Package 'biotmle'

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Title Targeted Learning for Biomarker Discovery with Moderated Statistics

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Description This package facilitates the discovery of biomarkers from biological sequencing data (e.g., microarrays, RNA-seq) based on the associations of potential biomarkers with exposure and outcome variables by implementing an estimation procedure that combines a generalization of moderated statistics with asymptotically linear statistical parameters estimated via targeted minimum loss-based estimation (TMLE).

Depends R (>= 3.4)

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URL https://github.com/nhejazi/biotmle

BugReports https://github.com/nhejazi/biotmle/issues

Encoding UTF-8

LazyData true

Imports dplyr, magrittr, ggplot2, superheat, wesanderson, doFuture, future, stats, Matrix, methods, DBI, limma, BiocParallel, SummarizedExperiment, biotmleData (>= 1.1.1), SuperLearner, tmle

Suggests testthat, knitr, rmarkdown, BiocStyle

VignetteBuilder knitr

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`biomarkertmle` *Biomarker Evaluation with Targeted Minimum Loss-Based Estimation (TMLE)*

Description

Computes the causal target parameter defined as the difference between the biomarker expression values under treatment and those same values under no treatment, using Targeted Minimum Loss-Based Estimation.

Usage

```
biomarkertmle(se, varInt, ngscounts = FALSE, parallel = TRUE,
  bppar_type = NULL, future_param = NULL, family = "gaussian",
  subj_ids = NULL, g_lib = c("SL.glm", "SL.randomForest", "SL.nnet",
  "SL.polymars", "SL.mean"), Q_lib = c("SL.glm", "SL.randomForest", "SL.nnet",
  "SL.mean"))
```

Arguments

`se`	(SummarizedExperiment) - containing expression or next-generation sequencing data in the "assays" slot and a matrix of phenotype-level data in the "colData" slot.
`varInt`	(numeric) - indicating the column of the design matrix corresponding to the treatment or outcome of interest (in the "colData" slot of the "se" argument above).
`ngscounts`	(logical) - whether the data are counts generated from a next-generation sequencing (NGS) experiment (e.g., RNA-seq). The default setting assumes continuous expression measures as generated by microarray-type platforms.
`parallel`	(logical) - whether or not to use parallelization in the estimation procedure. Invoking parallelization happens through a combination of calls to `future` and `BiocParallel`. If this argument is set to `TRUE`, `future::multiprocess` is used, and if `FALSE`, `future::sequential` is used, alongside `BiocParallel::bplapply`. Other options for evaluation through futures may be invoked by setting the argument `future_param`.
`bppar_type`	(character) - specifies the type of backend to be used with the parallelization invoked by `BiocParallel`. Consult the manual page for `BiocParallel::BiocParallelParam` for possible types and descriptions on their appropriate uses. The default for this argument is `NULL`, which silently uses `BiocParallel::DoparParam`.
`future_param`	(character) - specifies the type of parallelization to be invoked when using futures for evaluation. For a list of the available types, please consult the documentation for `future::plan`. The default setting (this argument set to `NULL`) silently invokes `future::multiprocess`. Be careful if changing this setting.

family	(character) - specification of error family: "binomial" or "gaussian".
subj_ids	(numeric vector) - subject IDs to be passed directly to the same subject should have the exact same numerical identifier; coerced to numeric if not provided in the appropriate form.
g_lib	(char vector) - library of learning algorithms to be used in fitting the "g" step of the standard TMLE procedure.
Q_lib	(char vector) - library of learning algorithms to be used in fitting the "Q" step of the standard TMLE procedure.

Value

S4 object of class `biotmle`, generated by sub-classing `SummarizedExperiment`, with additional slots containing `tmleOut` and `call`, among others, containing TMLE-based estimates of the relationship between a biomarker and exposure or outcome variable and the original call to this function (for user reference), respectively.

