Package 'MAST'

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Description Methods and models for handling zero-inflated single cell assay data.

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Collate 'AllGenerics.R' 'AllClasses.R' 'CovFromBoots.R' 'Fluidigm-methods.R' 'GSEA-by-boot.R' 'Hypothesis.R' 'LmWrapper.R' 'MAST-package.R' 'MultidimensionalScaling.R' 'RNASeqAssay-methods.R' 'Readers.R' 'SingleCellAssay-methods.R' 'UtilityFunctions.R' 'ZlmFit-bootstrap.R' 'ZlmFit-logFC.R' 'ZlmFit.R' 'bayesglm.R' 'convertMASTClassic.R' 'ebayes-helpers.R' 'filterEval.R' 'helper-methods.R' 'lmWrapper-bayesglm.R' 'lmWrapper-glm.R' 'lmWrapper-glmer.R' 'lmWrapper-ridge.R' 'lrtest.R' 'predict.R' 'stat_ell.R' 'thresholdSCRNA.R' 'zeroinf.R' 'zlmHooks.R'

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Contents

`MAST-package` *MAST: Model-based Analysis of Single- cell Transcriptomics*

Description

Methods for analysing single cell assay data using hurdle models.

Details

This packages provides data structures and functions for statistical analysis of single-cell assay data such as Fluidigm single cell gene expression assays.

Author(s)

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- Masanao Yajima <myajima@fredhutch.org>

References

Finak, et al. MAST: a flexible statistical framework for assessing transcriptional changes and characterizing heterogeneity in single-cell RNA sequencing data. Genome Biology (2015).

See Also

Useful links:

- https://github.com/RGLab/MAST/
- Report bugs at https://github.com/RGLab/MAST/issues

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applyFlat *Apply a vectorized binary operation recycling over last dimension*

Description

When x is an array of order K, and y is an array of order K-1, whose dimensions otherwise agree, apply FUN by recycling y as necessary over dimension K of x.

Usage

```
applyFlat(x, y, FUN = "-")
```

Arguments

x	array, order K
y	array, order K-1
FUN	vectorized binary operation

Value

array, order K equal to FUN(x,y)

Examples

```
##Dumb example, could be done with scale(...,scale=FALSE)
x0 = matrix(1:10, nrow=2)
y0 = rowMeans(x0)
dim(y0) = c(1, 2)
x1 = MAST:::applyFlat(x0,y0)
stopifnot(rowMeans(x1)==0)
```

BayesGLMlike-class *Wrapper for bayesian GLM*

Description

Wrapper for bayesian GLM

Slots

`prior numeric` optional 3d array used to specify prior for coefficients

`useContinuousBayes logical` should `bayesglm` be used to fit the continuous component as well?

---

calcZ *Get Z or T statistics and P values after running gseaAfterBoot*

Description

The Z or T statistics may be reported by component (discrete/continuous) when `combined='no'` or combined by Fisher's or Stouffer's method (`combined='fisher'` or `combined='stouffer'`. Fisher's method uses the product of the p-values, while Stouffer's method uses the sum of the Z/T scores. The "Z" score returned by Fisher is the normal quantile that would yield the observed Fisher P-value, whose sign is derived from the sign of the maximum component Z score. The "Z" score returned by Stouffer when `testType='normal'` is the sum of the Z scores, over sqrt(2). When `testType='t'` it is a weighted combination of the Z scores, with weights correponding to the degrees of freedom in each of the t statistics. A t-approximation to this sum of t-variables is derived by matching moments. It seems to be fairly accurate in practice.

Usage

```
calcZ(gseaObj, testType = "t", combined = "none")
```

Arguments

`gseaObj`	output from `gseaAfterBoot`
`testType`	either 'normal' or 't'. The 't' test adjusts for excess kurtosis due to the finite number of bootstrap replicates used to estimate the variance of the statistics. This will result in more conservative inference.
`combined`	`character` one of 'none', 'fisher' or 'stouffer'

Value

3D array with dimensions set (modules) comp ('cont'inuous or 'disc'rete) and metric ('Z' stat and two sided 'P' value that P(z>|Z|)) if `combined='no'`, otherwise just a matrix.

See Also

gseaAfterBoot

Examples

```
## See the examples in gseaAfterBoot
example(gseaAfterBoot)
```

`colData<-,SingleCellAssay,DataFrame-method` *Replace* `colData`

Description

Replace `colData` with a `DataFrame`. Checks to make sure that `row.names(value)` match `colnames{x}`, in contrast to the parent method Checks for a wellKey column, as well.

Usage

```
## S4 replacement method for signature 'SingleCellAssay,DataFrame'
colData(x) <- value
```

Arguments

x	SingleCellAssay
value	DataFrame

Value

modified `SingleCellAssay`

`collectResiduals` *Residual hooks and collection methods*

Description

After each gene is fit, a hook function can optionally be run and the output saved. This allows extended computations to be done using the fitted model, without keeping it in memory. Here this is used to calculate various residuals, though in some cases they can be done using only the information contained in the `ZlmFit`-class.

Usage

```
collectResiduals(x, sca, newLayerName = "Residuals")

discrete_residuals_hook(x)

continuous_residuals_hook(x)

combined_residuals_hook(x)

deviance_residuals_hook(x)

fitted_phat(x)

partialScore(x, effectRegex)
```

Arguments

x	ZlmFit-class
sca	SingleCellAssay object to which the residuals should be added
newLayerName	character name of the assay layer
effectRegex	a regular expression naming columns of the design corresponding to Z_0. Generally these should be the treatment effects of interest.

Value

copy of sca with new layer

Functions

- discrete_residuals_hook(): Hook to get the discrete residuals, ie, difference between expected probability of expression and observed
- continuous_residuals_hook(): Hook to get the continuous residuals, ie, residuals for conditionally positive observations. If an observation is zero, it's residual is defined to be zero as well.
- combined_residuals_hook(): Hook to get the combined residuals, ie, Y-E(U)*E(V)
- deviance_residuals_hook(): Standardized deviance residuals hook. Computes the sum of the standardized deviance residuals for the discrete and continuous models (scaled to have unit variance). If the observation is zero then only the discrete component is used.
- fitted_phat(): Hook to return p_hat, the predicted probability of expression.
- partialScore(): Compute $Y_i - E(V_i|X_i, Z_0)E(U|X_i, Z_0)$, where Z_0 is a treatment effect (being left in) and X_i is a nuisance effect (being regressed out).

Total residual types

Each component of the model contributes several flavors of residual, which can be combined in various fashions. The discrete residual can be on the response scale (thus subtracting the predicted probability of expression from the 0/1 expression value). Or it can be a deviance residual, revealing something about the log-likelihood.

Partial residuals

It's also possible to consider partial residuals, in which the contribution of a particular covariate is added back into the model.

See Also

zlm

Examples

```
data(vbetaFA)
svbeta <- subset(vbetaFA, ncells==1)
svbeta <- svbeta[freq(svbeta)>.4,]
window <- function(x1) lapply(assays(x1), function(x2) x2[1:3, 1:6])
#total residuals of the response
z1 <- zlm(~ Stim.Condition, svbeta, hook=discrete_residuals_hook)
window(collectResiduals(z1, svbeta))
z2 <- zlm(~ Stim.Condition, svbeta, hook=continuous_residuals_hook)
window(collectResiduals(z2, svbeta))
z3 <- zlm(~ Stim.Condition, svbeta, hook=combined_residuals_hook)
window(collectResiduals(z3, svbeta))
#partial residuals
colData(svbeta)$ngeneson <- colMeans(assay(svbeta)>0)
z5 <- zlm(~ Stim.Condition + ngeneson, svbeta)
partialScore(z5, 'Stim.Condition')
```

---

computeEtFromCt *Compute the Et from the Ct*

Description

Computes the Et value from the Ct value in an existing data frame and returns a new data frame with the Et column appended

Usage

```
computeEtFromCt(df, column = "Ct", Cmax = 40)
```

Arguments

df	a data.frame
column	The name of the Ct column. A character. 'Ct' by default.
Cmax	the maximum number of cycles performed. 40 by default.

Value

A copy of df with the 'Et' column appended

Author(s)

Greg Finak

Examples

```
data(vbeta)
vbeta <- computeEtFromCt(vbeta)
```

`convertMASTClassicToSingleCellAssay` *Convert a MASTClassic SingleCellAssay*

Description

Convert a SingleCellAssay object created with the MASTClassic package to an object recognized by the new MAST package

Usage

```
convertMASTClassicToSingleCellAssay(object = NULL)
```

Arguments

`object`	of class `SingleCellAssay` created by MASTClassic

Details

The function will extract the relevant information from the attributes of the old object and construct a new SingleCellAssay that is recognized by MAST. This function checks that the object is a MASTClassic SingleCellAssay object. It will stop if it is not a SingleCellAssay, return a converted SingleCellAssay if object was created by MASTClassic, and return the original object if the object is already compatible.

Value

A MAST SingleCellAssay object.

Note

Type checking for old object is not performed.

Examples

```
data(vbetaFA)
convertMASTClassicToSingleCellAssay(vbetaFA)
```

CovFromBoots — *Extract the inter-gene covariance matrices for continuous and discrete components of a MAST model for a given coefficient from bootstrap replicates*

Description

Computes the genewise covariance for a model coefficient from bootstrap replicates from ‘MAST::bootVcov1()‘. If coefficients are unestimable (i.e. NA) for a gene, that row/column in the covariance matrix will be NA. Returns a list with components "C" and "D" containing the covariance matrices for the "C"ontinuous and "D"iscrete components of the MAST model.

