug concentrations.
conc2	[vector] is a vector of drug concentrations.
viability1	[vector] is a vector whose entries are the viability values observed in the presence of the drug concentrations whose logarithms are in the corresponding entries of conc1, expressed as percentages of viability in the absence of any drug.
viability2	[vector] is a vector whose entries are the viability values observed in the presence of the drug concentrations whose logarithms are in the corresponding entries of conc2, expressed as percentages of viability in the absence of any drug.
Hill_fit1	[list or vector] In the order: c("Hill Slope", "E_inf", "EC50"), the parameters of a Hill Slope as returned by logLogisticRegression. If conc_as_log is set then the function assumes logEC50 is passed in, and if viability_as_pct flag is set, it assumes E_inf is passed in as a percent. Otherwise, E_inf is assumed to be a decimal, and EC50 as a concentration.
Hill_fit2	[list or vector] In the order: c("Hill Slope", "E_inf", "EC50"), the parameters of a Hill Slope as returned by logLogisticRegression. If conc_as_log is set then the function assumes logEC50 is passed in, and if viability_as_pct flag is set, it assumes E_inf is passed in as a percent. Otherwise, E_inf is assumed to be a decimal, and EC50 as a concentration.
conc_as_log	[logical], if true, assumes that log10-concentration data has been given rather than concentration data.
viability_as_pct	[logical], if false, assumes that viability is given as a decimal rather than a percentage, and returns ABC as a decimal. Otherwise, viability is interpreted as percent, and AUC is returned 0-100.
trunc	[logical], if true, causes viability data to be truncated to lie between 0 and 1 before curve-fitting is performed.
verbose	[logical], if true, causes warnings thrown by the function to be printed.



**Value**

The numeric area of the absolute difference between the two hill slopes

**Examples**

```
dose1 <- c("0.0025", "0.008", "0.025", "0.08", "0.25", "0.8", "2.53", "8")
viability1 <- c("108.67", "111", "102.16", "100.27", "90", "87", "74", "57")
dose2 <- c("0.0025", "0.008", "0.025", "0.08", "0.25", "0.8", "2.53", "8")
viability2 <- c("100.94", "112.5", "86", "104.16", "75", "68", "48", "29")
computeABC(dose1, dose2, viability1, viability2)
```

---

computeAmax	<i>Fits dose-response curves to data given by the user and returns the Amax of the fitted curve. Amax: 100 - viability at maximum concentration (in fitted curve)</i>
-------------	---

---

**Description**

Fits dose-response curves to data given by the user and returns the Amax of the fitted curve. Amax: 100 - viability at maximum concentration (in fitted curve)

**Usage**

```
computeAmax(concentration, viability, trunc = TRUE, verbose = FALSE)
```

**Arguments**

concentration	[vector] is a vector of drug concentrations.
viability	[vector] is a vector whose entries are the viability values observed in the presence of the drug concentrations whose logarithms are in the corresponding entries of the log_conc, expressed as percentages of viability in the absence of any drug.
trunc	[logical], if true, causes viability data to be truncated to lie between 0 and 1 before curve-fitting is performed.
verbose	[logical] should warnings be printed

**Value**

The numerical Amax

**Examples**

```
dose <- c("0.0025", "0.008", "0.025", "0.08", "0.25", "0.8", "2.53", "8")
viability <- c("108.67", "111", "102.16", "100.27", "90", "87", "74", "57")
computeAmax(dose, viability)
```

---

 computeAUC

*Computes the AUC for a Drug Dose Viability Curve*


---

### Description

Returns the AUC (Area Under the drug response Curve) given concentration and viability as input, normalized by the concentration range of the experiment. The area returned is the response (1-Viability) area, i.e. area under the curve when the response curve is plotted on a log10 concentration scale, with high AUC implying high sensitivity to the drug. The function can calculate both the area under a fitted Hill Curve to the data, and a trapz numeric integral of the actual data provided. Alternatively, the parameters of a Hill Slope returned by logLogisticRegression can be passed in if they already known.

### Usage

```
computeAUC(concentration, viability, Hill_fit, conc_as_log = FALSE,
  viability_as_pct = TRUE, trunc = TRUE, area.type = c("Fitted",
  "Actual"), verbose = TRUE)
```

### Arguments

concentration	[vector] is a vector of drug concentrations.
viability	[vector] is a vector whose entries are the viability values observed in the presence of the drug concentrations whose logarithms are in the corresponding entries of conc, where viability 0 indicates that all cells died, and viability 1 indicates that the drug had no effect on the cells.
Hill_fit	[list or vector] In the order: c("Hill Slope", "E_inf", "EC50"), the parameters of a Hill Slope as returned by logLogisticRegression. If conc_as_log is set then the function assumes logEC50 is passed in, and if viability_as_pct flag is set, it assumes E_inf is passed in as a percent. Otherwise, E_inf is assumed to be a decimal, and EC50 as a concentration.
conc_as_log	[logical], if true, assumes that log10-concentration data has been given rather than concentration data.
viability_as_pct	[logical], if false, assumes that viability is given as a decimal rather than a percentage, and returns AUC as a decimal. Otherwise, viability is interpreted as percent, and AUC is returned 0-100.
trunc	[logical], if true, causes viability data to be truncated to lie between 0 and 1 before curve-fitting is performed.
area.type	Should the area be computed using the actual data ("Actual"), or a fitted curve ("Fitted")
verbose	[logical], if true, causes warnings thrown by the function to be printed.

### Value

Numeric AUC value

**Examples**

```
dose <- c("0.0025", "0.008", "0.025", "0.08", "0.25", "0.8", "2.53", "8")
viability <- c("108.67", "111", "102.16", "100.27", "90", "87", "74", "57")
computeAUC(dose, viability)
```

---

computeIC50

*Computes the IC<sub>n</sub> for any n in 0-100 for a Drug Dose Viability Curve*


---

**Description**

Returns the IC<sub>n</sub> for any given nth percentile when given concentration and viability as input, normalized by the concentration range of the experiment. A Hill Slope is first fit to the data, and the IC<sub>n</sub> is inferred from the fitted curve. Alternatively, the parameters of a Hill Slope returned by `logLogisticRegression` can be passed in if they already known.

**Usage**

```
computeIC50(concentration, viability, Hill_fit, conc_as_log = FALSE,
            viability_as_pct = TRUE, verbose = TRUE, trunc = TRUE)
```

```
computeICn(concentration, viability, Hill_fit, n, conc_as_log = FALSE,
            viability_as_pct = TRUE, verbose = TRUE, trunc = TRUE)
```

**Arguments**

- |                  |  |
|------------------|--|
| concentration    | [vector] is a vector of drug concentrations.   |
| viability        | [vector] is a vector whose entries are the viability values observed in the presence of the drug concentrations whose logarithms are in the corresponding entries of conc, where viability 0 indicates that all cells died, and viability 1 indicates that the drug had no effect on the cells.  |
| Hill_fit         | [list or vector] In the order: c("Hill Slope", "E_inf", "EC50"), the parameters of a Hill Slope as returned by <code>logLogisticRegression</code> . If <code>conc_as_log</code> is set then the function assumes <code>logEC50</code> is passed in, and if <code>viability_as_pct</code> flag is set, it assumes <code>E_inf</code> is passed in as a percent. Otherwise, <code>E_inf</code> is assumed to be a decimal, and <code>EC50</code> as a concentration. |
| conc_as_log      | [logical], if true, assumes that log10-concentration data has been given rather than concentration data, and that <code>log10(IC<sub>n</sub>)</code> should be returned instead of IC <sub>n</sub> .   |
| viability_as_pct | [logical], if false, assumes that viability is given as a decimal rather than a percentage, and that <code>E_inf</code> passed in as decimal.  |
| verbose          | [logical], if true, causes warnings thrown by the function to be printed.  |
| trunc            | [logical], if true, causes viability data to be truncated to lie between 0 and 1 before curve-fitting is performed.  |
| n                | [numeric] The percentile concentration to compute. If <code>viability_as_pct</code> set, assumed to be percentage, otherwise assumed to be a decimal value.  |

**Value**

a numeric value for the concentration of the nth percentile viability reduction

**Functions**

- `computeIC50`: Returns the IC50 of a Drug Dose response curve

**Examples**

```
dose <- c("0.0025", "0.008", "0.025", "0.08", "0.25", "0.8", "2.53", "8")
viability <- c("108.67", "111", "102.16", "100.27", "90", "87", "74", "57")
computeIC50(dose, viability)
computeICn(dose, viability, n=10)
```

---

`computeSlope`

*Return Slope (normalized slope of the drug response curve) for an experiment of a pSet by taking its concentration and viability as input.*

---

**Description**

Return Slope (normalized slope of the drug response curve) for an experiment of a pSet by taking its concentration and viability as input.

