# Package 'DNAshapeR'

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Title High-throughput prediction of DNA shape features
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Description DNAhapeR is an R/BioConductor package for ultra-fast,
     high-throughput predictions of DNA shape features.
     The package allows to predict,
     visualize and encode DNA shape features
     for statistical learning.
License GPL-2
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DNAsh		DNAshapeR package for high-throughput prediction of DNA shape						
	features							

# Description

The main functions in the package are getFasta and getShape. Shape predictions can be additionally plotted with plotShape. See package vignette for examples.

# **Details**

All support questions should be posted to the Bioconductor support site: support.bioconductor. org, using DNAshapeR as a tag.

## Author(s)

Tsu-Pei Chiu and Federico Comoglio

# References

## DNAshapeR reference:

T.-P. Chiu\*, F. Comoglio\*, T. Zhou, L. Yang, R. Paro, and R. Rohs: DNAshapeR: an R/Bioconductor package for DNA shape prediction and feature encoding (2016). Bioinformatics <a href="http://bioinformatics.oxfordjournals.org/content/early/2016/01/09/bioinformatics.btv735">http://bioinformatics.oxfordjournals.org/content/early/2016/01/09/bioinformatics.btv735</a> (\*equal contributor in alphabetic order)

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 ${\tt convertMethFile}$ 

Convert fasta file to methylated file format

# Description

Convert fasta file to methylated file format

# Usage

```
convertMethFile(fastaFileName, methPositionFileName)
```

# **Arguments**

fastaFileName The name of the input fasta format file, including full path to file if it is located outside the current working directory.

methPositionFileName

The name of the input position file indicating the methlation position

#### Value

methFileName fasta file containing methylated Cytosine

# Author(s)

Satyanarayan Rao & Tsu-Pei Chiu

encodeKmerHbond

encode Hbond

# Description

encode Hbond

# Usage

```
encodeKmerHbond (k, dnaStringSet )
```

## **Arguments**

k k-mer sequence dnaStringSet dnaStringSet

# Value

featureVector

				_
Δn	200	NΔK	Ma	rSea
C1	LUU	ノニハ	שויו	ı ocu

Encode k-mer DNA sequence features

#### **Description**

DNAshapeR can be used to generate feature vectors for a user-defined model. The model can be a k-mer sequence. Sequence is encoded in four binary features (i.e., in terms of 1-mers, 0001 for adenine, 0010 for cytosine, 0100 for guanine, and 1000 for thymine) at each nucleotide position (Zhou, et al., 2015). The function permits an encoding of 2-mers and 3-mers (16 and 64 binary features at each position, respectively).

## Usage

```
encodeKMerSeq(k, dnaStringSet)
```

# **Arguments**

k A number indicating k-mer sequence encoding dnaStringSet A DNAStringSet object of the inputted fasta file

#### Value

featureVector A matrix containing encoded features. Sequence feature is represented as binary numbers

#### Author(s)

Tsu-Pei Chiu

# ${\tt encodeNstOrderShape}$

Encode n-st order shape features DNAshapeR can be used to generate feature vectors for a user-defined model. The model can be a shape model. There are four structural parameters including MGW, Roll, ProT and HelT. The second order shape features are product terms of values for the same category of shape features at adjacent positions.

# Description

Encode n-st order shape features DNAshapeR can be used to generate feature vectors for a user-defined model. The model can be a shape model. There are four structural parameters including MGW, Roll, ProT and HelT. The second order shape features are product terms of values for the same category of shape features at adjacent positions.