Usage

```
CovFromBoots(boots = NULL, coefficient = NULL)
```

Arguments

boots	a multidimensional array returned by ‘bootVcov1‘ or ‘pbootVcov1‘.
coefficient	‘character‘ the name of the model coefficient for which to return the inter-gene covariance matrices.

Value

list with components "C" and "D" containing covariance matrices for the continuous and discrete components of the model.

defaultPrior — *Initialize a prior to be used a prior for BayeGLMlike/BayesGLMlike2*

Description

Initialize a prior to be used a prior for BayeGLMlike/BayesGLMlike2

Usage

```
defaultPrior(names)
```

Arguments

names	character vector of coefficients. The ‘(Intercept)‘ will be ignored.

Value

3d array, with leading dimension giving the prior ’loc’ation, ’scale’ and degrees of freedom (df), second dimension giving the component (’C’ontinuous or ’D’iscrete) and trailing dimension giving the coefficient to which the prior applies. The location is initialized to be 0, the scale to 2, and degrees of freedom of 1, following the default of bayesglm.

Examples

```
dp <- defaultPrior('Stim.ConditionUnstim')
## Not run:
data(vbetaFA)
zlmVbeta <- zlm(~ Stim.Condition, vbetaFA, method='bayesglm', coefPrior=dp)

## End(Not run)
```

`dof` *Degrees of freedom of Zero inflated model*

Description

Degrees of freedom of Zero inflated model

Usage

```
dof(object)
```

Arguments

`object`	LMlike or subclass

Value

vector giving the model degrees of freedom for continuous and discrete

---

`Drop` *Drop specified dimension from an array*

Description

Like drop(x) but only dropping specified dimensions. There is no testing that the specified dimensions are actually singletons.

Usage

```
Drop(x, d)
```

Arguments

`x`	array of at least d dimensions
`d`	dimension(s) to drop

Value

array x

Examples

```
x = array(1:4, dim=c(1, 2, 1, 2))
dx = MAST:::Drop(x, 1)
stopifnot(all(dim(dx)==c(2,1,2)))
```

---

ebayes *Estimate hyperparameters for hierarchical variance model for continuous component*

Description

ebayesControl is a named list with (optional) components 'method' (one of 'MOM' or 'MLE') and 'model' (one of 'H0' or 'H1') method MOM uses a method-of-moments estimator, while MLE using the marginal likelihood. H0 model estimates the precisions using the intercept alone in each gene, while H1 fits the full model specified by mm

Usage

```
ebayes(assay_t, ebayesControl, mm, truncate = Inf)
```

Arguments

assay_t	cells X genes matrix
ebayesControl	list with (optional) components 'method', 'model'. See details.
mm	a model matrix, used when model='H1'.
truncate	Genes with sample precisions exceeding this value are discarded when estimating the hyper parameters

Value

numeric of length two, giving the hyperparameters in terms of a variance (v) and prior observations (df), inside a structure, with component hess, giving the Fisher Information of the hyperparameters.

---

expavg *Exponential average*

Description

Puts log transformed values onto natural scale and takes mean of vector. Calculates mean(2^x - 1)

Usage

```
expavg(x)
```

Arguments

x	numeric

Value

numeric

Examples

```
x <- 1:10
logmean(expavg(x))
```

---

filterLowExpressedGenes *Filter low-expressing genes*

Description

Filter out genes that have less than some percent threshold expression across all libraries

Usage

```
filterLowExpressedGenes(assay, threshold = 0.1)
```

Arguments

assay	a SingleCellAssay object
threshold	a numeric between 0, and 1, specifying the threshold frequency below which genes will be filtered out

Value

SingleCellAssay

Examples

```
data(vbetaFA)
filterLowExpressedGenes(vbetaFA)
```

---

fit *fit a zero-inflated regression*

Description

Given a design and formula, fit the zero inflated regression, storing the fits in slots fitC and fitD

Usage

```
fit(object, response, ...)

## S4 method for signature 'LMERlike,missing'
fit(object, response, silent = TRUE, ...)
```

Arguments

object	inheriting from LMlike
response	a vector, same length as the design, or if missing then use the current response
...	currently ignored
silent	mute some warnings emitted from the underlying modeling functions

Value

LMlike or subclass

---

freq *Summary statistics for genes in an experiment*

Description

freq returns the frequency of expression, i.e., the proportion of non-zero values in sc. NAs can be optionally removed

Usage

```
freq(sc, na.rm = TRUE)

condmean(sc)

condSd(sc)

numexp(sc)
```

Arguments

sc	SingleCellAssay
na.rm	should NAs be removed, or carried through?

Value

vector of proportions

Functions

- condmean(): Report the mean non-zero expression value for each gene. NAs are always removed.
- condSd(): Report standard deviation of expression, for positive et for each gene. NAs are always removed.
- numexp(): Report number of expressing cells (>0) per gene. NAs are removed.

Examples

```
data(vbetaFA)
freq(vbetaFA)
condmean(vbetaFA)
```

FromFlatDF *Construct a SingleCellAssay (or derived subclass) from a 'flat' (melted) data.frame/data.table*

Description

SingleCellAssay are a generic container for such data and are simple wrappers around SummarizedExperiment objects. Subclasses exist that embue the container with additional attributes, eg FluidigmAssay.

Usage

```
FromFlatDF(
  dataframe,
  idvars,
  primerid,
  measurement,
  id = numeric(0),
  cellvars = NULL,
  featurevars = NULL,
  phenovars = NULL,
  class = "SingleCellAssay",
  check_sanity = TRUE,
  ...
)
```

Arguments

dataframe	A 'flattened' data.frame or data.table containing columns giving cell and feature identifiers and a measurement column
idvars	character vector naming columns that uniquely identify a cell
primerid	character vector of length 1 that names the column that identifies what feature (i.e. gene) was measured
measurement	character vector of length 1 that names the column containing the measurement
id	An identifier (eg, experiment name) for the resulting object
cellvars	Character vector naming columns containing additional cellular metadata
featurevars	Character vector naming columns containing additional feature metadata
phenovars	Character vector naming columns containing additional phenotype metadata
class	desired subclass of object. Default SingleCellAssay.
check_sanity	(default: TRUE) Set FALSE to override sanity checks that try to ensure that the default assay is log-transformed and has at least one exact zero. See defaultAssay for details on the "default assay" which is assumed to contain log transformed data.
...	additional arguments are ignored

Value

SingleCellAssay, or derived, object

Examples

```
data(vbeta)
colnames(vbeta)
vbeta <- computeEtFromCt(vbeta)
vbeta.fa <- FromFlatDF(vbeta, idvars=c("Subject.ID", "Chip.Number", "Well"),
primerid='Gene', measurement='Et', ncells='Number.of.Cells',
geneid="Gene",cellvars=c('Number.of.Cells', 'Population'),
phenovars=c('Stim.Condition','Time'), id='vbeta all', class='FluidigmAssay')
show(vbeta.fa)
nrow(vbeta.fa)
ncol(vbeta.fa)
head(mcols(vbeta.fa)$primerid)
table(colData(vbeta.fa)$Subject.ID)
vbeta.sub <- subset(vbeta.fa, Subject.ID=='Sub01')
show(vbeta.sub)
```

FromMatrix *Construct a SingleCellAssay from a matrix or array of expression*

Description

If the gene expression measurements are already in a rectangular form, then this function allows an easy way to construct a SingleCellAssay object while still doing some sanity checking of inputs.

Usage

```
FromMatrix(
  exprsArray,
  cData,
  fData,
  class = "SingleCellAssay",
  check_sanity = TRUE,
  check_logged = check_sanity
)
```

Arguments

exprsArray	matrix, or a list of matrices, or an array. Columns are cells, rows are genes.
cData	cellData an object that can be coerced to a DataFrame, ie, data.frame, AnnotatedDataFrame. Must have as many rows as `ncol(exprsArray)`
fData	featureData an object that can be coerced to a DataFrame, ie, data.frame, AnnotatedDataFrame. Must have as many rows as `nrow(exprsArray)`.
class	desired subclass of object. Default `SingleCellAssay`.
check_sanity	(default: `TRUE`) Set `FALSE` to override sanity checks that try to ensure that the default assay is log-transformed and has at least one exact zero. See defaultAssay for details on the "default assay" which is assumed to contain log transformed data.
check_logged	alias for `check_sanity`

Value

an object of class `class`

See Also

defaultAssay

Examples

```
ncells <- 10
ngenes <- 5
fData <- data.frame(primerid=LETTERS[1:ngenes])
cData <- data.frame(wellKey=seq_len(ncells))
mat <- matrix(rnorm(ncells*ngenes), nrow=ngenes)
sca <- FromMatrix(mat, cData, fData)
stopifnot(inherits(sca, 'SingleCellAssay'))
stopifnot(inherits(sca, 'SummarizedExperiment'))
##If there are mandatory keywords expected by a class, you'll have to manually set them yourself
cData$ncells <- 1
fd <- FromMatrix(mat, cData, fData)
stopifnot(inherits(fd, 'SingleCellAssay'))
```

---

getConcordance *Get the concordance between two experiments*

Description

Return the concordance between two assays (i.e. single cell and hundred cell). The "average" of `singleCellRef` (after adjusting for the number of cells) and `singleCellComp` are taken per gene, per `groups`. A `data.frame` with one row per gene-`groups` is returned with some additional columns.