**Usage**

```
computeSlope(concentration, viability, trunc = TRUE, verbose = TRUE)
```

**Arguments**

<code>concentration</code>	[vector] A concentration range that the AUC should be computed for that range. Concentration range by default considered as not logarithmic scaled. Converted to numeric by function if necessary.
<code>viability</code>	[vector] Viabilities corresponding to the concentration range passed as first parameter. The range of viability values by definition should be between 0 and 100. But the viabilities greater than 100 and lower than 0 are also accepted.
<code>trunc</code>	[binary] A flag that identify if the viability values should be truncated to be in the range of (0,100)
<code>verbose</code>	[boolean] If 'TRUE' the function will retrun warnings and other infomrative messages.

**Value**

Returns the normalized linear slope of the drug response curve

**Examples**

```
dose <- c("0.0025", "0.008", "0.025", "0.08", "0.25", "0.8", "2.53", "8")
viability <- c("108.67", "111", "102.16", "100.27", "90", "87", "74", "57")
computeSlope(dose, viability)
```

---

connectivityScore      *Function computing connectivity scores between two signatures*

---

### Description

A function for finding the connectivity between two signatures, using either the GSEA method based on the KS statistic, or the gwc method based on a weighted spearman statistic. The GSEA analysis is implemented in the piano package.

### Usage

```
connectivityScore(x, y, method = c("gsea", "fgsea", "gwc"), nperm = 10000,
  nthread = 1, gwc.method = c("spearman", "pearson"), ...)
```

### Arguments

x	A matrix with the first gene signature. In the case of GSEA the vector of values per gene for GSEA in which we are looking for an enrichment. In the case of gwc, this should be a matrix, with the per gene responses in the first column, and the significance values in the second.
y	A matrix with the second signature. In the case of GSEA, this is the vector of up and down regulated genes we are looking for in our signature, with the direction being determined from the sign. In the case of gwc, this should be a matrix of identical size to x, once again with the per gene responses in the first column, and their significance in the second.
method	character string identifying which method to use, out of 'gsea' and 'gwc'
nperm	numeric, how many permutations should be done to determine significance through permutation testing? The minimum is 100, default is 1e4.
nthread	numeric, how many cores to run parallel processing on.
gwc.method	character, should gwc use a weighted spearman or pearson statistic?
...	Additional arguments passed down to gsea and gwc functions

### Value

numeric a numeric vector with the score and the p-value associated with it

### References

F. Pozzi, T. Di Matteo, T. Aste, "Exponential smoothing weighted correlations", The European Physical Journal B, Vol. 85, No 6, 2012. DOI: 10.1140/epjb/e2012-20697-x

Varemo, L., Nielsen, J. and Nookaew, I. (2013) Enriching the gene set analysis of genome-wide data by incorporating directionality of gene expression and combining statistical hypotheses and methods. Nucleic Acids Research. 41 (8), 4378-4391. doi: 10.1093/nar/gkt111

**Examples**

```
xValue <- c(1,5,23,4,8,9,2,19,11,12,13)
xSig <- c(0.01, 0.001, .97, 0.01,0.01,0.28,0.7,0.01,0.01,0.01,0.01)
yValue <- c(1,5,10,4,8,19,22,19,11,12,13)
ySig <- c(0.01, 0.001, .97,0.01, 0.01,0.78,0.9,0.01,0.01,0.01,0.01)
xx <- cbind(xValue, xSig)
yy <- cbind(yValue, ySig)
rownames(xx) <- rownames(yy) <- c('1','2','3','4','5','6','7','8','9','10','11')
data.cor <- connectivityScore(xx,yy,method="gwc", gwc.method="spearman", nperm=300)
```

cosinePerm

*Computes the cosine similarity and significance using permutation test***Description**

Computes the cosine similarity and significance using permutation test

**Usage**

```
cosinePerm(x, y, nperm = 1000, alternative = c("two.sided", "less",
"greater"), include.perm = FALSE, setseed = 12345, nthread = 1)
```

**Arguments**

x	[factor] is the factors for the first variable
y	[factor] is the factors for the second variable
nperm	[integer] is the number of permutations to compute the null distribution of MCC estimates
alternative	[string] indicates the alternative hypothesis and must be one of "two.sided", "greater" or "less". You can specify just the initial letter. "greater" corresponds to positive association, "less" to negative association. Options are "two.sided", "less", or "greater"
include.perm	[boolean] indicates whether the estimates for the null distribution should be returned. Default set to 'FALSE'
setseed	[integer] is the seed specified by the user. Defaults is '12345'
nthread	[integer] is the number of threads to be used to perform the permutations in parallel

**Value**

list estimate of the cosine similarity, p-value and estimates after random permutations (null distribution) in include.perm is set to 'TRUE'

**Examples**

```
x <- factor(c(1,2,1,2,1))
y <- factor(c(2,2,1,1,1))
cosinePerm(x, y)
```

---

dateCreated	<i>dateCreated Generic</i>
-------------	----------------------------

---

**Description**

A generic for the dateCreated method

**Usage**

```
dateCreated(pSet)
```

**Arguments**

pSet	A PharmacoSet
------	---------------

**Value**

The date the PharmacoSet was created

**Examples**

```
data(CCLEsmall)
dateCreated(CCLEsmall)
```

---

dim, PharmacoSet-method

*Get the dimensions of a PharmacoSet*

---

**Description**

Get the dimensions of a PharmacoSet

**Usage**

```
## S4 method for signature 'PharmacoSet'
dim(x)
```

**Arguments**

x	PharmacoSet
---	-------------

**Value**

A named vector with the number of Cells and Drugs in the PharmacoSet

---

downloadPertSig      *Download Drug Perturbation Signatures*

---

### Description

This function allows you to download an array of drug perturbation signatures, as would be computed by the `drugPerturbationSig` function, for the available perturbation `PharmacoSets`. This function allows the user to skip these very lengthy calculation steps for the datasets available, and start their analysis from the already computed signatures

### Usage

```
downloadPertSig(name, saveDir = file.path(".", "Psets", "Sigs"),
  myfn = NULL, verbose = TRUE)
```

### Arguments

name	Character string, the name of the <code>PharmacoSet</code> for which to download signatures. The name should match the names returned in the <code>availablePsets</code> table.
saveDir	Character string with the folder path where the <code>PharmacoSet</code> should be saved. Defaults to <code>"/Psets/Sigs/"</code> . Will create directory if it does not exist.
myfn	character string, the file name to save the dataset under
verbose	bool Should status messages be printed during download. Defaults to <code>TRUE</code> .

### Value

An array type object containing the signatures

### Examples

```
if (interactive()){
  downloadPertSig("CMAP")
}
```

---

downloadPSet      *Download a PharmacoSet object*

---

### Description

This function allows you to download a `PharmacoSet` object for use with this package. The `PharmacoSets` have been extensively curated and organised within a `PharmacoSet` class, enabling use with all the analysis tools provided in `PharmacoGx`.

### Usage

```
downloadPSet(name, saveDir = file.path(".", "Psets"), pSetFileName = NULL,
  verbose = TRUE)
```



**Arguments**

name	Character string, the name of the PhamracoSet to download.
saveDir	Character string with the folder path where the PharmacoSet should be saved. Defaults to './Psets/'. Will create directory if it does not exist.
pSetFileName	character string, the file name to save the dataset under
verbose	bool Should status messages be printed during download. Defaults to TRUE.