Examples

```
library(dplyr)
library(biotmleData)
data(illuminaData)
library(SummarizedExperiment)
"%ni%" = Negate("%in%")

colData(illuminaData) <- colData(illuminaData) %>%
     data.frame %>%
     dplyr::mutate(age = as.numeric(age > median(age))) %>%
     DataFrame

varInt_index <- which(names(colData(illuminaData)) %in% "benzene")

biomarkerTMLEout <- biomarkertmle(se = illuminaData[1:2, ],
                                  varInt = varInt_index,
                                  parallel = FALSE,
                                  family = "gaussian",
                                  g_lib = c("SL.mean", "SL.glm"),
                                  Q_lib = "SL.mean"
                                 )
```

`biomarkerTMLE_exposure`

TMLE procedure for Biomarker Identication from Exposure

Description

This function performs influence curve-based estimation of the effect of an exposure on biological expression values associated with a given biomarker, controlling for a user-specified set of baseline covariates

Usage

```
biomarkerTMLE_exposure(Y, W, A, a, subj_ids = NULL, family = "gaussian",
  g_lib, Q_lib)
```

Arguments

Y	(numeric vector) - a vector of expression values for a single biomarker.
W	(numeric matrix) - a matrix of baseline covariates to be controlled in the estimation process.
A	(numeric vector) - a discretized exposure vector (e.g., from a design matrix whose effect on expression values is of interest.
a	(numeric vector) - the levels of A against which comparisons are to be made.
subj_ids	(numeric vector) - subject IDs to be passed directly to the same subject should have the exact same numerical identifier. coerced to numeric if not provided in the appropriate form.
family	(character) - specification of error family: "binomial" or "gaussian"
g_lib	(char vector) - library of learning algorithms to be used in fitting the "g" step of the standard TMLE procedure.
Q_lib	(char vector) - library of learning algorithms to be used in fitting the "Q" step of the standard TMLE procedure.

Value

TMLE-based estimate of the relationship between biomarker expression and changes in an exposure variable, computed iteratively and saved in the `tmleOut` slot in a `biotmle` object.

---

bioTMLE-class *Constructor for class bioTMLE*

Description

Constructor for class bioTMLE

Value

class `biotmle` object, sub-classed from SummarizedExperiment.

Examples

```
library(SummarizedExperiment)
library(biotmleData)
data(illuminaData)

example_biotmle_class <- function(se) {

    call <- match.call(expand.dots = TRUE)
    biotmle <- .biotmle(
          SummarizedExperiment(
             assays = assay(se),
             rowData = rowData(se),
```

```
            colData = colData(se)
          ),
          call = call,
          tmleOut = as.data.frame(matrix(NA, 10, 10)),
          topTable = as.data.frame(matrix(NA, 10, 10))
    )
    return(biotmle)
}

example_class <- example_biotmle_class(se = illuminaData)
```

---

data.frame_OR_EList-class *S4 class union data.frame_OR_EList*

Description

Class union contaning `data.frame` and `limma::Elist`, used internally to handle situations when a returned object has a type that cannot be guessed from the function call.

Value

fusion of classes `data.frame` and `EList`, used within `.biotmle` by class `bioTMLE` to handle uncertainty in the object passed to slot "tmleOut".

---

heatmap_ic *Heatmap for class biotmle*

Description

Heatmap of the contributions of a select subset of biomarkers to the variable importance measure changes as assessed by influence curve-based estimation, across all subjects.

Usage

```
heatmap_ic(x, ..., design, FDRcutoff = 0.05, top = 25)
```

Arguments

`x`	object of class `biotmle` as produced by an appropriate call to `biomarkertmle`
`...`	additional arguments passed to `superheat::superheat` as necessary
`design`	a vector providing the contrast to be displayed in the heatmap.
`FDRcutoff`	cutoff to be used in controlling the False Discovery Rate
`top`	number of identified biomarkers to plot in the heatmap

Value

heatmap (from the superheat package) using hierarchical clustering to plot the changes in the variable importance measure for all subjects across a specified top number of biomarkers.