Usage

```
getConcordance(
  singleCellRef,
  singleCellcomp,
  groups = NULL,
  fun.natural = expavg,
  fun.cycle = logmean
)

getwss(concord, nexp)

getss(concord)

getrc(concord)
```

Arguments

singleCellRef	"reference" SingleCellAssay
singleCellcomp	"comparison" SingleCellAssay
groups	character vector giving variable(s) on which the comparison is conditioned
fun.natural	function to transform the SingleCellAssays to a mRNA proportional level
fun.cycle	inverse function of fun.natural
concord	data.frame returned by getConcordance
nexp	number of expressed cells per row in concord

Value

concordance between two assays

Functions

- getwss(): getrc the sum of squares, weighted by nexp
- getss(): return the sum of squares
- getrc(): Return Lin's (1989) concordance correlation coefficient

Author(s)

Andrew McDavid

See Also

plotSCAConcordance

Examples

```
data(vbetaFA)
sca1 <- subset(vbetaFA, ncells==1)
sca100 <- subset(vbetaFA, ncells==100)
concord <- getConcordance(sca1, sca100)
getss(concord)
getrc(concord)
```

---

getwellKey *Accessor for wellKey*

Description

This returns the wellKey, which is a unique identifier generated by idvars in the mapping

Usage

```
getwellKey(sc)
```

Arguments

sc	An object with a wellKey

Value

integer giving the unique id generated

Examples

```
data(vbetaFA)
getwellKey(vbetaFA)
colData(vbetaFA)$wellKey
```

GLMlike-class *Wrapper for regular glm/lm*

Description

Wrapper for regular glm/lm

Usage

```
## S4 method for signature 'GLMlike'
vcov(object, which, ...)
```

Arguments

object	GLMlike
which	character, one of 'C', 'D'.
...	ignored

Value

covariance matrix

Methods (by generic)

- vcov(GLMlike): return the variance/covariance of component which

Slots

weightFun function to map expression values to probabilities of expression. Currently unused.

`gseaAfterBoot` *Gene set analysis for hurdle model*

Description

Modules defined in `sets` are tested for average differences in expression from the "average" gene. By using bootstraps, the between-gene covariance of terms in the hurdle model is found, and is used to adjust for coexpression between genes. We drop genes if the coefficient we are testing was not estimible in original model fit in `zFit` or in any of the bootstrap replicates (evidenced an `NA` in the bootstrap array). This might yield overly conservative inference. Since bootstrapping is a randomized procedure, the degrees of freedom of a module (and its variance parameters) might differ from run-to-run. You might try setting `var_estimate='modelbased'` to relax this requirement by assuming independence between genes and then using the asymptotic covariance estimates, which are deterministic, but may result in overly-generous inference.

Usage

```
gseaAfterBoot(
  zFit,
  boots,
  sets,
  hypothesis,
  control = gsea_control(n_randomize = Inf, var_estimate = "bootall")
)

gsea_control(n_randomize = Inf, var_estimate = "bootall")
```

Arguments

`zFit`	object of class ZlmFit
`boots`	bootstraps of zFit
`sets`	list of indices of genes
`hypothesis`	a `Hypothesis` to test. Currently only one degree `CoefficientHypothesis` are supported.
`control`	parameters as provided by `gsea_control`. See details.
`n_randomize`	the number of genes to sample to approximate the non-module average expression. Set to `Inf` to turn off the approximation (the default).
`var_estimate`	the method used to estimate the variance of the modules, one of `bootall`, `bootdiag`, or `modelbased`.

Value

Object of class `GSEATests`, containing slots `tests`, 4D array and `bootR`, the number of boostrap replicates.

Functions

- `gsea_control()`: set control parameters. See Details.

`control`

`control` is a list with elements:

- `n_randomize`, giving the number of genes to sample to approximate the non-module average expression. Set to `Inf` to turn off the approximation (the default).
- `var_estimate`, giving the method used to estimate the variance of the modules. `bootall` uses the bootstrapped covariance matrices. `bootdiag` uses only the diagonal of the bootstrapped covariance matrix (so assuming independence across genes). `modelbased` assumes independence across genes and uses the variance estimated from the model.

Return Value

A 4D array is returned, with dimensions "set" (each module), "comp" ('disc'rete or 'cont'inuous), "metric" ('stat' gives the average of the coefficient, 'var' gives the variance of that average, 'dof' gives the number of genes that were actually tested in the set), "group" ('test' for the genes in test-set, "null" for all genes outside the test-set).

See Also

calcZ

summary,GSEATests-method

Examples

```
data(vbetaFA)
vb1 = subset(vbetaFA, ncells==1)
vb1 = vb1[,freq(vb1)>.1][1:15,]
zf = zlm(~Stim.Condition, vb1)
boots = bootVcov1(zf, 5)
sets = list(A=1:5, B=3:10, C=15, D=1:5)
gsea = gseaAfterBoot(zf, boots, sets, CoefficientHypothesis('Stim.ConditionUnstim'))
## Use a model-based estimate of the variance/covariance.
gsea_mb = gseaAfterBoot(zf, boots, sets, CoefficientHypothesis('Stim.ConditionUnstim'),
control = gsea_control(var_estimate = 'modelbased'))
calcZ(gsea)
summary(gsea)
```

GSEATests-class *An S4 class for Gene Set Enrichment output*

Description

This holds output from a call to gseaAfterBoot. It primarily provides a summary method.

Slots

`tests` array: gene sets X discrete,continuous X stat, variance, degrees of freedom, avg correlation X test, null

`bootR` number of bootstrap replicates

See Also

gseaAfterBoot

calcZ

summary,GSEATests-method

`hushWarning` *Selectively muffle warnings based on output*

Description

Selectively muffle warnings based on output

Usage

```
hushWarning(expr, regexp)
```

Arguments

`expr`	an expression
`regexp`	a regexp to be matched (with str_detect)

Value

the result of expr

Examples

```
hushWarning(warning('Beware the rabbit'), 'rabbit')
hushWarning(warning('Beware the rabbit'), 'hedgehog')
```

`Hypothesis` *Describe a linear model hypothesis to be tested*

Description

A `Hypothesis` can be any linear combination of coefficients, compared to zero. Specify it as a character vector that can be parsed to yield the desired equalities ala `makeContrasts`. A `CoefficientHypothesis` is a hypothesis for which terms are singly or jointly tested to be zero (generally the case in a t-test or F-test), by dropping coefficients from the model.

Usage

```
Hypothesis(hypothesis, terms)
```

Arguments

`hypothesis`	a character vector specifying a hypothesis, following makeContrasts, or a character vector naming coefficients to be dropped.
`terms`	an optional character vector giving the terms (column names from the `model.matrix`) out of which the contrasts will be contrasted. If missing then most functions will attempt to fill this in for you at run time.

Value

a Hypothesis with a "transformed" component

See Also

zlm waldTest lrTest

Examples

```
h <- Hypothesis('Stim.ConditionUnstim', c('(Intercept)', 'Stim.ConditionUnstim'))
h@contrastMatrix
```

`impute` *impute missing continuous expression for plotting*

Description

If there are no positive observations for a contrast, it is generally not estimible. However, for the purposes of testing we can replace it with the least favorable value with respect to the contrasts that are defined.

Usage

```
impute(object, groupby)
```

Arguments

`object`	Output of predict
`groupby`	Variables (column names in predict) to group by for imputation (facets of the plot)

Value

data.table

Examples

```
##See stat_ell
example(stat_ell)
```

influence.bayesglm *Influence bayesglm object*

Description

The influence function

Usage

```
## S3 method for class 'bayesglm'
influence(model, do.coef = TRUE, ...)
```

Arguments

model	bayesglm
do.coef	see influence.glm
...	ignored

Value

see influence.glm

invlogit *Inverse of logistic transformation*

Description

Inverse of logistic transformation

Usage

```
invlogit(x)
```

Arguments

x	numeric

Value

numeric

Examples

```
x <- 1:5
invlogit(log(x/(1-x)))
```

LMERlike-class *Wrapper for lmer/glmer*

Description

A horrendous hack is employed in order to do arbitrary likelihood ratio tests: the model matrix is built, the names possibly mangled, then fed in as a symbolic formula to glmer/lmer. This is necessary because there is no (easy) way to specify an arbitrary fixed-effect model matrix in glmer.

Usage

```
## S4 method for signature 'LMERlike'
update(object, formula., design, keepDefaultCoef = FALSE, ...)

## S4 method for signature 'LMERlike'
vcov(object, which, ...)

## S4 method for signature 'LMERlike'
coef(object, which, singular = TRUE, ...)

## S4 method for signature 'LMERlike'
logLik(object)
```

Arguments

object	LMERlike
formula.	formula
design	something coercible to a data.frame
keepDefaultCoef	logical. Should the coefficient names be preserved from object or updated if the model matrix has changed?
...	In the case of vcov, ignored. In the case of update, passed to model.matrix.
which	character, one of 'C', 'D'.
singular	logical. Should NA coefficients be returned?