**Value**

A PSet object with the dataset, downloaded from our server

**Examples**

```
if (interactive()){
  downloadPSet("CMAF")
}
```

---

drugDoseResponseCurve *Plot drug response curve of a given drug and a given cell for a list of pSets (objects of the PharmacoSet class).*

---

**Description**

Given a list of PharmacoSets, the function will plot the drug\_response curve, for a given drug/cell pair. The y axis of the plot is the viability percentage and x axis is the log transformed concentrations. If more than one pSet is provided, a light gray area would show the common concentration range between pSets. User can ask for type of sensitivity measurement to be shown in the plot legend. The user can also provide a list of their own concentrations and viability values, as in the examples below, and it will be treated as experiments equivalent to values coming from a pset. The names of the concentration list determine the legend labels.

**Usage**

```
drugDoseResponseCurve(drug, cellline, pSets = list(),
  concentrations = list(), viabilities = list(), conc_as_log = FALSE,
  viability_as_pct = TRUE, trunc = TRUE,
  legends.label = c("ic50_published", "gi50_published", "auc_published",
  "auc_recomputed", "ic50_recomputed"), ylim = c(0, 100), xlim, mycol, title,
  plot.type = c("Fitted", "Actual", "Both"), summarize.replicates = TRUE,
  lwd = 0.5, cex = 0.7, cex.main = 0.9, legend.loc = "topright",
  verbose = TRUE)
```

**Arguments**

drug	[string] A drug name for which the drug response curve should be plotted. If the plot is desirable for more than one pharmaco set, A unique drug id should be provided.
cellline	[string] A cell line name for which the drug response curve should be plotted. If the plot is desirable for more than one pharmaco set, A unique cell id should be provided.

<code>pSets</code>	[list] a list of <code>PharmacoSet</code> objects, for which the function should plot the curves.
<code>concentrations, viabilities</code>	[list] A list of concentrations and viabilities to plot, the function assumes that <code>concentrations[[i]]</code> is plotted against <code>viabilities[[i]]</code> . The names of the concentration list are used to create the legend labels
<code>conc_as_log</code>	[logical], if true, assumes that log10-concentration data has been given rather than concentration data, and that <code>log10(ICn)</code> should be returned instead of <code>ICn</code> . Applies only to the concentrations parameter.
<code>viability_as_pct</code>	[logical], if false, assumes that viability is given as a decimal rather than a percentage, and that <code>E_inf</code> passed in as decimal. Applies only to the viabilities parameter.
<code>trunc</code>	[bool] Should the viability values be truncated to lie in [0-100] before doing the fitting
<code>legends.label</code>	[vector] A vector of sensitivity measurement types which could be any combination of <code>ic50_published</code> , <code>auc_published</code> , <code>auc_recomputed</code> and <code>auc_recomputed_star</code> . A legend will be displayed on the top right of the plot which each line of the legend is the values of requested sensitivity measurements for one of the requested <code>pSets</code> . If this parameter is missed no legend would be provided for the plot.
<code>ylim</code>	[vector] A vector of two numerical values to be used as <code>ylim</code> of the plot. If this parameter would be missed <code>c(0,100)</code> would be used as the <code>ylim</code> of the plot.
<code>xlim</code>	[vector] A vector of two numerical values to be used as <code>xlim</code> of the plot. If this parameter would be missed the minimum and maximum concentrations between all the <code>pSets</code> would be used as plot <code>xlim</code> .
<code>mycol</code>	[vector] A vector with the same length of the <code>pSets</code> parameter which will determine the color of the curve for the pharmac sets. If this parameter is missed default colors from <code>Rcolorbrewer</code> package will be used as curves color.
<code>title</code>	[character] The title of the graph. If no title is provided, then it defaults to <code>'Drug':'Cell Line'</code> .
<code>plot.type</code>	[character] Plot type which can be the actual one ("Actual") or the one fitted by log1 logistic regression ("Fitted") or both of them ("Both"). If this parameter is missed by default actual curve is plotted.
<code>summarize.replicates</code>	[character] If this parameter is set to true replicates are summarized and replicates are plotted individually otherwise
<code>lwd</code>	[numeric] The line width to plot with
<code>cex</code>	[numeric] The <code>cex</code> parameter passed to plot
<code>cex.main</code>	[numeric] The <code>cex.main</code> parameter passed to plot, controls the size of the titles
<code>legend.loc</code>	And argument passable to <code>xy.coords</code> for the position to place the legend.
<code>verbose</code>	[boolean] Should warning messages about the data passed in be printed?

**Value**

Plots to the active graphics device and returns and invisible `NULL`.

**Examples**

```
if (interactive()) {
  drugDoseResponseCurve(concentrations=list("Experiment 1"=c(.008, .04, .2, 1)),
    viabilities=list(c(100,50,30,1)), plot.type="Both")
}
```

---

drugInfo

*drugInfo Generic*


---

**Description**

Generic for drugInfo method

**Usage**

```
drugInfo(pSet)
```

**Arguments**

pSet                    The PharmacoSet to retrieve drug info from

**Value**

a data.frame with the drug annotations

**Examples**

```
data(CCLEsmall)
drugInfo(CCLEsmall)
```

---

drugInfo<-

*drugInfo<- Generic*


---

**Description**

Generic for drugInfo replace method

**Usage**

```
drugInfo(object) <- value
```

**Arguments**

object                    The PharmacoSet to replace drug info in  
value                      A data.frame with the new drug annotations

**Value**

Updated PharmacoSet

**Examples**

```
data(CCLEsmall)
drugInfo(CCLEsmall) <- drugInfo(CCLEsmall)
```

---

drugNames	<i>drugNames Generic</i>
-----------	--------------------------

---

**Description**

A generic for the drugNames method

**Usage**

```
drugNames(pSet)
```

**Arguments**

pSet            The PharmacoSet to return drug names from

**Value**

A vector of the drug names used in the PharmacoSet

**Examples**

```
data(CCLEsmall)
drugNames(CCLEsmall)
```

---

drugNames<-	<i>drugNames&lt;- Generic</i>
-------------	-------------------------------

---

**Description**

A generic for the drugNames replacement method

**Usage**

```
drugNames(object) <- value
```

**Arguments**

object            The PharmacoSet to update  
value            A character vector of the new drug names

**Value**

Updated PharmacoSet

**Examples**

```
data(CCLEsmall)
drugNames(CCLEsmall) <- drugNames(CCLEsmall)
```

---

**drugPerturbationSig**     *Creates a signature representing gene expression (or other molecular profile) change induced by administrating a drug, for use in drug effect analysis.*

---

**Description**

Given a Pharmacoset of the perturbation experiment type, and a list of drugs, the function will compute a signature for the effect of drug concentration on the molecular profile of a cell. The algorithm uses a regression model which corrects for experimental batch effects, cell specific differences, and duration of experiment to isolate the effect of the concentration of the drug applied. The function returns the estimated coefficient for concentration, the t-stat, the p-value and the false discovery rate associated with that coefficient, in a 3 dimensional array, with genes in the first direction, drugs in the second, and the selected return values in the third.

**Usage**

```
drugPerturbationSig(pSet, mDataType, drugs, cells, features, nthread = 1,
  returnValues = c("estimate", "tstat", "pvalue", "fdr"), verbose = FALSE)
```

**Arguments**

pSet	[PharmacoSet] a PharmacoSet of the perturbation experiment type
mDataType	[character] which one of the molecular data types to use in the analysis, out of dna, rna, rnaseq, snp, cnv
drugs	[character] a vector of drug names for which to compute the signatures. Should match the names used in the PharmacoSet.
cells	[character] a vector of cell names to use in computing the signatures. Should match the names used in the PharmacoSet.
features	[character] a vector of features for which to compute the signatures. Should match the names used in correspondant molecular data in PharmacoSet.
nthread	[numeric] if multiple cores are available, how many cores should the computation be parallelized over?
returnValues	[character] Which of estimate, t-stat, p-value and fdr should the function return for each gene drug pair?
verbose	[bool] Should diagnostic messages be printed? (default false)

**Value**

list a 3D array with genes in the first dimension, drugs in the second, and return values in the third.