## Usage

```
encodeNstOrderShape(n, shapeMatrix, shapeType)
```

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#### **Arguments**

n A number indicating n-st order shape encoding shapeMatrix A matrix containing DNAshape prediction result

shapeType A character name of shape (MGW, Roll, ProT, HelT) features

#### Value

featureVector A matrix containing encoded features. shape feature is represented as continuous numbers

#### Author(s)

Tsu-Pei Chiu

encodeSeqShape Encode k-mer DNA sequence and n-th order DNA Shape features

#### **Description**

DNAshapeR can be used to generate feature vectors for a user-defined model. These models can be based on DNA sequence (1-mer, 2-mer, 3-mer) or DNA shape (MGW, Roll, ProT, HelT) features or any combination thereof. Sequence is encoded as four binary features (i.e., 0001 for adenine, 0010 for cytosine, 0100 for guanine, and 1000 for thymine, for encoding of 1-mers) at each nucleotide position (Zhou, et al., 2015). Encoding of 2-mers and 3-mers (16 and 64 binary features at each position, respectively) is also supported. Shape features include first and second order (or higher order) values for the four structural parameters MGW, Roll, ProT and HelT. The second order shape features are product terms of values for the same category of shape features at adjacent positions. The function allows to generate any subset of these features, e.g. a given shape category or first order shape features, and any desired combination of shape and sequence features. Feature encoding returns a feature matrix for a dataset of multiple sequences, in which each sequence generates a concatenated feature vector. The output of this function can be used directly for any statistical machine learning method.

# Usage

encodeSeqShape(fastaFileName, shapeMatrix, featureNames, normalize)

#### **Arguments**

fastaFileName A character name of the input fasta format file, including full path to file if it is

located outside the current working directory.

shapeMatrix A matrix containing DNAshape prediction result

featureNames A vector containing a combination of user-defined sequence and shape parame-

ters. The parameters can be any combination of "k-mer", "n-shape", "n-MGW",

"n-ProT", "n-Roll", "n-HelT" (k, n are integers)

normalize A logical indicating whether to perform normalization. Default to TRUE.

## Value

featureVector A matrix containing encoded features. Sequence features are represented as binary numbers, while shape features are represented as real numbers.

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#### Author(s)

Tsu-Pei Chiu

## **Examples**

```
fn <- system.file("extdata", "CGRsample_short.fa", package = "DNAshapeR")
pred <- getShape(fn)
featureNames <- c("1-shape")
featureVector <- encodeSeqShape(fn, pred, featureNames)</pre>
```

getFasta

Extract fasta sequence given a set of genomic intervals and a reference genome.

#### **Description**

DNAshapeR can predict DNA shape features from custom FASTA files or directly from genomic coordinates in the form of a GRanges object within BioConductor (see <a href="https://bioconductor.org/packages/release/bioc/h">https://bioconductor.org/packages/release/bioc/h</a> for more information).

## Usage

```
getFasta(GR, BSgenome, width = 1e3, filename = 'tmp.fa')
```

## **Arguments**

GR A GRanges object indicating genomic coordinates

BSgenome A BSgenome object indicating the genome of interest width A number indicating a fixed width of sequences

filename The Name of the input fasta format file, including full path to file if it is located

outside the current working directory

## Value

writes a fasta file

# Author(s)

Federico Comoglio

## **Examples**

```
gr <- GRanges(seqnames = c("chrI"),
strand = c("+", "-", "+"),
ranges = IRanges(start = c(100, 200, 300), width = 100))
library(BSgenome.Scerevisiae.UCSC.sacCer3)
getFasta(gr, BSgenome = Scerevisiae, width = 100, filename = "tmp.fa")
fn <- "tmp.fa"
pred <- getShape(fn)</pre>
```

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getShape Predict DNA shape from a FASTA file
----------------------------------------------