Value

see the section "Methods (by generic)"

Methods (by generic)

- update(LMERlike): update the formula or design matrix
- vcov(LMERlike): return the variance/covariance of component which
- coef(LMERlike): return the coefficients. The horrendous hack is attempted to be undone.
- logLik(LMERlike): return the log-likelihood

Slots

pseudoMM part of this horrendous hack.

strictConvergence logical (default: TRUE) return results even when the optimizer or *lmer complains about convergence

optimMsg character record warnings from lme. NA_character_ means no warnings.

LMlike-class *Linear Model-like Class*

Description

Wrapper around modeling function to make them behave enough alike that Wald tests and Likelihood ratio are easy to do. To implement a new type of zero-inflated model, extend this class. Depending on how different the method is, you will definitely need to override the `fit` method, and possibly the `model.matrix`, `model.matrix<-`, `update`, `coef`, `vcov`, and `logLik` methods.

Usage

```
## S4 method for signature 'LMlike'
summary(object)

## S4 method for signature 'LMlike'
update(object, formula., design, keepDefaultCoef = FALSE, ...)

## S4 method for signature 'LMlike,CoefficientHypothesis'
waldTest(object, hypothesis)

## S4 method for signature 'LMlike,matrix'
waldTest(object, hypothesis)

## S4 method for signature 'LMlike,character'
lrTest(object, hypothesis)

## S4 method for signature 'LMlike,CoefficientHypothesis'
lrTest(object, hypothesis)

## S4 method for signature 'LMlike,Hypothesis'
lrTest(object, hypothesis)

## S4 method for signature 'LMlike,matrix'
lrTest(object, hypothesis)

## S4 method for signature 'GLMlike'
logLik(object)
```

Arguments

object	LMlike
formula.	formula
design	something coercible to a data.frame

`keepDefaultCoef`
: `logical`. Should the coefficient names be preserved from `object` or updated if the model matrix has changed?

`...`
: passed to `model.matrix`

`hypothesis`
: one of a `CoefficientHypothesis`, `Hypothesis` or contrast `matrix`.

Value

see section "Methods (by generic)"

Methods (by generic)

- `summary(LMlike)`: Print a summary of the coefficients in each component.
- `update(LMlike)`: update the formula or design from which the `model.matrix` is constructed
- `waldTest(object = LMlike, hypothesis = CoefficientHypothesis)`: Wald test dropping single term specified by `CoefficientHypothesis` `hypothesis`
- `waldTest(object = LMlike, hypothesis = matrix)`: Wald test of contrast specified by contrast matrix `hypothesis`
- `lrTest(object = LMlike, hypothesis = character)`: Likelihood ratio test dropping entire term specified by `character` `hypothesis` naming a term in the symbolic formula.
- `lrTest(object = LMlike, hypothesis = CoefficientHypothesis)`: Likelihood ratio test dropping single term specified by `CoefficientHypothesis` `hypothesis`
- `lrTest(object = LMlike, hypothesis = Hypothesis)`: Likelihood ratio test dropping single term specified by `Hypothesis` `hypothesis`
- `lrTest(object = LMlike, hypothesis = matrix)`: Likelihood ratio test dropping single term specified by contrast matrix `hypothesis`
- `logLik(GLMlike)`: return the log-likelihood of a fitted model

Slots

design a data.frame from which variables are taken for the right hand side of the regression

fitC The continuous fit

fitD The discrete fit

response The left hand side of the regression

fitted A `logical` with components "C" and "D", TRUE if the respective component has converged

formula A `formula` for the regression

fitArgsC

fitArgsD Both `lists` giving arguments that will be passed to the fitter (such as convergence criteria or case weights)

See Also

coef

lrTest

waldTest

vcov

logLik

logFC *Calculate log-fold changes from hurdle model components*

Description

Using the delta method, estimate the log-fold change from a state given by a vector contrast0 and the state(s) given by contrast1.

Usage

```
logFC(zlmfit, contrast0, contrast1)

getLogFC(zlmfit, contrast0, contrast1)
```

Arguments

zlmfit	ZlmFit output
contrast0	vector of coefficients giving baseline contrast, or a Hypothesis. If missing, then the '(Intercept)' is used as baseline.
contrast1	matrix of coefficients giving comparison contrasts, or a Hypothesis. If missing, then all non-(Intercept) coefficients are compared.

Details

The log-fold change is defined as follows. For each gene, let $u(x)$ be the expected value of the continuous component, given a covariate x and the estimated coefficients coefC, ie, $u(x) =$ crossprod(x, coefC). Likewise, Let $v(x) =$ 1/(1+exp(-crossprod(coefD, x))) be the expected value of the discrete component. The log fold change from contrast0 to contrast1 is defined as

$$u(contrast1)v(contrast1) - u(contrast0)v(contrast0).$$

Note that for this to be a log-fold change, then the regression for u must have been fit on the log scale. This is returned in the matrix logFC. An approximation of the variance of logFC (applying the delta method to formula defined above) is provided in varLogFC.

Value

list of matrices 'logFC' and 'varLogFC', giving the log-fold-changes for each contrast (columns) and genes (rows) and the estimated sampling variance thereof

Functions

- getLogFC(): Return results as a perhaps friendlier data.table

Caveats

1. When method='bayesglm' (the default), it's no longer necessarily true that the log fold change from condition A to B will be the inverse of the log fold change from B to A if the models are fit separately. This is due to the shrinkage in bayesglm.

2. The log fold change can be small, but the Hurdle p-value small and significant when the sign of the discrete and continuous model components are discordant so that the marginal log fold change

cancels out. The large sample sizes present in many single cell experiments also means that there is substantial power to detect even small changes.

3. When there is no expression in a gene for a coefficient that is non-zero in either `condition0` or `condition1` we return `NA` because there is not any information to estimate the continuous component. Technically we might return plus or minus infinity, but there is not a straightforward way to estimate a confidence interval in any case. See https://support.bioconductor.org/p/99244/ for details

See Also

Hypothesis

summary,ZlmFit-method

Examples

```
data(vbetaFA)
zz <- zlm( ~ Stim.Condition+Population, vbetaFA[1:5,])
##log-fold changes in terms of intercept (which is Stim(SEB) and CD154+VbetaResponsive)
lfcStim <- logFC(zz)
##If we want to compare against unstim, we can try the following
coefnames <- colnames(coef(zz, 'D'))
contrast0 <- setNames(rep(0, length(coefnames)), coefnames)
contrast0[c('(Intercept)', 'Stim.ConditionUnstim')] <- 1
contrast1 <- diag(length(coefnames))
rownames(contrast1)<-colnames(contrast1)<-coefnames
contrast1['(Intercept)',]<-1
lfcUnstim <- logFC(zz, contrast0, contrast1)
##log-fold change with itself is 0
stopifnot(all(lfcUnstim$logFC[,2]==0))
##inverse of log-fold change with Stim as reference
stopifnot(all(lfcStim$logFC[,1]==(-lfcUnstim$logFC[,1])))
##As a data.table:
getLogFC(zz)
```

---

logmean *Log mean*

Description

Takes mean of natural scaled values and then logrithm Approximately the inverse operation of `expavg` Calculates log2(mean(x) + 1)

Usage

```
logmean(x)
```

Arguments

x	numeric

Value

numeric

Examples

```
x <- 1:10
expavg(logmean(x))
```

LRT *Likelihood Ratio Tests for SingleCellAssays*

Description

Tests for a change in ET binomial proportion or mean of positive ET Likelihood Ratio Test for SingleCellAssay objects

Usage

```
LRT(sca, comparison, ...)

## S4 method for signature 'SingleCellAssay,character'
LRT(sca, comparison, referent = NULL, groups = NULL, returnall = FALSE)
```

Arguments

sca	A SingleCellAssay class object
comparison	A character specifying the factor for comparison
...	ignored
referent	A character specifying the reference level of comparison.
groups	A optional character specifying a variable on which to stratify the test. For each level of groups, there will be a separate likelihood ratio test.
returnall	A logical specifying if additional rows should be returned with information about the different components of the test.

Details

Combined Likelihood ratio test (binomial and normal) for SingleCellAssay and derived objects. This function is deprecated, please use lrTest instead.

Value

data.frame

See Also

zlm ZlmFit

Examples

```
data(vbetaFA)
LRT(vbetaFA, 'Stim.Condition', 'Unstim')
```

lrTest *Run a likelihood-ratio test*

Description

Compares the change in likelihood between the current model and one subject to contrasts tested in `hypothesis`. `hypothesis` can be one of a `character` giving complete factors or terms to be dropped from the model, `CoefficientHypothesis` giving names of coefficients to be dropped, `Hypothesis` giving contrasts using the symbolically, or a contrast `matrix`, with one row for each coefficient in the full model, and one column for each contrast being tested.

Usage

```
lrTest(object, hypothesis, ...)
```

Arguments

`object`	LMlike or subclass
`hypothesis`	the hypothesis to be tested. See details.
`...`	optional arguments, passed to fitting functions

Value

array giving test statistics

See Also

fit

waldTest

Hypothesis

CoefficientHypothesis

Examples

```
#see ZlmFit-class for examples
example('ZlmFit-class')
```

lrTest,ZlmFit,character-method *Likelihood ratio test*

Description

A 3D array with first dimension being the genes, next dimension giving information about the test (the degrees of freedom, Chisq statistic, and P value), and final dimension being the value of these quantities on the discrete, continuous and hurdle (combined) levels.