**Examples**

```
data(CMAPsmall)
drug.perturbation <- drugPerturbationSig(CMAPsmall, mDataType="rna", nthread=1)
print(drug.perturbation)
```

---

drugSensitivitySig	<i>Creates a signature representing the association between gene expression (or other molecular profile) and drug dose response, for use in drug sensitivity analysis.</i>
--------------------	--

---

## Description

Given a Pharmacoset of the sensitivity experiment type, and a list of drugs, the function will compute a signature for the effect gene expression on the molecular profile of a cell. The function returns the estimated coefficient, the t-stat, the p-value and the false discovery rate associated with that coefficient, in a 3 dimensional array, with genes in the first direction, drugs in the second, and the selected return values in the third.

## Usage

```
drugSensitivitySig(pSet, mDataType, drugs, features,
  sensitivity.measure = "auc_recomputed", molecular.summary.stat = c("mean",
    "median", "first", "last", "or", "and"),
  sensitivity.summary.stat = c("mean", "median", "first", "last"),
  returnValues = c("estimate", "pvalue", "fdr"), sensitivity.cutoff,
  standardize = c("SD", "rescale", "none"), molecular.cutoff = NA,
  molecular.cutoff.direction = c("less", "greater"), nthread = 1,
  verbose = TRUE, ...)
```

## Arguments

pSet	[PharmacoSet] a PharmacoSet of the perturbation experiment type
mDataType	[character] which one of the molecular data types to use in the analysis, out of dna, rna, rnaseq, snp, cnv
drugs	[character] a vector of drug names for which to compute the signatures. Should match the names used in the PharmacoSet.
features	[character] a vector of features for which to compute the signatures. Should match the names used in correspondant molecular data in PharmacoSet.
sensitivity.measure	[character] which measure of the drug dose sensitivity should the function use for its computations? Use the sensitivityMeasures function to find out what measures are available for each PSet.
molecular.summary.stat	What summary statistic should be used to summarize duplicates for cell line molecular profile measurements?
sensitivity.summary.stat	What summary statistic should be used to summarize duplicates for cell line sensitivity measurements?
returnValues	[character] Which of estimate, t-stat, p-value and fdr should the function return for each gene drug pair?
sensitivity.cutoff	[numeric] Allows the user to binarize the sensitivity data using this threshold.

standardize	[character] One of "SD", "rescale", or "none", for the form of standardization of the data to use. If "SD", the the data is scaled so that SD = 1. If rescale, then the data is scaled so that the 95 interquantile range lies in [0,1]. If none no rescaling is done.
molecular.cutoff	Allows the user to binarize the sensitivity data using this threshold.
molecular.cutoff.direction	[character] One of "less" or "greater", allows to set direction of binarization.
nthread	[numeric] if multiple cores are available, how many cores should the computation be parallelized over?
verbose	[boolean] 'TRUE' if the warnings and other infomrative message should be displayed
...	additional arguments not currently fully supported by the function

**Value**

list a 3D array with genes in the first dimension, drugs in the second, and return values in the third.

**Examples**

```
data(GDSCsmall)
drug.sensitivity <- drugSensitivitySig(GDSCsmall, mDataType="rna",
                                     nthread=1, features = fNames(GDSCsmall, "rna")[1])
print(drug.sensitivity)
```

---

featureInfo	<i>featureInfo Generic</i>
-------------	----------------------------

---

**Description**

Generic for featureInfo method

**Usage**

```
featureInfo(pSet, mDataType)
```

**Arguments**

pSet	The PharmacoSet to retrieve feature annotations from
mDataType	the type of molecular data

**Value**

a data.frame with the experiment info

**Examples**

```
data(CCLEsmall)
featureInfo(CCLEsmall, "rna")
```

---

```
featureInfo<-          featureInfo<- Generic
```

---

**Description**

Generic for featureInfo replace method

**Usage**

```
featureInfo(object, mDataType) <- value
```

**Arguments**

object	The PharmacoS <sub>et</sub> to replace gene annotations in
mDataType	The type of molecular data to be updated
value	A data.frame with the new feature annotations

**Value**

Updated PharmacoS<sub>et</sub>

**Examples**

```
data(CCLEsmall)
featureInfo(CCLEsmall, "rna") <- featureInfo(CCLEsmall, "rna")
```

---

filterNoisyCurves	<i>Viability measurements in dose-reponse curves must remain stable or decrease monotonically reflecting response to the drug being tested. filterNoisyCurves flags dose-response curves that strongly violate these assumptions.</i>
-------------------	---

---

**Description**

Viability measurements in dose-reponse curves must remain stable or decrease monotonically reflecting response to the drug being tested. filterNoisyCurves flags dose-response curves that strongly violate these assumptions.

**Usage**

```
filterNoisyCurves(pSet, epsilon = 25, positive.cutoff.percent = 0.8,
  mean.viability = 200, nthread = 1)
```



**Arguments**

pSet	[PharmacoSet] a PharmacoSet object
epsilon	[numeric] a value indicates assumed threshold for the distance between to consecutive viability values on the drug-response curve in the analysis, out of dna, rna, rnaseq, snp, cnv
positive.cutoff.percent	[numeric] This value indicates that function may violate epsilon rule for how many points on drug-response curve
mean.viability	[numeric] average expected viability value
nthread	[numeric] if multiple cores are available, how many cores should the computation be parallelized over?

**Value**

a list with two elements 'noisy' containing the rownames of the noisy curves, and 'ok' containing the rownames of the non-noisy curves

**Examples**

```
data(GDSCsmall)
filterNoisyCurves(GDSCsmall)
```

---

fNames	<i>fNames Generic</i>
--------	-----------------------

---

**Description**

A generic for the fNames method

**Usage**

```
fNames(pSet, mDataType)
```

**Arguments**

pSet	The PharmacoSet
mDataType	The molecular data type to return feature names for

**Value**

A character vector of the feature names

**Examples**

```
data(CCLEsmall)
fNames(CCLEsmall, "rna")
```

---

 GDSCsmall

*Genomics of Drug Sensitivity in Cancer Example PharmacoSet*


---

**Description**

A small example version of the Genomics of Drug Sensitivity in Cancer Project PharmacoSet, used in the documentation examples. All credit for the data goes to the Genomics of Drug Sensitivity in Cancer Project group at the Sanger. This is not a full version of the dataset, most of the dataset was removed to make runnable example code. For the full dataset, please download using the `downloadPSet` function.

**Usage**

```
data(GDSCsmall)
```

**Format**

PharmacoSet object

**References**

Garnett et al. Systematic identification of genomic markers of drug sensitivity in cancer cells. *Nature*, 2012.

---

 gwc

*Calculate the gwc score between two vectors, using either a weighted spearman or pearson correlation*


---

**Description**

Calculate the gwc score between two vectors, using either a weighted spearman or pearson correlation

**Usage**

```
gwc(x1, p1, x2, p2, method.cor = c("pearson", "spearman"), nperm = 10000,
    truncate.p = 1e-16, ...)
```

**Arguments**

x1	numeric vector of effect sizes (e.g., fold change or t statistics) for the first experiment
p1	numeric vector of p-values for each corresponding effect size for the first experiment
x2	numeric effect size (e.g., fold change or t statistics) for the second experiment
p2	numeric vector of p-values for each corresponding effect size for the second experiment
method.cor	character string identifying if a pearson or spearman correlation should be used

nperm            numeric how many permutations should be done to determine  
truncate.p        numeric Truncation value for extremely low p-values  
...                Other passed down to internal functions

**Value**

numeric a vector of two values, the correlation and associated p-value.