Examples

```
library(dplyr)
library(biotmleData)
library(SummarizedExperiment)
data(illuminaData)
data(biomarkertmleOut)

colData(illuminaData) <- colData(illuminaData) %>%
     data.frame %>%
     dplyr::mutate(age = as.numeric(age > median(age))) %>%
     DataFrame

varInt_index <- which(names(colData(illuminaData)) %in% "benzene")
designVar <- as.data.frame(colData(illuminaData))[, varInt_index]
design <- as.numeric(designVar == max(designVar))

limmaTMLEout <- modtest_ic(biotmle = biomarkerTMLEout)

heatmap_ic(x = limmaTMLEout, design = design, FDRcutoff = 0.05, top = 15)
```

`modtest_ic` *Moderated Statistical Tests for Influence Curves*

Description

Performs variance shrinkage via the empirical Bayes procedure of LIMMA on the observed data after a transformation moving the data to influence curve space, based on the average treatment effect parameter.

Usage

```
modtest_ic(biotmle, adjust = "BH")
```

Arguments

`biotmle`	`biotmle` object as generated by `biomarkertmle`
`adjust`	the multiple testing correction to be applied to p-values that are generated from the moderated tests. The recommended (and default) method is that of Benjamini and Hochberg. See topTable for a list of appropriate methods.

Value

`biotmle` object containing output from `limma::lmFit` and `limma::topTable`

Examples

```
library(biotmleData)
library(SummarizedExperiment)
data(biomarkertmleOut)

limmaTMLEout <- modtest_ic(biotmle = biomarkerTMLEout)
```

plot.bioTMLE *Plot p-values from moderated statistical tests for class biotmle*

Description

Histogram of raw or FDR-adjusted p-values from the moderated t-test.

Usage

```
## S3 method for class 'bioTMLE'
plot(x, ..., type = "pvals_adj")
```

Arguments

x	object of class biotmle as produced by an appropriate call to biomarkertmle
...	additional arguments passed plot as necessary
type	character describing whether to provide a plot of unadjusted or adjusted p-values (adjustment performed via Benjamini-Hochberg)

Value

object of class ggplot containing a histogram of the raw or Benjamini-Hochberg corrected p-values (depending on user input).

Examples

```
library(dplyr)
library(biotmleData)
library(SummarizedExperiment)
data(biomarkertmleOut)

limmaTMLEout <- modtest_ic(biotmle = biomarkerTMLEout)

plot(x = limmaTMLEout, type = "pvals_adj")
```

rnaseq_ic *Transformation utility for using "voom" with biomarker TMLE procedure*

Description

This function prepares next-generation sequencing data (counts) for use with the biomarker TMLE procedure by invoking the voom transform of limma.

Usage

```
rnaseq_ic(biotmle, weights = TRUE, ...)
```

Arguments

biotmle	(bioTMLE) - subclass of SummarizedExperiment containing next-generation sequencing (NGS) count data in the "assays" slot.
weights	(logical) - whether to return quality weights of samples in the output object.
...	- other arguments to be passed to functions limma::voom or limma::voomWithQualityWeights as appropriate.

Value

EList object containing voom-transformed "expression" measures of count data (actually, the mean-variance trend) in the "E" slot, to be passed into the biomarker TMLE procedure.

---

volcano_ic *Volcano plot for class biotmle*

Description

Volcano plot of the log-changes in the target causal paramter against the log raw p-values from the moderated t-test.

Usage

```
volcano_ic(biotmle, fc_bound = 3, pval_bound = 0.2)
```

Arguments

biotmle	object of class biotmle as produced by an appropriate call to biomarkertmle
fc_bound	(numeric) - indicates the highest magnitude of the fold to be colored along the x-axis of the volcano plot; this limits the observations to be considered differentially expressed to those in a user-specified interval.
pval_bound	(numeric) - indicates the largest corrected p-value to be colored along the y-axis of the volcano plot; this limits observations considered as differentially expressed to those in a user-specified interval.

Value

object of class ggplot containing a standard volcano plot of the log-fold change in the causal target parameter against the raw log p-value computed from the moderated tests in modtest_ic.

Examples

```
library(dplyr)
library(biotmleData)
library(SummarizedExperiment)
data(biomarkertmleOut)

limmaTMLEout <- modtest_ic(biotmle = biomarkerTMLEout)

volcano_ic(biotmle = limmaTMLEout)
```

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