Usage

```
## S4 method for signature 'ZlmFit,character'
lrTest(object, hypothesis, ...)
```

Arguments

object	ZlmFit
hypothesis	See Details
...	Arguments passed on to zlm
	formula a formula with the measurement variable on the LHS and predictors present in colData on the RHS
	sca SingleCellAssay object
	method character vector, either 'glm', 'glmer' or 'bayesglm'
	silent Silence common problems with fitting some genes
	ebayes if TRUE, regularize variance using empirical bayes method
	ebayesControl list with parameters for empirical bayes procedure. See ebayes.
	force Should we continue testing genes even after many errors have occurred?
	hook a function called on the fit after each gene.
	parallel If TRUE and option(mc.cores)>1 then multiple cores will be used in fitting.
	LMlike if provided, then the model defined in this object will be used, rather than following the formulas. This is intended for internal use.
	onlyCoef If TRUE then only an array of model coefficients will be returned (probably only useful for bootstrapping).
	exprs_values character or integer passed to 'assay' specifying which assay to use for testing

Value

3D array

---

magic_assay_names *Default* assay *returned*

Description

Methods in this package operate on log-transformed (multiplicative scale) expression. We attempt to check for this at construction, and then over-ride the assay method to return the "layer" containing such log-transformed data.

Usage

```
magic_assay_names()

assay_idx(x)

## S4 method for signature 'SingleCellAssay,missing'
assay(x, i, withDimnames = TRUE, ...)
```

Arguments

x	SingleCellAssay
i	must be missing for this method to apply
withDimnames	A logical(1), indicating whether the dimnames of the SummarizedExperiment object should be applied (i.e. copied) to the extracted assays. More precisely, setting withDimnames=FALSE in the *getter* returns the assays *as-is* whereas setting withDimnames=FALSE return them with possibly modified dimnames. Setting withDimnames=FALSE in the *setter* (assays<-) is required when the dimnames on the supplied assays are not identical to the dimnames on the SummarizedExperiment object; it does not influence actual assignment of dimnames to assays (they're always stored as-is). Note that `assays(x, withDimnames=FALSE) <- assays(x, withDimnames=FALSE)` is guaranteed to always work and be a no-op. This is not the case if withDimnames=TRUE is used or if withDimnames is not specified.
...	passed to parent method

Details

By default we return the assay whose names, as given by assayNames(x), matches the first element in the vector c('thresh', 'et', 'Et', 'lCount', 'logTPM', 'logCounts', 'logcounts').

Functions

- magic_assay_names(): list of names assumed to represent log-transformed data, in order of usage preference
- assay_idx(): what index is returned by default by 'assay'

Examples

```
data(vbetaFA)
assay(vbetaFA)[1:3,1:3]
assay(vbetaFA, 'thresh', withDimnames = FALSE) = assay(vbetaFA)*0 - 9
assay(vbetaFA)[1:3, 1:3]
```

---

maits *MAITs data set, RNASeq*

Description

MAITs data set, RNASeq

Format

a list containing an expression matrix (expressionmat), cell cdat and feature fdat.

See Also

FromMatrix

MAST-defunct *Defunct functions in package 'MAST'*

Description

These functions are defunct or have been renamed.

Functions (and replacements, if available)

filter mast_filter

`cData` colData

`fData` mcols

`exprs` assay

`zlm.SingleCellAssay` zlm

`combine` cbind or rbind

`deviance_residuals_hook` No replacement available, underlying API changed

mast_filter *Filter a SingleCellAssay*

Description

Remove, or flag wells that are outliers in discrete or continuous space.

Usage

```
mast_filter(sc, groups = NULL, filt_control = NULL, apply_filter = TRUE)

burdenOfFiltering(sc, groups, byGroup = FALSE, filt_control = NULL)
```

Arguments

`sc`	The `SingleCellAssay` object
`groups`	An optional `character` naming the grouping variable
`filt_control`	The `list` with configuration parameters for the filter.
`apply_filter`	`logical` should the filter be applied, or should a matrix of booleans giving if a well would be subject to a filtering criteria be returned?
`byGroup`	in the case of `burdenOfFiltering` should the filter be stratified by groups, or only the plotting.

Details

The function filters wells that don't pass filtering criteria described in filt_control. filt_control is a list with named elements `nOutlier` (minimum nmber of outlier cells for a cell to be filtered [default = 2] `sigmaContinuous` (the z-score outlier threshold for the continuous part of the signal) [default = 7] and `sigmaProportion` (the z-score outlier threshold for the discrete part of the signal) [default = 7].

If `groups` is provided, the filtering is calculated within each level of the group, then combined again as output.

Value

A filtered result

Functions

- `burdenOfFiltering()`: plot the proportions of wells are filtered due to different criteria

Author(s)

Andrew McDavid

See Also

burdenOfFiltering

Examples

```
data(vbetaFA)
## Split by 'ncells', apply to each component, then recombine
vbeta.filtered <- mast_filter(vbetaFA, groups='ncells')
## Returned as boolean matrix
was.filtered <- mast_filter(vbetaFA, apply_filter=FALSE)
## Wells filtered for being discrete outliers
head(subset(was.filtered, pctout))
burdenOfFiltering(vbetaFA, groups='ncells', byGroup=TRUE)
burdenOfFiltering(vbetaFA, groups='ncells')
```

`meld_list_left` *Combine lists, preferentially taking elements from x if there are duplicate names*

Description

Combine lists, preferentially taking elements from x if there are duplicate names

Usage

```
meld_list_left(x, y)
```

Arguments

`x`	list
`y`	list

Examples

```
MAST:::meld_list_left(list(A=1, B=2), list(A = 0))
```

melt.SingleCellAssay *"Melt" a* SingleCellAssay *matrix*

Description

Return a molten (flat) representation, taking the cross-product of the expression values, the colData (column meta data), and the feature data (mcols).

Usage

```
## S3 method for class 'SingleCellAssay'
melt(data, ..., na.rm = FALSE, value.name = "value")
```

Arguments

data	SingleCellAssay
...	ignored
na.rm	ignored
value.name	name of 'values' column in returned value

Value

A data.table, with the cartesian product of the row and column attributes and the expression values

Examples

```
data(vbetaFA)
melt.SingleCellAssay(vbetaFA[1:10,])
as(vbetaFA[1:10,], 'data.table')
```

model.matrix *Model matrix accessor*

Description

Model matrix accessor

Usage

```
model.matrix(object, ...)

## S4 method for signature 'LMlike'
model.matrix(object, ...)
```

Arguments

object	LMlike or subclass
...	ignored

Value

model.matrix if present

Methods (by class)

- `model.matrix(LMlike)`: return the `model.matrix`

`model.matrix<-` *Replace model matrix*

Description

Replace model matrix

Usage

```
model.matrix(object) <- value
```

Arguments

`object`	LMlike or subclass
`value`	matrix

Value

modify object

`myBiplot` *Makes a nice BiPlot*

Description

Creates a custom BiPlot for visualizing the results of PCA

Usage

```
myBiplot(pc, colorfactor, scaling = 100, nudge = 1.2, N = 10, dims = 1:2, ...)
```

Arguments

`pc`	output of `prcomp`
`colorfactor`	a `factor` the same length as `nrow(pc$x)` to color the points
`scaling`	`integer` to scale the vectors showing loadings
`nudge`	`numeric` to offset labels for loadings
`N`	number of variables with longest `dim[1]` or `dim[2]` projections to display
`dims`	`numeric` vector of length 2 indicating which PCs to plot
`...`	passed to plot

Value

printed plot

new_with_repaired_slots *Instantiate a class, but warn rather than error for badly named slots*

Description

Instantiate a class, but warn rather than error for badly named slots

Usage

```
new_with_repaired_slots(classname, ..., extra)
```

Arguments

classname	‘character‘ naming a class
...	slots in ‘classname‘
extra	named list giving other slots in ‘classname‘

Value

‘new(classname)‘

Examples

```
MAST:::new_with_repaired_slots("SimpleList",  listData = list(x = LETTERS),
extra = list(elementType = 'character', food = "tasty", beer = "cold"))
```

pbootVcov1 *Bootstrap a zlmfit*

Description

Sample cells with replacement to find bootstrapped distribution of coefficients

Usage

```
pbootVcov1(cl, zlmfit, R = 99)

bootVcov1(zlmfit, R = 99, boot_index = NULL)
```

Arguments

cl	a cluster object created by makeCluster
zlmfit	class ZlmFit
R	number of bootstrap replicates
boot_index	list of indices to resample. Only one of R or boot_index can be offered.

Value

array of bootstrapped coefficients

array of bootstrapped coefficients

Functions

- `pbootVcov1()`: parallel version of bootstrapping

Examples

```
data(vbetaFA)
zlmVbeta <- zlm(~ Stim.Condition, subset(vbetaFA, ncells==1)[1:5,])
#Only run 3 boot straps, which you wouldn't ever want to do in practice...
bootVcov1(zlmVbeta, R=3)
```

`plot.thresholdSCRNACountMatrix` *Plot cutpoints and densities for thresholding*

Description

Plot cutpoints and densities for thresholding

Usage

```
## S3 method for class 'thresholdSCRNACountMatrix'
plot(x, ask = FALSE, wait.time = 0, type = "bin", indices = NULL, ...)
```

Arguments

`x`	output of `thresholdSCRNACountMatrix`
`ask`	if TRUE then will prompt before displaying each plot
`wait.time`	pause (in seconds) between each plot
`type`	one or more of the following: 'bin' (plot the genes by the binning used for thresholding), or 'gene' (plot thresholding by gene – see next argument)
`indices`	if `type` is equal to 'gene', and is a integer of length 1, then a random sample of `indices` genes is taken. If it is NULL, then 10 genes are sampled. If it is a integer vector of length > 1, then it is interpreted as giving a list of indices of genes to be displayed.
`...`	further arguments passed to `plot`

Value

displays plots

Examples

```
## See thresholdSCRNACountMatrix
example(thresholdSCRNACountMatrix)
```

plotlrt *Plot a likelihood ratio test object*

Description

Constructs a forest-like plot of signed log10 p-values, possibly adjusted for multiple comparisons `adjust` can be one of "holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr", "none".