**Examples**

```
data(CCLEsmall)
x <- molecularProfiles(CCLEsmall,"rna")[,1]
y <- molecularProfiles(CCLEsmall,"rna")[,2]
x_p <- rep(0.05, times=length(x))
y_p <- rep(0.05, times=length(y))
names(x_p) <- names(x)
names(y_p) <- names(y)
gwc(x,x_p,y,y_p, nperm=100)
```

---

HDAC\_genes

*HDAC Gene Signature*


---

**Description**

A gene signature for HDAC inhibitors, as detailed by Glaser et al. The signature is mapped from the probe to gene level using probeGeneMapping

**Usage**

```
data(HDAC_genes)
```

**Format**

a 13x2 data.frame with gene identifiers in the first column and direction change in the second

**References**

Glaser et al. Gene expression profiling of multiple histone deacetylase (HDAC) inhibitors: defining a common gene set produced by HDAC inhibition in T24 and MDA carcinoma cell lines. Molecular cancer therapeutics, 2003.

---

intersectList	<i>Utility to find the intersection between a list of more than two vectors or lists</i>
---------------	--

---

**Description**

This function extends the native intersect function to work on two or more arguments.

**Usage**

```
intersectList(...)
```

**Arguments**

...            A list of or any number of vector like objects of the same mode, which could also be operated on by the native R set operations

**Value**

A vector like object of the same mode as the first argument, containing only the intersection common to all arguments to the function

**Examples**

```
list1 <- list('a', 'b', 'c')
list2 <- list('a', 'c')
list3 <- list('a', 'c', 'd')
listAll <- intersectList(list1, list2, list3)
listAll
```

---

intersectPSet	<i>Intersects objects of the PharmacoSet class, subsetting them to the common drugs and/or cell lines as selected by the user.</i>
---------------	--

---

**Description**

Given a list of PharmacoSets, the function will find the common drugs, and/or cell lines, and return PharmacoSets that contain data only pertaining to the common drugs, and/or cell lines. The mapping between dataset drug and cell names is done using annotations found in the PharmacoSet object's internal curation slot

**Usage**

```
intersectPSet(pSets, intersectOn = c("drugs", "cell.lines", "concentrations"),
  cells, drugs, strictIntersect = FALSE, verbose = TRUE, nthread = 1)
```

**Arguments**

pSets	[list] a list of PharmacoSet objects, of which the function should find the intersection
intersectOn	[character] which identifiers to intersect on, drugs or cell lines
cells	a vector of common cell lines between pSets. In case user is intersted on getting intersection on certain cell lines, they can provide their list of cell lines
drugs	a vector of common drugs between pSets. In case user is intersted on getting intersection on certain drugs, they can provide their list of drugs.
strictIntersect	[boolean] Should the intersection keep only the drugs and cell lines that have been tested on together?
verbose	[boolean] Should the function announce its key steps?
nthread	[numeric] The number of cores to use to run intersection on concentrations

**Value**

list a list of pSets, contatining only the intersection

**Examples**

```
data(GDSCsmall)
data(CCLEsmall)
common <- intersectPSet(list('GDSC'=GDSCsmall,
  'CCLE'=CCLEsmall), intersectOn = c("drugs", "cell.lines", "concentrations"))
common$CGP
common$CCLE
```

---

logLogisticRegression *Fits curves of the form  $E = E_{inf} + (1 - E_{inf})/(1 + (c/EC50)^{HS})$  to dose-response data points  $(c, E)$  given by the user and returns a vector containing estimates for  $HS$ ,  $E_{inf}$ , and  $EC50$ .*

---

**Description**

By default, logLogisticRegression uses an L-BFGS algorithm to generate the fit. However, if this fails to converge to solution, logLogisticRegression samples lattice points throughout the parameter space. It then uses the lattice point with minimal least-squares residual as an initial guess for the optimal parameters, passes this guess to drm, and re-attempts the optimization. If this still fails, logLogisticRegression uses the PatternSearch algorithm to fit a log-logistic curve to the data.

**Usage**

```
logLogisticRegression(conc, viability, density = c(2, 10, 2),
  step = 0.5/density, precision = 0.05, lower_bounds = c(0, 0, -6),
  upper_bounds = c(4, 1, 6), scale = 0.07, family = c("normal", "Cauchy"),
  median_n = 1, conc_as_log = FALSE, viability_as_pct = TRUE,
  trunc = TRUE, verbose = FALSE)
```

**Arguments**

<code>conc</code>	[vector] is a vector of drug concentrations.
<code>viability</code>	[vector] is a vector whose entries are the viability values observed in the presence of the drug concentrations whose logarithms are in the corresponding entries of the <code>log_conc</code> , where viability 0 indicates that all cells died, and viability 1 indicates that the drug had no effect on the cells.
<code>density</code>	[vector] is a vector of length 3 whose components are the numbers of lattice points per unit length along the HS-, $E_{inf}$ -, and base-10 logarithm of the EC50-dimensions of the parameter space, respectively.
<code>step</code>	[vector] is a vector of length 3 whose entries are the initial step sizes in the HS, $E_{inf}$ , and base-10 logarithm of the EC50 dimensions, respectively, for the PatternSearch algorithm.
<code>precision</code>	is a positive real number such that when the ratio of current step size to initial step size falls below it, the PatternSearch algorithm terminates. A smaller value will cause LogisticPatternSearch to take longer to complete optimization, but will produce a more accurate estimate for the fitted parameters.
<code>lower_bounds</code>	[vector] is a vector of length 3 whose entries are the lower bounds on the HS, $E_{inf}$ , and base-10 logarithm of the EC50 parameters, respectively.
<code>upper_bounds</code>	[vector] is a vector of length 3 whose entries are the upper bounds on the HS, $E_{inf}$ , and base-10 logarithm of the EC50 parameters, respectively.
<code>scale</code>	is a positive real number specifying the shape parameter of the Cauchy distribution.
<code>family</code>	[character], if "cauchy", uses MLE under an assumption of Cauchy-distributed errors instead of sum-of-squared-residuals as the objective function for assessing goodness-of-fit of dose-response curves to the data. Otherwise, if "normal", uses MLE with a gaussian assumption of errors
<code>median_n</code>	If the viability points being fit were medians of measurements, they are expected to follow a median of family distribution, which is in general quite different from the case of one measurement. <code>Median_n</code> is the number of measurements the median was taken of. If the measurements are means of values, then both the Normal and the Cauchy distributions are stable, so means of Cauchy or Normal distributed variables are still Cauchy and normal respectively.
<code>conc_as_log</code>	[logical], if true, assumes that log10-concentration data has been given rather than concentration data, and that log10(EC50) should be returned instead of EC50.
<code>viability_as_pct</code>	[logical], if false, assumes that viability is given as a decimal rather than a percentage, and that $E_{inf}$ should be returned as a decimal rather than a percentage.
<code>trunc</code>	[logical], if true, causes viability data to be truncated to lie between 0 and 1 before curve-fitting is performed.
<code>verbose</code>	[logical], if true, causes warnings thrown by the function to be printed.

**Value**

A vector containing estimates for HS,  $E_{inf}$ , and EC50

**Examples**

```
dose <- c("0.0025", "0.008", "0.025", "0.08", "0.25", "0.8", "2.53", "8")
viability <- c("108.67", "111", "102.16", "100.27", "90", "87", "74", "57")
computeAUC(dose, viability)
```

mcc

*Compute a Matthews Correlation Coefficient***Description**

The function computes a Matthews correlation coefficient for two factors provided to the function. It assumes each factor is a factor of class labels, and the entries are paired in order of the vectors.

**Usage**

```
mcc(x, y, nperm = 1000, setseed = 12345, nthread = 1)
```

**Arguments**

x, y	factor of the same length with the same number of levels
nperm	number of permutations for significance estimation. If 0, no permutation testing is done
setseed	seed for permutation testing
nthread	can parallelize permutation testing using parallel's mclapply

**Value**

A list with the MCC as the \$estimate, and p value as \$p.value

**Examples**

```
x <- factor(c(1,2,1,2,3,1))
y <- factor(c(2,1,1,1,2,2))
mcc(x,y)
```

mDataNames

*mDataNames***Description**

Returns the molecular data names for the PharmacoSet.