#### **Description**

The DNA prediction uses a sliding pentamer window where structural features unique to each of the 512 distinct pentamers define a vector of minor groove width (MGW), Roll, propeller twist (ProT), and helix twist (HelT) at each nucleotide position (Zhou, et al., 2013). MGW and ProT define basepair parameter whereas Roll and HelT represent base pair-step parameters. The values for each DNA shape feature as function of its pentamer sequence were derived from all-atom Monte Carlo simulations where DNA structure is sampled in collective and internal degrees of freedom in combination with explicit counter ions (Zhang, et al., 2014). The Monte Carlo simulations were analyzed with a modified Curves approach (Zhou, et al., 2013). Through data mining, average values for each shape feature were calculated for the on average 44 occurrences of each pentamer in an ensemble of Monte Carlo trajectories for 2,121 DNA fragments of 12-27 base pairs in length. DNAshapeR predicts four DNA shape features, which were observed in various co-crystal structures playing an important role in specific protein-DNA binding. The core prediction algorithm enables ultra-fast, high-throughput predictions of shape features for thousands of genomic sequences and is implemented in C++. Since it is likely that features describing additional structural properties or equivalent features derived from different experimental or computational sources will become available, the package has a flexible modular design that easily allows future expansions. In the latest version, we further added additional 9 DNA shape features beyond our previous set of 4 features, and expanded our available repertoire to a total of 13 features, including 6 inter-base pair or base pair-step parameters (HelT, Rise, Roll, Shift, Slide, and Tilt), 6 intra-base pair or base pair-step parameters (Buckle, Opening, ProT, Shear, Stagger, and Stretch), and MGW.

#### Usage

```
getShape(filename, shapeType = 'Default', parse = TRUE,
methylate = FALSE, methylatedPosFile = NULL)
```

#### **Arguments**

filename The name of input fasta format file, including full path to file if it is located

outside the current working directory.

shapeType A character indicating the shape parameters which can be "MGW", "ProT",

"Roll", "HelT" or "All" (meaning all four shapes)

parse A logical value indicating whether parse the prediction result

methylate A logical value indicating wheter consider methlatation

methylatedPosFile

The name of input postion file indicating methlated position

## Details

# Predict biophysical feature

Our previous work explained protein-DNA binding specificity based on correlations between MGW and electrostatic potential (EP) observed in experimentally available structures (Joshi, et al., 2007). However, A/T and C/G base pairs carry different partial charge distributions in the minor groove (due primarily to the guanine amino group), which will affect minor-groove EP. We developed a high-throughput method to predict minor-groove EP based on data mining of results from solving

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the nonlinear Poisson-Boltzmann calculations (Honig & Nicholls, 1995) on 2,297 DNA structures derived from Monte Carlo simulations. DNAshapeR includes EP as an additional feature.

#### Value

shapeList A List containing shapre prediction result

#### Author(s)

Federico Comoglio & Tsu-Pei Chiu

#### **Examples**

```
fn <- system.file("extdata", "CGRsample.fa", package = "DNAshapeR")
pred <- getShape(fn)</pre>
```

heatShape

Plot heatmap of DNA shape features

#### **Description**

Plot heatmap of DNA shape features

#### Usage

```
heatShape(shapeMatrix, nBins, ordRow = NULL, useRaster = TRUE, ...)
```

#### **Arguments**

shapeMatrix A matrix containing DNAshape prediction results.

nBins An integer specifying the number of equally-sized bins in which shape predic-

tions should be aggregated. Summarized predictions can be visualized by setting

nBins=1.

ordRow A numeric vector (of the same length as the number of rows of shapeMatrix)

defining the permutation of the rows of shapeMatrix to be used for plotting.

Default to NULL, i.e. rows are ordered by coefficients of variation.

useRaster Logical, if TRUE a bitmap raster is used to plot the image instead of polygons

(see ?graphics::image for details).

... Additional parameters to be passed to the image.plot function (see ?fields::image.plot

for details).

## Value

Called for its effects

# Author(s)

Federico Comoglio

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#### **Examples**

```
fn <- system.file("extdata", "CGRsample.fa", package = "DNAshapeR")
pred <- getShape(fn)
library(fields)
heatShape(pred$MGW, 20)</pre>
```

normalize

Min-Max normalization

## **Description**

Min-Max normalization

## Usage

```
normalize(x, max, min)
```

## **Arguments**

x A matrix containing encoded features

max A number maximum number for Min-Max Normalization
min A number minimum number for Min-Max Normalization

#### Value

featureVector A matrix containing encoded features. shape feature is represented as continuous numbers

# Author(s)

Tsu-Pei Chiu

normalizeShape

Normalize n-st order shape features

## **Description**

Normalize n-st order shape features

# Usage

```
normalizeShape(featureVector, thOrder, shapeType, normalize)
```

# Arguments

featureVector A matrix containing encoded features.

thOrder A number indicating n-st order shape encoding

shapeType A character name of shape (MGW, Roll, ProT, HelT) features

normalize A logical indicating whether to perform normalization. Default to TRUE.