Usage

```
plotlrt(lr, adjust = "fdr", thres = 0.1, trunc = 1e-06, groups = NULL)
```

Arguments

`lr`	output from lrtest, with returnall=FALSE
`adjust`	`character`, passed along to `p.adjust`, see below
`thres`	`numeric` genes with adjusted pvalues above this value are not depicted
`trunc`	`numeric` p values below this value are truncated at this value
`groups`	`character` grouping value. If provided, must match groups argument passed to lrtest. Plots done separately for each group.

Value

Constructs a dotplot

Author(s)

andrew

plotSCAConcordance *Concordance plots of filtered single vs n-cell assays*

Description

Plot the average expression value of two subsets of the data. Generally these might be 1 cell and multiple-cell replicates, in which case if the `mcols` column `ncells` is set then the averages will be adjusted accordingly. But it could be any grouping.

Usage

```
plotSCAConcordance(
  SCellAssay,
  NCellAssay,
  filterCriteria = list(nOutlier = 2, sigmaContinuous = 9, sigmaProportion = 9),
  groups = NULL,
  ...
)
```

Arguments

SCellAssay	is a FluidigmAssay for the 1-cell per well assay
NCellAssay	is a FluidigmAssay for the n-cell per well assay
filterCriteria	is a list of filtering criteria to apply to the SCellAssay and NCellAssay
groups	is a character vector naming the group within which to perform filtering. NULL by default.
...	passed to getConcordance

Value

printed plot

See Also

getConcordance

Examples

```
data(vbetaFA)
sca1 <- subset(vbetaFA, ncells==1)
sca100 <- subset(vbetaFA, ncells==100)
plotSCAConcordance(sca1, sca100)
```

---

predict.ZlmFit *Return predictions from a ZlmFit object.*

Description

Return predictions from a ZlmFit object.

Usage

```
## S3 method for class 'ZlmFit'
predict(object, newdata = NULL, modelmatrix = NULL, ...)
```

Arguments

object	A ZlmFit
newdata	The data to predict from. Currently ignored, will use the data in the object.
modelmatrix	The model matrix specifying the linear combination of coefficients.
...	ignored

Value

Predictions (on the link scale) and standard errors.

Examples

```
##See stat_ell
example(stat_ell)
```

predicted_sig *Predicted signatures*

Description

Predicted signatures

Format

A data frame of predicted gene expresion signatures for stimulated and unstimulated cells.

primerAverage *Average expression values for duplicated/redundant genes*

Description

Takes an average, potentially on a different scale given by `fun.natural` of some genes. The average is then transformed with `fun.cycle`.

Usage

```
primerAverage(fd, geneGroups, fun.natural = expavg, fun.cycle = logshift)
```

Arguments

`fd`	`SingleCellAssay` or subclass
`geneGroups`	`character` naming a column in the `featureData` that keys the duplicates
`fun.natural`	transformation to be used to collapse the duplicate expression values
`fun.cycle`	transformation to be used after collapsing

Value

averaged version of `fd`.

Note

This code needs to be tested more extensively after a refactoring. Caveat calculator.

print.summaryZlmFit *Print summary of a ZlmFit*

Description

Shows the top ‘n‘ genes by z score on ‘by‘

Usage

```
## S3 method for class 'summaryZlmFit'
print(x, n = 2, by = "logFC", ...)
```

Arguments

x	output from summary(ZlmFit)
n	number of genes to show
by	one of 'C' , 'D' or 'logFC' for continuous, discrete and log fold change z-scores for each contrast
...	ignored

Value

prints a pretty table and invisibly returns a `data.table` representing the table.

See Also

summary,ZlmFit-method

read.fluidigm *Reads a Fluidigm Biomark (c. 2011) raw data file (or set of files)*

Description

This function reads a raw Fluidigm Biomark data file or set of files and constructs a SingleCellAssay (or FluigidmAssay) object. This was written c. 2011 and has not been tested lately. The Biomark format may have changed.

Usage

```
read.fluidigm(
  files = NULL,
  metadata = NULL,
  header.size = 2,
  skip = 8,
  cycle.threshold = 40,
  metadataColClasses = NULL,
  meta.key = NULL,
  idvars = NULL,
  splitby = NULL,
```

```
  unique.well.id = "Chamber.ID",
  raw = TRUE,
  assay = NULL,
  geneid = "Assay.Name",
  sample = NULL,
  well = "Well",
  measurement = "X40.Ct",
  measurement.processed = "Ct",
  ncells = "SampleRConc"
)
```

Arguments

files	A character vector of files to read.
metadata	A character path and filename of a CSV file containing additional metadata about the samples
header.size	A numeric indicating the number of lines in the header (default 2)
skip	numeric how many lines to skip before reading (default 8)
cycle.threshold	The maximum number of PCR cycles performed (default 40) numeric
metadataColClasses	Optional character vector giving the column classes of the metadata file. See read.table.
meta.key	Optional character vector that identifies the key column between the metadata and the fluidigm data
idvars	Optional character vector that defines the set of columns uniquely identifying a well (unique cell, gene, and condition).
splitby	Optional character that defines the column / variable used to split the resulting data into a list of SingleCellAssay, such that unique levels of splitby each fall into their own SingleCellAssay. Ususally the experimental unit subjected to different treatments.
unique.well.id	The column that uniquely identifies a sample well in the data. Default is "Chamber.ID".
raw	logical flag indicating this is raw data coming off the instrument. Thus we make some assumptions about the column names that are present.
assay	character name of a column that uniquely identifies an Assay (i.e. gene). Default is NULL
geneid	character names of the column that identifies a gene. Default is "Assay.Name"
sample	character name of a column that uniquely identifies a sample
well	character name of a column that uniquely identifies a well. Default "Well".
measurement	character name of the column that holds the measurement. Default "X40.Ct".
measurement.processed	character one of "Ct","40-Ct", or "et". If not "Ct", the measurement will be transformed.
ncells	The column with the number of cells in this well.

Value

list of SingleCellAssay holding the data.

Author(s)

Greg Finak

`removeResponse` *Remove the left hand side (response) from a formula*

Description

The order of terms will be rearrange to suit R's liking for hierarchy but otherwise the function should be idempotent for

Usage

```
removeResponse(Formula, warn = TRUE)
```

Arguments

`Formula`	formula
`warn`	Issue a warning if a response variable is found?

Value

formula

Author(s)

Andrew

`rstandard.bayesglm` *rstandard for bayesglm objects.*

Description

rstandard bayesglm object S3 method

Usage

```
## S3 method for class 'bayesglm'
rstandard(
  model,
  infl = influence(model, do.coef = FALSE),
  type = c("deviance", "pearson"),
  ...
)
```

Arguments

model	bayesglm
infl	see rstandard
type	see rstandard
...	ignored

Value

numeric residuals

SceToSingleCellAssay *Coerce a SingleCellExperiment to some class defined in MAST*

Description

Coerce a SingleCellExperiment to some class defined in MAST

Usage

```
SceToSingleCellAssay(sce, class = "SingleCellAssay", check_sanity = TRUE)
```

Arguments

sce	object inheriting from SingleCellExperiment
class	character naming the class to be coerced to
check_sanity	(default: TRUE) Set FALSE to override sanity checks that try to ensure that the default assay is log-transformed and has at least one exact zero. See defaultAssay for details on the "default assay" which is assumed to contain log transformed data.

Value

object of the indicated class.

---

se.coef *Return coefficient standard errors*

Description

Given a fitted model, return the standard errors of the coefficient

Usage

```
se.coef(object, ...)
```

Arguments

object	a model implementing vcov
...	passed to methods

Value

vector or matrix

See Also

ZlmFit-class

Examples

```
#see ZlmFit-class for examples
example('ZlmFit-class')
```

---

show,LMlike-method *show*

Description

Display info

Usage

```
## S4 method for signature 'LMlike'
show(object)

## S4 method for signature 'ZlmFit'
show(object)
```

Arguments

object	an object of some type

Details

Prints information on a LMlike object

Value

side effect of printing to console

Methods (by class)

- show(ZlmFit): print info on ZlmFit

split,SingleCellAssay,character-method *Split into* list

Description

Splits a SingleCellAssay into a list by a factor (or something coercible into a factor) or a character giving a column of colData(x)

Usage

```
## S4 method for signature 'SingleCellAssay,character'
split(x, f, drop = FALSE, ...)
```

Arguments

x	SingleCellAssay
f	length-1 character, or atomic of length ncol(x)
drop	drop unused factor levels
...	ignored

Value

List

Examples

```
data(vbetaFA)
split(vbetaFA, 'ncells')
fa <- as.factor(colData(vbetaFA)$ncells)
split(vbetaFA, fa)
```

stat_ell *Plot confidence ellipse in 2D*

Description

The focus of the ellipse will be the point (x, y) and semi-major axes aligned with the coordinate axes and scaled by xse, yse and the level.