**Usage**

```
mDataNames(pSet)
```

**Arguments**

pSet                    PharamcoSet object

**Value**

Vector of names of the molecular data types

**Examples**

```
data(CCLEsmall)
mDataNames(CCLEsmall)
```

---

molecularProfiles        *molecularProfiles Generic*

---

**Description**

Generic for molecularProfiles method

**Usage**

```
molecularProfiles(pSet, mDataType)
```

**Arguments**

pSet                    The PharmacoSet to retrieve molecular profiles from  
mDataType                the type of molecular data

**Value**

a data.frame with the experiment info

**Examples**

```
data(CCLEsmall)
molecularProfiles(CCLEsmall, "rna")
```



---

molecularProfiles<-    *molecularProfiles<- Generic*

---

**Description**

Generic for molecularProfiles replace method

**Usage**

```
molecularProfiles(object, mDataType) <- value
```

**Arguments**

object	The PharmacoSet to replace molecular profiles in
mDataType	The type of molecular data to be updated
value	A matrix with the new profiles

**Value**

Updated PharmacoSet

**Examples**

```
data(CCLEsmall)
molecularProfiles(CCLEsmall, "rna") <- molecularProfiles(CCLEsmall, "rna")
```

---

pertNumber                    *pertNumber Generic*

---

**Description**

A generic for the pertNumber method

**Usage**

```
pertNumber(pSet)
```

**Arguments**

pSet	A PharmacoSet
------	---------------

**Value**

A 3D array with the number of perturbation experiments per drug and cell line, and data type

**Examples**

```
data(CCLEsmall)
pertNumber(CCLEsmall)
```

---

pertNumber<-                    *pertNumber<- Generic*

---

### Description

A generic for the pertNumber method

### Usage

```
pertNumber(object) <- value
```

### Arguments

object	A PharmacoSet
value	A new 3D array with the number of perturbation experiments per drug and cell line, and data type

### Value

The updated PharmacoSet

### Examples

```
data(CCLEsmall)
pertNumber(CCLEsmall) <- pertNumber(CCLEsmall)
```

---

PharmacoSet                    *PharmacoSet constructor*

---

### Description

A constructor that simplifies the process of creating PharmacoSets, as well as creates empty objects for data not provided to the constructor. Only objects returned by this constructor are expected to work with the PharmacoSet methods. For a much more detailed instruction on creating PharmacoSets, please see the "CreatingPharmacoSet" vignette.

### Usage

```
PharmacoSet(name, molecularProfiles = list(), cell = data.frame(),
  drug = data.frame(), sensitivityInfo = data.frame(),
  sensitivityRaw = array(dim = c(0, 0, 0)), sensitivityProfiles = matrix(),
  sensitivityN = matrix(nrow = 0, ncol = 0), perturbationN = array(NA, dim =
  c(0, 0, 0)), curationDrug = data.frame(), curationCell = data.frame(),
  curationTissue = data.frame(), datasetType = c("sensitivity",
  "perturbation", "both"), verify = TRUE)
```

**Arguments**

name	A character string detailing the name of the dataset
molecularProfiles	A list of ExpressionSet objects containing molecular profiles
cell	A data.frame containing the annotations for all the cell lines profiled in the data set, across all data types
drug	A data.frame containing the annotations for all the drugs profiled in the data set, across all data types
sensitivityInfo	A data.frame containing the information for the sensitivity experiments
sensitivityRaw	A 3 Dimensional array containing the raw drug dose response data for the sensitivity experiments
sensitivityProfiles	data.frame containing drug sensitivity profile statistics such as IC50 and AUC
sensitivityN, perturbationN	A data.frame summarizing the available sensitivity/perturbation data
curationDrug, curationCell, curationTissue	A data.frame mapping the names for drugs, cells and tissues used in the data set to universal identifiers used between different PharmacoSet objects
datasetType	A character string of 'sensitivity', 'preturbation', or both detailing what type of data can be found in the PharmacoSet, for proper processing of the data
verify	boolean Should the function verify the PharmacoSet and print out any errors it finds after construction?

**Value**

An object of class PharmacoSet

**Examples**

```
## For help creating a PharmacoSet object, please see the following vignette:
browseVignettes("PharmacoGx")
```

---

PharmacoSet-class	<i>A Class to Contain PharmacoGenomic datasets together with their curations</i>
-------------------	--

---

**Description**

The PharmacoSet (PSet) class was developed to contain and organise large PharmacoGenomic datasets, and aid in their metanalysis. It was designed primarily to allow bioinformaticians and biologists to work with data at the level of genes, drugs and cell lines, providing a more naturally intuitive interface and simplifying analyses between several datasets. As such, it was designed to be flexible enough to hold datasets of two different natures while providing a common interface. The class can accomodate datasets containing both drug dose response data, as well as datasets containing genetic profiles of cell lines pre and post treatment with compounds, known respectively as sensitivity and perturbation datasets.

**Usage**

```
## S4 method for signature 'PharmacoSet'
cellInfo(pSet)

## S4 replacement method for signature 'PharmacoSet,data.frame'
cellInfo(object) <- value

## S4 method for signature 'PharmacoSet'
drugInfo(pSet)

## S4 replacement method for signature 'PharmacoSet,data.frame'
drugInfo(object) <- value

## S4 method for signature 'PharmacoSet'
phenoInfo(pSet, mDataType)

## S4 replacement method for signature 'PharmacoSet,character,data.frame'
phenoInfo(object,
  mDataType) <- value

## S4 method for signature 'PharmacoSet'
molecularProfiles(pSet, mDataType)

## S4 replacement method for signature 'PharmacoSet,character,matrix'
molecularProfiles(object,
  mDataType) <- value

## S4 method for signature 'PharmacoSet'
featureInfo(pSet, mDataType)

## S4 replacement method for signature 'PharmacoSet,character,data.frame'
featureInfo(object,
  mDataType) <- value

## S4 method for signature 'PharmacoSet'
sensitivityInfo(pSet)

## S4 replacement method for signature 'PharmacoSet,data.frame'
sensitivityInfo(object) <- value

## S4 method for signature 'PharmacoSet'
sensitivityProfiles(pSet)

## S4 replacement method for signature 'PharmacoSet,data.frame'
sensitivityProfiles(object) <- value

## S4 replacement method for signature 'PharmacoSet,matrix'
sensitivityProfiles(object) <- value

## S4 method for signature 'PharmacoSet'
sensitivityMeasures(pSet)
```

```

## S4 method for signature 'PharmacoSet'
drugNames(pSet)

## S4 replacement method for signature 'PharmacoSet,character'
drugNames(object) <- value

## S4 method for signature 'PharmacoSet'
cellNames(pSet)

## S4 replacement method for signature 'PharmacoSet,character'
cellNames(object) <- value

## S4 method for signature 'PharmacoSet'
fNames(pSet, mDataType)

## S4 method for signature 'PharmacoSet'
dateCreated(pSet)

## S4 method for signature 'PharmacoSet'
pSetName(pSet)

## S4 method for signature 'PharmacoSet'
pertNumber(pSet)

## S4 method for signature 'PharmacoSet'
sensNumber(pSet)

## S4 replacement method for signature 'PharmacoSet,array'
pertNumber(object) <- value

## S4 replacement method for signature 'PharmacoSet,matrix'
sensNumber(object) <- value

```

**Arguments**

pSet	A PharmacoSet object
object	A PharmacoSet object
value	A replacement value
mDataType	A character with the type of molecular data to return/update

**Value**

An object of the PharmacoSet class

**Methods (by generic)**

- cellInfo: Returns the annotations for all the cell lines tested on in the PharmacoSet
- cellInfo<-: Update the cell line annotations
- drugInfo: Returns the annotations for all the drugs tested in the PharmacoSet
- drugInfo<-: Update the drug annotations
- phenoInfo: Return the experiment info from the given type of molecular data in PharmacoSet