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#### Value

featureVector A matrix containing encoded features.

## Author(s)

Tsu-Pei Chiu

plotShape	Plot metaprofiles of DNA shape features

# Description

DNA shape features can be visualized as aggregated line plots (also known as metaprofiles, see Comoglio et al., 2015), heat maps (Yang et al., 2014) and genome browser tracks (Chiu et al., 2014).

# Usage

```
plotShape(shapeMatrix, background = NULL,
colDots = rgb( 0, 0, 1, 0.1),
colDotsBg = rgb( 0, 0, 0, 0.1),
colLine = 'steelblue', colLineBg = 'gray50', cex = 0.5, lwd = 2, ylim, ...)
```

# Arguments

shapeMatrix	A matrix containing DNAshape prediction results
background	A matrix containing DNAshape prediction results for a set of background regions. Default to NULL, i.e. background not provided.
colDots	A character vector specifying the color of the points representing the column mean of shapeMatrix. Default to $rgb(0, 0, 1, 0.1)$ .
colDotsBg	A character vector specifying the color of the points representing the column mean of background. Default to $rgb(0, 0, 0, 0.1)$ .
colLine	A character string giving the color name of line representing the column mean of shapeMatrix. Default to 'steelblue'.
colLineBg	A character string giving the color name of line representing the column mean of background. Default to 'gray50'.
cex	A numerical value giving the amount by which plotting text and symbols should be magnified relative to the default. Default to 0.5.
lwd	A numerical value specifying the line width. Default to 2.
ylim	A numerical vector of size 2 specifying the y-axis plot range.
	Additional parameters to be passed to the R plot function.

## Value

Called for its effects

#### Author(s)

Federico Comoglio

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## **Examples**

```
fn <- system.file("extdata", "CGRsample.fa", package = "DNAshapeR")
pred <- getShape(fn)
plotShape(pred$MGW)
plotShape(pred$ProT)
plotShape(pred$Roll)
plotShape(pred$HelT)</pre>
```

 ${\tt readNonStandardFastaFile}$ 

Read the position fasta file

# Description

Read the position fasta file

# Usage

readNonStandardFastaFile(filename)

## **Arguments**

filename

The name of the input position file indicating the methlation position

## Value

df dataframe

# Author(s)

Satyanarayan Rao & Tsu-Pei Chiu

readShape

Read (parse) DNA shape predictions

# **Description**

Read DNA shape predictions

## Usage

readShape(filename)

# **Arguments**

 $\quad \hbox{filename} \quad$ 

character name of the file containing shape predictions, including full path to file if it is located outside the current working directory.

# Value

shapeMatrix matrix containing the shape prediction result

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## Author(s)

Federico Comoglio & Tsu-Pei Chiu

## **Examples**

```
fn <- system.file("extdata", "CGRsample.fa", package = "DNAshapeR")
pred <- readShape(fn)</pre>
```

trackShape

Plot track view of DNA shape features

# **Description**

Plot track view of DNA shape features

## Usage

```
trackShape( filename, shapeList )
```

# **Arguments**

filename The name of the input fasta format file, including full path to file if it is located

outside the current working directory

shapeList A list containing four DNAshape prediction results

## Value

Called for its effects

#### Note

None.

# Author(s)

Tsu-Pei Chiu

# **Examples**

```
fn2 <- system.file("extdata", "SingleSeqsample.fa", package = "DNAshapeR")
pred2 <- getShape(fn2)
trackShape(fn2, pred2) # Only for single sequence file</pre>
```

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