Usage

```
stat_ell(
  mapping = NULL,
  data = NULL,
  geom = "polygon",
  position = "identity",
  na.rm = FALSE,
  show.legend = NA,
  inherit.aes = TRUE,
  fill = NA,
  level = 0.95,
  lty = 2,
  invert = FALSE,
  alpha = 1,
  ...
)
```

Arguments

mapping	Set of aesthetic mappings created by aes or aes_. If specified and inherit.aes = TRUE (the default), it is combined with the default mapping at the top level of the plot. You must supply mapping if there is no plot mapping.
data	The data to be displayed in this layer. There are three options: If NULL, the default, the data is inherited from the plot data as specified in the call to ggplot. A data.frame, or other object, will override the plot data. All objects will be fortified to produce a data frame. See fortify for which variables will be created. A function will be called with a single argument, the plot data. The return value must be a data.frame., and will be used as the layer data.
geom	The geometric object to use display the data
position	Position adjustment, either as a string, or the result of a call to a position adjustment function.
na.rm	If FALSE (the default), removes missing values with a warning. If TRUE silently removes missing values.
show.legend	logical. Should this layer be included in the legends? NA, the default, includes if any aesthetics are mapped. FALSE never includes, and TRUE always includes.
inherit.aes	If FALSE, overrides the default aesthetics, rather than combining with them. This is most useful for helper functions that define both data and aesthetics and shouldn't inherit behaviour from the default plot specification, e.g. borders.
fill	A color or aesthetic mapping to fill color. Defaults to NA for empty ellipses.
level	The confidence level at which to draw an ellipse (default is level=0.95).
lty	The linetype to use. Can map to a variable. Defaults to 2 (dashed line)
invert	vector of length 1 that should either be "x","y",or TRUE. Specifies whether to plot the estimates from the discrete component on the inverse logit scale. invert specifies which axis to invert.
alpha	transparency
...	other arguments passed on to layer. These are often aesthetics, used to set an aesthetic to a fixed value, like color = "red" or size = 3. They may also be parameters to the paired geom/stat.

Value

ggplot layer

Examples

```
data(vbetaFA)
library(ggplot2)
zlmCond <- zlm(~Stim.Condition, vbetaFA[1:10,])
MM <- model.matrix(~Stim.Condition,unique(colData(vbetaFA)[,c("Stim.Condition"),drop=FALSE]))
predicted <- predict(zlmCond,modelmatrix=MM)
plt <- ggplot(predicted)+aes(x=invlogit(etaD),y=muC,xse=seD,yse=seC,col=sample)+
   facet_wrap(~primerid,scales="free_y")+theme_linedraw()+
   geom_point(size=0.5)+scale_x_continuous("Proportion expression")+
   scale_y_continuous("Estimated Mean")+
   stat_ell(aes(x=etaD,y=muC),level=0.95, invert='x')
## plot with inverse logit transformed x-axis
print(plt)
# doesn't do anything in this case because there are no inestimable coefficients
predictI <- impute(predicted, groupby='primerid')
```

---

subset,SingleCellAssay-method

Subset a SingleCellAssay *by cells (columns)*

Description

Evaluates the expression in ... in the context of colData(x) and returns a subsetted version of x

Usage

```
## S4 method for signature 'SingleCellAssay'
subset(x, ...)
```

Arguments

x	SingleCellAssay
...	expression

Value

SingleCellAssay

Examples

```
data(vbetaFA)
subset(vbetaFA, ncells==1)
```

summarize *Return programmatically useful summary of a fit*

Description

Return programmatically useful summary of a fit

Usage

```
summarize(object, ...)
```

Arguments

object	LMlike or subclass
...	other arguments

Value

list of parameters characterizing fit

summary,GSEATests-method *Summarize gene set enrichment tests*

Description

Returns a data.table with one row per gene set. This data.table contains columns:

set name of gene set

cond_Z Z statistic for continuous component

cont_P wald P value

cont_effect difference in continuous regression coefficients between null and test sets (ie, the numerator of the Z-statistic.)

disc_Z Z statistic for discrete

disc_P wald P value

disc_effect difference in discrete regression coefficients between null and test sets.

combined_Z combined discrete and continuous Z statistic using Stouffer's method

combined_P combined P value

combined_adj FDR adjusted combined P value

Usage

```
## S4 method for signature 'GSEATests'
summary(object, ...)
```

Arguments

object	A GSEATests object
...	passed to calcZ

Value

data.table

See Also

gseaAfterBoot

Examples

```
## See the examples in gseaAfterBoot
example(gseaAfterBoot)
```

---

summary,ZlmFit-method *Summarize model features from a* ZlmFit *object*

Description

Returns a data.table with a special print method that shows the top 2 most significant genes by contrast. This data.table contains columns:

primerid the gene

component C=continuous, D=discrete, logFC=log fold change, S=combined using Stouffer's method, H=combined using hurdle method

contrast the coefficient/contrast of interest

ci.hi upper bound of confidence interval

ci.lo lower bound of confidence interval

coef point estimate

z z score (coefficient divided by standard error of coefficient)

Pr(>Chisq) likelihood ratio test p-value (only if doLRT=TRUE)

Some of these columns will contain NAs if they are not applicable for a particular component or contrast.

Usage

```
## S4 method for signature 'ZlmFit'
summary(
  object,
  logFC = TRUE,
  doLRT = FALSE,
  level = 0.95,
  parallel = FALSE,
  ...
)
```

Arguments

object	A ZlmFit object
logFC	If TRUE, calculate log-fold changes, or output from a call to getLogFC.
doLRT	if TRUE, calculate lrTests on each coefficient, or a character vector of such coefficients to consider.
level	what level of confidence coefficient to return. Defaults to 95 percent.
parallel	If TRUE and option(mc.cores)>1 then multiple cores will be used in fitting.
...	ignored

Value

data.table

See Also

print.summaryZlmFit

Examples

```
data(vbetaFA)
z <- zlm(~Stim.Condition, vbetaFA[1:5,])
zs <- summary(z)
names(zs)
print(zs)
##Select `datatable` copmonent to get normal print method
zs$datatable
## Can use parallel processing for LRT now
summary(z, doLRT = TRUE, parallel = TRUE)
```

---

summary.thresholdSCRNACountMatrix

Summarize the effect of thresholding

Description

Returns the proportion of (putative) expression, the variance of expressed cells, and -log10 shapiro-wilk tests for normality on the expressed cells

Usage

```
## S3 method for class 'thresholdSCRNACountMatrix'
summary(object, ...)

## S3 method for class 'summaryThresholdSCRNA'
print(x, ...)
```

Arguments

object	a thresholdSCRNACountMatrix
...	currently ignored
x	a summaryThresholdSCRNA object, ie output from summary.thresholdSCRNACountMatrix

Value

a list of statistics on the original data, and thresholded data

Functions

- `print(summaryThresholdSCRNA)`: prints five-number distillation of the statistics and invisibly returns the table used to generate the summary

---

`thresholdSCRNACountMatrix` *Threshold a count matrix using an adaptive threshold.*

Description

An adaptive threshold is calculated from the conditional mean of expression, based on 10 bins of the genes with similar expression levels. Thresholds are chosen by estimating cutpoints in the bimodal density estimates of the binned data. These density estimates currently exclude the zeros due to complications with how the bandwidth is selected. (If the bandwith is too small, then extra peaks/modes are found and everything goes haywire). If the diagnostic plots don't reveal any bimodal bins, this is probably the reason, and you may not need to threshold since background in the data are exact zeros.

Usage

```
thresholdSCRNACountMatrix(
  data_all,
  conditions = NULL,
  cutbins = NULL,
  nbins = 10,
  bin_by = "median",
  qt = 0.975,
  min_per_bin = 50,
  absolute_min = 0,
  data_log = TRUE,
  adj = 1
)
```

Arguments

`data_all`	`matrix` of (possibly log-transformed) counts or TPM. Rows are genes and columns are cells.
`conditions`	Bins are be determined per gene and per condition. Typically contrasts of interest should be specified.
`cutbins`	`vector` of cut points.
`nbins`	`integer` number of bins when cutbins is not specified.
`bin_by`	`character` "median", "proportion", "mean"
`qt`	when `bin_by` is "quantile", what quantile should be used to form the bins
`min_per_bin`	minimum number of genes within a bin
`absolute_min`	`numeric` giving a hard threshold below which everything is assumed to be noise

data_log	is data_all log+1 transformed? If so, it will be returned on the (log+1)-scale as well.
adj	bandwith adjustment, passed to density

Value

list of thresholded counts (on natural scale), thresholds, bins, densities estimated on each bin, and the original data

Examples

```
data(maits,package='MAST', envir = environment())
sca <- FromMatrix(t(maits$expressionmat[,1:1000]), maits$cdat, maits$fdat[1:1000,])
tt <- thresholdSCRNACountMatrix(assay(sca))
tt <- thresholdSCRNACountMatrix(2^assay(sca)-1, data_log=FALSE)
opar <- par(no.readonly = TRUE)
on.exit(par(opar))
par(mfrow=c(4,2))
plot(tt)
```

---

vbeta *Vbeta Data Set*

Description

Vbeta Data Set

Format

a data frame with 11 columns. Column Ct contains the cycle threshold, with NA denoting that the threshold was never crossed. So it is inversely proportional to the log2 mRNA, and should be negated (and NAs set to zero) if it is used as a expression measurement for a FluidigmAssay.