- `phenoInfo<-`: Update the the given type of molecular data experiment info in the PharmacoSet
- `molecularProfiles`: Return the given type of molecular data from the PharmacoSet
- `molecularProfiles<-`: Update the given type of molecular data from the PharmacoSet
- `featureInfo`: Return the feature info for the given molecular data
- `featureInfo<-`: Replace the gene info for the molecular data
- `sensitivityInfo`: Return the drug dose sensitivity experiment info
- `sensitivityInfo<-`: Update the sensitivity experiment info
- `sensitivityProfiles`: Return the phenotypic data for the drug dose sensitivity
- `sensitivityProfiles<-`: Update the phenotypic data for the drug dose sensitivity
- `sensitivityProfiles<-`: Update the phenotypic data for the drug dose sensitivity
- `sensitivityMeasures`: Returns the available sensitivity profile summaries, for example, whether there are IC50 values available
- `drugNames`: Return the names of the drugs used in the PharmacoSet
- `drugNames<-`: Update the drug names used in the dataset
- `cellNames`: Return the cell names used in the dataset
- `cellNames<-`: Update the cell names used in the dataset
- `fNames`: Return the feature names used in the dataset
- `dateCreated`: Return the date the PharmacoSet was created
- `pSetName`: Return the name of the PharmacoSet
- `pertNumber`: Return the summary of available perturbation experiments
- `sensNumber`: Return the summary of available sensitivity experiments
- `pertNumber<-`: Update the summary of available perturbation experiments
- `sensNumber<-`: Update the summary of available sensitivity experiments

## Slots

- `annotation` A list of annotation data about the PharmacoSet, including the `$name` and the session information for how the object was creating, detailing the exact versions of R and all the packages used
- `molecularProfiles` A list containing 4 `Biobase::ExpressionSet` type object for holding data for RNA, DNA, SNP and Copy Number Variation measurements respectively, with associated `fData` and `pData` containing the row and column metadata
- `cell` A `data.frame` containing the annotations for all the cell lines profiled in the data set, across all data types
- `drug` A `data.frame` containing the annotations for all the drugs profiled in the data set, across all data types
- `sensitivity` A list containing all the data for the sensitivity experiments, including `$info`, a `data.frame` containing the experimental info, `$raw` a 3D array containing raw data, `$profiles`, a `data.frame` containing sensitivity profiles statistics, and `$n`, a `data.frame` detailing the number of experiments for each cell-drug pair
- `perturbation` A list containing `$n`, a `data.frame` summarizing the available perturbation data,
- `curation` A list containing mappings for `$drug`, `cell`, `tissue` names used in the data set to universal identifiers used between different PharmacoSet objects
- `datasetType` A character string of 'sensitivity', 'perturbation', or both detailing what type of data can be found in the PharmacoSet, for proper processing of the data

---

phenoInfo	<i>phenoInfo Generic</i>
-----------	--------------------------

---

**Description**

Generic for phenoInfo method

**Usage**

```
phenoInfo(pSet, mDataType)
```

**Arguments**

pSet	The PharmacoSet to retrieve rna annotations from
mDataType	the type of molecular data

**Value**

a data.frame with the experiment info

**Examples**

```
data(CCLEsmall)
phenoInfo(CCLEsmall, mDataType="rna")
```

---

phenoInfo<-	<i>phenoInfo&lt;- Generic</i>
-------------	-------------------------------

---

**Description**

Generic for phenoInfo replace method

**Usage**

```
phenoInfo(object, mDataType) <- value
```

**Arguments**

object	The PharmacoSet to retrieve molecular experiment annotations from
mDataType	the type of molecular data
value	a data.frame with the new experiment annotations

**Value**

The updated PharmacoSet

**Examples**

```
data(CCLEsmall)
phenoInfo(CCLEsmall, mDataType="rna") <- phenoInfo(CCLEsmall, mDataType="rna")
```

---

pSetName	<i>pSetName Generic</i>
----------	-------------------------

---

**Description**

A generic for the pSetName method

**Usage**

```
pSetName(pSet)
```

**Arguments**

pSet            A PharmacoSet

**Value**

The name of the PharmacoSet

**Examples**

```
data(CCLEsmall)
pSetName(CCLEsmall)
```

---

sensitivityInfo	<i>sensitivityInfo Generic</i>
-----------------	--------------------------------

---

**Description**

Generic for sensitivityInfo method

**Usage**

```
sensitivityInfo(pSet)
```

**Arguments**

pSet            The PharmacoSet to retrieve sensitivity experiment annotations from

**Value**

a data.frame with the experiment info



### Examples

```
data(CCLEsmall)
sensitivityInfo(CCLEsmall)
```

---

sensitivityInfo<-      *sensitivityInfo<- Generic*

---

### Description

A generic for the sensitivityInfo replacement method

### Usage

```
sensitivityInfo(object) <- value
```

### Arguments

object	The PharmacoSet to update
value	A data.frame with the new sensitivity annotations

### Value

Updated PharmacoSet

### Examples

```
data(CCLEsmall)
sensitivityInfo(CCLEsmall) <- sensitivityInfo(CCLEsmall)
```

---

sensitivityMeasures      *sensitivityMeasures Generic*

---

### Description

A generic for the sensitivityMeasures method

### Usage

```
sensitivityMeasures(pSet)
```

### Arguments

pSet	The PharmacoSet
------	-----------------

### Value

A character vector of all the available sensitivity measures

**Examples**

```
data(CCLEsmall)
sensitivityMeasures(CCLEsmall)
```

---

```
sensitivityProfiles    sensitivityProfiles Generic
```

---

**Description**

Generic for sensitivityProfiles method

**Usage**

```
sensitivityProfiles(pSet)
```

**Arguments**

pSet                    The PharmacoSet to retrieve sensitivity experiment data from

**Value**

a data.frame with the experiment info

**Examples**

```
data(CCLEsmall)
sensitivityProfiles(CCLEsmall)
```

---

```
sensitivityProfiles<-    sensitivityProfiles<- Generic
```

---

**Description**

A generic for the sensitivityProfiles replacement method

**Usage**

```
sensitivityProfiles(object) <- value
```

**Arguments**

object                    The PharmacoSet to update  
value                      A data.frame with the new sensitivity profiles. If a matrix object is passed in, converted to data.frame before assignment

**Value**

Updated PharmacoSet

**Examples**

```
data(CCLEsmall)
sensitivityProfiles(CCLEsmall) <- sensitivityProfiles(CCLEsmall)
```

---

sensNumber	<i>sensNumber Generic</i>
------------	---------------------------

---

**Description**

A generic for the sensNumber method

**Usage**

```
sensNumber(pSet)
```

**Arguments**

pSet            A PharmacoS

**Value**

A data.frame with the number of sensitivity experiments per drug and cell line

**Examples**

```
data(CCLEsmall)
sensNumber(CCLEsmall)
```

---

sensNumber<-	<i>sensNumber&lt;- Generic</i>
--------------	--------------------------------

---

**Description**

A generic for the sensNumber method

**Usage**

```
sensNumber(object) <- value
```

**Arguments**

object            A PharmacoS  
value            A new data.frame with the number of sensitivity experiments per drug and cell line

**Value**

The updated PharmacoS

**Examples**

```
data(CCLEsmall)
sensNumber(CCLEsmall) <- sensNumber(CCLEsmall)
```

---

show,PharmacoSet-method

*Show a PharamcoSet*

---

**Description**

Show a PharamcoSet

**Usage**

```
## S4 method for signature 'PharmacoSet'
show(object)
```

**Arguments**

object            PharmacoSet

**Value**

Prints the PharmacoSet object to the output stream, and returns invisible NULL.

**Examples**

```
data(CCLEsmall)
CCLEsmall
```

---

show,PharmacoSig-method

*Show PharmacoGx Signatures*

---

**Description**

Show PharmacoGx Signatures

**Usage**

```
## S4 method for signature 'PharmacoSig'
show(object)
```

**Arguments**

object            PharmacoSig

**Value**

Prints the PharmacGx Signatures object to the output stream, and returns invisible NULL.

**Examples**

```
data(GDSCsmall)
drug.sensitivity <- drugSensitivitySig(GDSCsmall, mDataType="rna",
                                     nthread=1, features = fName(GDSCsmall, "rna")[1])
drug.sensitivity
```

---

showSigAnnot

*Show the Annotations of a signature object*

---

**Description**

This function prints out the information about the call used to compute the drug signatures, and the session info for the session in which the computation was done. Useful for determining the exact conditions used to generate signatures.

**Usage**

```
showSigAnnot(Sigs)
```

**Arguments**

**Sigs** An object of the PharmacSig Class, as returned by drugPerturbationSig or drugSensitivitySig

**Value**

Prints the PharmacGx Signatures annotations to the output stream, and returns invisible NULL.

**Examples**

```
data(GDSCsmall)
drug.sensitivity <- drugSensitivitySig(GDSCsmall, mDataType="rna",
                                     nthread=1, features = fName(GDSCsmall, "rna")[1])
showSigAnnot(drug.sensitivity)
```

---

subsetTo	<i>A function to subset a PharmacoSet to data containing only specified drugs, cells and genes</i>
----------	--

---

## Description

This is the preferred method of subsetting a PharmacoSet. This function allows abstraction of the data to the level of biologically relevant objects: drugs and cells. The function will automatically go through all of the combined data in the PharmacoSet and ensure only the requested drugs and cell lines are found in any of the slots. This allows quickly picking out all the experiments for a drug or cell of interest, as well removes the need to keep track of all the metadata conventions between different datasets.

## Usage

```
subsetTo(pSet, cells = NULL, drugs = NULL, molecular.data.cells = NULL,  
keep.controls = TRUE, ...)
```

## Arguments

pSet	A PharmacoSet to be subsetted
cells	A list or vector of cell names as used in the dataset to which the object will be subsetted. If left blank, then all cells will be left in the dataset.
drugs	A list or vector of drug names as used in the dataset to which the object will be subsetted. If left blank, then all drugs will be left in the dataset.
molecular.data.cells	A list or vector of cell names to keep in the molecular data
keep.controls	If the dataset has perturbation type experiments, should the controls be kept in the dataset? Defaults to true.
...	Other arguments passed by other function within the package

## Value

A PharmacoSet with only the selected drugs and cells

## Examples

```
data(CCLEsmall)  
CCLDrugs <- drugNames(CCLEsmall)  
CCLCells <- cellNames(CCLEsmall)  
PSet <- subsetTo(CCLEsmall, drugs = CCLDrugs[1], cells = CCLCells[1])  
PSet
```

---

**summarizeMolecularProfiles**

*Takes molecular data from a PharmacoSet, and summarises them into one entry per drug*

---

**Description**

Given a PharmacoSet with molecular data, this function will summarize the data into one profile per cell line, using the chosen summary.stat. Note that this does not really make sense with perturbation type data, and will combine experiments and controls when doing the summary if run on a perturbation dataset.

**Usage**

```
summarizeMolecularProfiles(pSet, mDataType, cell.lines, features,
  summary.stat = c("mean", "median", "first", "last", "and", "or"),
  fill.missing = TRUE, summarize = TRUE, verbose = TRUE,
  binarize.threshold = NA, binarize.direction = c("less", "greater"))
```

**Arguments**

pSet	PharmacoSet The PharmacoSet to summarize
mDataType	character which one of the molecular data types to use in the analysis, out of all the molecular data types available for the pset for example: rna, rnaseq, snp
cell.lines	character The cell lines to be summarized. If any cell.line has no data, missing values will be created
features	character A vector of the feature names to include in the summary
summary.stat	character which summary method to use if there are repeated cell.lines? Choices are "mean", "median", "first", or "last" In case molecular data type is mutation or fusion "and" and "or" choices are available
fill.missing	boolean should the missing cell lines not in the molecular data object be filled in with missing values?
summarize	A flag which when set to FALSE (defaults to TRUE) disables summarizing and returns the data unchanged as a ExpressionSet
verbose	boolean should messages be printed
binarize.threshold	numeric A value on which the molecular data is binarized. If NA, no binarization is done.
binarize.direction	character One of "less" or "greater", the direction of binarization on binarize.threshold, if it is not NA.

**Value**

matrix An updated PharmacoSet with the molecular data summarized per cell line.

**Examples**

```
data(GDSCsmall)
GDSCsmall <- summarizeMolecularProfiles(GDSCsmall,
                                       mDataType = "rna", cell.lines=cellNames(GDSCsmall),
                                       summary.stat = 'median', fill.missing = TRUE, verbose=TRUE)
GDSCsmall
```

---

**summarizeSensitivityProfiles**

*Takes the sensitivity data from a PharmacoSet, and summarises them into a drug vs cell line table*

---

**Description**

This function creates a table with cell lines as rows and drugs as columns, summarising the drug sensitivity data of a PharmacoSet into drug-cell line pairs

**Usage**

```
summarizeSensitivityProfiles(pSet, sensitivity.measure = "auc_recomputed",
                             cell.lines, drugs, summary.stat = c("mean", "median", "first", "last",
                             "max", "min"), fill.missing = TRUE, verbose = TRUE)
```

**Arguments**

pSet	[PharmacoSet] The PharmacoSet from which to extract the data
sensitivity.measure	[character] which sensitivity measure to use? Use the sensitivityMeasures function to find out what measures are available for each PSet.
cell.lines	character The cell lines to be summarized. If any cell lines has no data, it will be filled with missing values
drugs	character The drugs to be summarized. If any drugs has no data, it will be filled with missing values
summary.stat	character which summary method to use if there are repeated cell line-drug experiments? Choices are "mean", "median", "first", or "last"
fill.missing	boolean should the missing cell lines not in the molecular data object be filled in with missing values?
verbose	Should the function print progress messages?

**Value**

matrix A matrix with cell lines going down the rows, drugs across the columns, with the selected sensitivity statistic for each pair.

**Examples**

```
data(GDSCsmall)
GDSCauc <- summarizeSensitivityProfiles(GDSCsmall, sensitivity.measure='auc_published')
```



---

symSetDiffList	<i>Utility to find the symmetric set difference of a list of two or more vectors or lists</i>
----------------	---

---

**Description**

The function finds the symmetric set differences between all the arguments, defined as  $\text{Union}(\text{args}) - \text{Intersection}(\text{args})$

**Usage**

```
symSetDiffList(...)
```

**Arguments**

... A list of or any number of vector like objects of the same mode, which could also be operated on by the native R set operations

**Value**

A vector like object of the same mode as the first argument, containing only the symmetric set difference

**Examples**

```
list1 <- list('a', 'b', 'c')
list2 <- list('a', 'c')
list3 <- list('a', 'c', 'd')
listAll <- symSetDiffList(list1, list2, list3)
listAll
```

---

unionList	<i>Utility to find the union between a list of more than two vectors or lists</i>
-----------	---

---

**Description**

This function extends the native union function to work on two or more arguments.

**Usage**

```
unionList(...)
```

**Arguments**

... A list of or any number of vector like objects of the same mode, which could also be operated on by the native R set operations

**Value**

A vector like object of the same mode as the first argument, containing all the elements of all arguments passed to the function

**Examples**

```
list1 <- list('a', 'b')
list2 <- list('a', 'c')
list3 <- list('c', 'd')
listAll <- unionList(list1, list2, list3)
listAll
```

---

```
[,PharmacoSet,ANY,ANY,ANY-method
  'I'
```

---

**Description**

'['

**Usage**

```
## S4 method for signature 'PharmacoSet,ANY,ANY,ANY'
x[i, j, ..., drop = FALSE]
```

**Arguments**

x	PSet
i	Cell lines to keep in PSet
j	Drugs to keep in PSet
...	further arguments
drop	A boolean flag of whether to drop single dimensions or not

**Value**

Returns the subsetting PSet

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