---

vbetaFA *Vbeta Data Set, FluidigmAssay*

Description

Vbeta Data Set, FluidigmAssay

Format

a FluidigmAssay of the vbeta data set.

See Also

vbeta, FromFlatDF

waldTest *Run a Wald test*

Description

Run a Wald tests on discrete and continuous components `hypothesis` can be one of a `character` giving complete factors or terms to be dropped from the model, `CoefficientHypothesis` giving names of coefficients to be dropped, `Hypothesis` giving contrasts using the symbolically, or a contrast `matrix`, with one row for each coefficient in the full model, and one column for each contrast being tested.

Usage

```
waldTest(object, hypothesis)
```

Arguments

`object`	LMlike or subclass
`hypothesis`	the hypothesis to be tested. See details.

Value

array giving test statistics

See Also

fit

lrTest

lht

Examples

```
#see ZlmFit-class for examples
example('ZlmFit-class')
```

waldTest,ZlmFit,matrix-method *Wald test*

Description

A 3D array with first dimension being the genes, next dimension giving information about the test (the degrees of freedom, Chisq statistic, and P value), and final dimension being the value of these quantities on the discrete, continuous and hurdle (combined) levels.

Usage

```
## S4 method for signature 'ZlmFit,matrix'
waldTest(object, hypothesis)
```

Arguments

object	ZlmFit
hypothesis	See Details

Value

3D array

---

xform *Make matrix of continuous expression values, orthogonal to discrete*

Description

This centers each column of mat around the mean of its non-zero values.

Usage

```
xform(mat, scale = FALSE)
```

Arguments

mat	matrix (such as produced by exprs)
scale	should the columns also be scaled to have unit variance

Value

matrix

---

zlm *Zero-inflated regression for SingleCellAssay*

Description

For each gene in sca, fits the hurdle model in formula (linear for et>0), logistic for et==0 vs et>0. Return an object of class ZlmFit containing slots giving the coefficients, variance-covariance matrices, etc. After each gene, optionally run the function on the fit named by 'hook'

Usage

```
zlm(
  formula,
  sca,
  method = "bayesglm",
  silent = TRUE,
  ebayes = TRUE,
  ebayesControl = NULL,
  force = FALSE,
  hook = NULL,
  parallel = TRUE,
```

```
  LMlike,
  onlyCoef = FALSE,
  exprs_values = assay_idx(sca)$aidx,
  ...
)
```

Arguments

formula	a formula with the measurement variable on the LHS and predictors present in colData on the RHS
sca	SingleCellAssay object
method	character vector, either 'glm', 'glmer' or 'bayesglm'
silent	Silence common problems with fitting some genes
ebayes	if TRUE, regularize variance using empirical bayes method
ebayesControl	list with parameters for empirical bayes procedure. See ebayes.
force	Should we continue testing genes even after many errors have occurred?
hook	a function called on the fit after each gene.
parallel	If TRUE and option(mc.cores)>1 then multiple cores will be used in fitting.
LMlike	if provided, then the model defined in this object will be used, rather than following the formulas. This is intended for internal use.
onlyCoef	If TRUE then only an array of model coefficients will be returned (probably only useful for bootstrapping).
exprs_values	character or integer passed to 'assay' specifying which assay to use for testing
...	arguments passed to the S4 model object upon construction. For example, fitArgsC and fitArgsD, or coefPrior.

Value

a object of class ZlmFit with methods to extract coefficients, etc. OR, if data is a data.frame just a list of the discrete and continuous fits.

Empirical Bayes variance regularization

The empirical bayes regularization of the gene variance assumes that the precision (1/variance) is drawn from a gamma distribution with unknown parameters. These parameters are estimated by considering the distribution of sample variances over all genes. The procedure used for this is determined from ebayesControl, a named list with components 'method' (one of 'MOM' or 'MLE') and 'model' (one of 'H0' or 'H1') method MOM uses a method-of-moments estimator, while MLE using the marginal likelihood. H0 model estimates the precisions using the intercept alone in each gene, while H1 fits the full model specified by formula

See Also

ZlmFit-class, ebayes, GLMlike-class, BayesGLMlike-class

Examples

```
data(vbetaFA)
zlmVbeta <- zlm(~ Stim.Condition, subset(vbetaFA, ncells==1)[1:10,])
slotNames(zlmVbeta)
#A matrix of coefficients
coef(zlmVbeta, 'D')['CCL2',]
#An array of covariance matrices
vcov(zlmVbeta, 'D')[,,'CCL2']
waldTest(zlmVbeta, CoefficientHypothesis('Stim.ConditionUnstim'))

## Can also provide just a \code{data.frame} instead
data<- data.frame(x=rnorm(500), z=rbinom(500, 1, .3))
logit.y <- with(data, x*2 + z*2); mu.y <- with(data, 10+10*x+10*z + rnorm(500))
y <- (runif(500)<exp(logit.y)/(1+exp(logit.y)))*1
y[y>0] <- mu.y[y>0]
data$y <- y
fit <- zlm(y ~ x+z, data)
summary.glm(fit$disc)
```

ZlmFit-class *An S4 class to hold the output of a call to zlm*

Description

This holds output from a call to zlm. Many methods are defined to operate on it. See below.

Usage

```
## S4 method for signature 'ZlmFit,CoefficientHypothesis'
lrTest(object, hypothesis, ...)

## S4 method for signature 'ZlmFit,Hypothesis'
lrTest(object, hypothesis, ...)

## S4 method for signature 'ZlmFit,matrix'
lrTest(object, hypothesis, ...)

## S4 method for signature 'ZlmFit,CoefficientHypothesis'
waldTest(object, hypothesis)

## S4 method for signature 'ZlmFit,Hypothesis'
waldTest(object, hypothesis)

## S4 method for signature 'ZlmFit'
coef(object, which, ...)

## S4 method for signature 'ZlmFit'
vcov(object, which, ...)

## S4 method for signature 'ZlmFit'
se.coef(object, which, ...)
```

Arguments

object	ZlmFit
hypothesis	call to Hypothesis or CoefficientHypothesis or a matrix giving such contrasts.
...	ignored
which	character vector, one of "C" (continuous) or "D" (discrete) specifying which component should be returned

Value

see "Methods (by generic)"

Methods (by generic)

- lrTest(object = ZlmFit, hypothesis = CoefficientHypothesis): Returns an array with likelihood-ratio tests on contrasts defined using CoefficientHypothesis().
- lrTest(object = ZlmFit, hypothesis = Hypothesis): Returns an array with likelihood-ratio tests specified by Hypothesis, which is a Hypothesis.
- lrTest(object = ZlmFit, hypothesis = matrix): Returns an array with likelihood-ratio tests specified by Hypothesis, which is a contrast matrix.
- waldTest(object = ZlmFit, hypothesis = CoefficientHypothesis): Returns an array with Wald Tests on contrasts defined using CoefficientHypothesis().
- waldTest(object = ZlmFit, hypothesis = Hypothesis): Returns an array with Wald Tests on contrasts defined in Hypothesis()
- coef(ZlmFit): Returns the matrix of coefficients for component which.
- vcov(ZlmFit): Returns an array of variance/covariance matrices for component which.
- se.coef(ZlmFit): Returns a matrix of standard error estimates for coefficients on component which.

Slots

coefC matrix of continuous coefficients

coefD matrix of discrete coefficients

vcovC array of variance/covariance matrices for coefficients

vcovD array of variance/covariance matrices for coefficients

LMlike the LmWrapper object used

sca the SingleCellAssay object used

deviance matrix of deviances

loglik matrix of loglikelihoods

df.null matrix of null (intercept only) degrees of freedom

df.resid matrix of residual DOF

dispersion matrix of dispersions (after shrinkage)

dispersionNoShrink matrix of dispersion (before shrinkage)

priorDOF shrinkage weight in terms of number of psuedo-obs

priorVar shrinkage target

converged output that may optionally be set by the underlying modeling function

hookOut a list of length ngenes containing output from a hook function, if zlm was called with one

exprs_values ‘character‘ or ‘integer‘ with the ‘assay‘ used.

See Also

zlm summary,ZlmFit-method

Examples

```
data(vbetaFA)
zlmVbeta <- zlm(~ Stim.Condition+Population, subset(vbetaFA, ncells==1)[1:10,])
#Coefficients and standard errors
coef(zlmVbeta, 'D')
coef(zlmVbeta, 'C')
se.coef(zlmVbeta, 'C')
#Test for a Population effect by dropping the whole term (a 5 degree of freedom test)
lrTest(zlmVbeta, 'Population')
#Test only if the VbetaResponsive cells differ from the baseline group
lrTest(zlmVbeta, CoefficientHypothesis('PopulationVbetaResponsive'))
# Test if there is a difference between CD154+/Unresponsive and CD154-/Unresponsive.
# Note that because we parse the expression
# the columns must be enclosed in backquotes
# to protect the \quote{+} and \quote{-} characters.
lrTest(zlmVbeta, Hypothesis('`PopulationCD154+VbetaUnresponsive` -
        `PopulationCD154-VbetaUnresponsive`'))
waldTest(zlmVbeta, Hypothesis('`PopulationCD154+VbetaUnresponsive` -
        `PopulationCD154-VbetaUnresponsive`'))
```

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