# Package 'ZygosityPredictor'

January 4, 2025

Type Package

Title Package for prediction of zygosity for variants/genes in NGS data

Version 1.7.0

Date 2023-12-08

**Imports** GenomicAlignments, GenomicRanges, Rsamtools, IRanges, VariantAnnotation, DelayedArray, dplyr, stringr, purrr, tibble, methods, knitr, igraph, readr, stats, magrittr, rlang

License GPL-2

# **Description**

The ZygosityPredictor allows to predict how many copies of a gene are affected by small variants. In addition to the basic calculations of the affected copy number of a variant, the Zygosity-Predictor can integrate the influence of several variants on a gene and ultimately make a statement if and how many wild-type copies of the gene are left. This information proves to be of particular use in the context of translational medicine. For example, in cancer genomes, the Zygosity-Predictor can address whether unmutated copies of tumor-suppressor genes are present. Beyond this, it is possible to make this statement for all genes of an organism. The Zygosity-Predictor was primarily developed to handle SNVs and INDELs (later addressed as small-variants) of somatic and germline origin. In order not to overlook severe effects outside of the small-variant context, it has been extended with the assessment of large scale deletions, which cause losses of whole genes or parts of them.

RoxygenNote 7.2.3

**Encoding UTF-8** 

**biocViews** BiomedicalInformatics, FunctionalPrediction, SomaticMutation, GenePrediction

**Depends** R (>= 4.3.0)

LazyData false

Suggests rmarkdown, testthat, BiocStyle

VignetteBuilder knitr

git\_url https://git.bioconductor.org/packages/ZygosityPredictor

git\_branch devel

2 aff\_germ\_copies

Maintainer Marco Rheinnecker <marco.rheinnecker@dkfz-heidelberg.de>

# **Contents**

	aff_germ_copies
	aff_som_copies
	gene_ov
	GR_GENE_MODEL
	GR_GERM_SMALL_VARS
	GR_HAPLOBLOCKS
	GR_SCNA
	GR_SOM_SMALL_VARS
	predict_per_variant
	predict_zygosity
	ZP_ov
Index	16
aff_	germ_copies calculates how many copies are affected by a germnline small variant

# Description

calculates how many copies are affected by a germnline small variant

# Usage

```
aff_germ_copies(chr, af, tcn, purity, sex, c_normal = NULL, af_normal = 0.5)
```

# Arguments

chr	chromosome of the variant (either format 1,2,,X,Y or chr1,,chrX)
af	Allele-frequency of the variant (numeric value between 0 and 1)
tcn	total-copynumber at position of the variant (numeric value >0)

aff\_som\_copies 3

purity	purity of the sample (numeric value between 0 and 1 indicating the fraction of relevant sample with control/unrelevant tissue)
sex	sex of the sample (character: "male", "female", "m", "f")
c_normal	expected copy number at position of the variant in normal tissue, 1 for gonosomes in male samples, and 2 for male autosomes and all chromosomes in female samples. (The function can also assess the c_normal parameter by itself, but then the following two inputs must be provided: chr and sex)
af_normal	Allele-frequency in normal tissue (numeric value between 0 and 1) 0.5 represents heterozygous variants in diploid genome, 1 would be homozygous. Could be relevant if germline CNVs are present at the position. Then also the c_normal parameter would have to be adjusted.

#### Value

A numeric value indicating the affecting copies for the variant

# **Examples**

```
library(dplyr)
library(purrr)
library(stringr)
aff_germ_copies(af=0.67, tcn=2, purity=0.9, chr="chrX", sex="female")
```

aff\_som\_copies

calculates how many copies are affected by a somatic small variant

#### **Description**

calculates how many copies are affected by a somatic small variant

# Usage

```
aff_som_copies(chr, af, tcn, purity, sex, c_normal = NULL)
```

# Arguments

chr	chromosome of the variant (either format 1,2,,X,Y or chr1,,chrX)
af	Allele-frequency of the variant (numeric value between 0 and 1)
tcn	total-copynumber at position of the variant (numeric value >0)

purity purity of the sample (numeric value between 0 and 1 indicating the fraction of

relevant sample with control/unrelevant tissue)

sex sex of the sample (character: "male", "female", "m", "f")

c\_normal expected copy number at the position of the variant in normal tissue, 1 for gono-

somes in male samples, and 2 for male autosomes and all chromosomes in female samples. (The function can also assess the c\_normal parameter by itself,

but then the following two inputs must be provided: chr and sex)

gene\_ov

#### Value

A numeric value indicating the affecting copies for the variant

#### **Examples**

```
library(dplyr)
library(purrr)
library(stringr)
aff_som_copies(chr="chrX", af=0.67, tcn=2, purity=0.9, sex="female")
```

gene\_ov

accesor for gene predictions printing detailed info about how a gene status was assigned

# **Description**

accesor for gene predictions printing detailed info about how a gene status was assigned

# Usage

```
gene_ov(fp, inp_gene, n = 20)
```

### **Arguments**

fp full prediction (output of predict\_zygoisty())
inp\_gene name of gene that should be printed with detailed information
n max number of rows to print, as some gene status depend on loads of phasing results#'

#### Value

prints overview about run from function predict\_zygoisty() with specific information about provided gene

```
cnvs = GenomicRanges::GRanges(
    dplyr::tibble(
        chr = "chr17",
        start = c(170060, 34520990),
        end = c(34520990, 83198614),
        tcn = c(2, 1),
        cna_type = c("neutral", "LOH")
    )
)
somatic_vars = GenomicRanges::GRanges(
    dplyr::tibble(
        chr="chr17",
```

GR\_GENE\_MODEL 5

```
start = 7675088,
    end = 7675088,
    ref = "C",
    alt = T,
    af = 0.65,
    gene = "TP53"
  )
)
germline_vars = GenomicRanges::GRanges(
  dplyr::tibble(
    chr="chr17",
    start = 41771694,
    end = 41771694,
    ref = "GTGT",
    alt = "G",
    af = 0.95,
    gene = "JUP"
  )
)
reference = GenomicRanges::GRanges(
  dplyr::tibble(
    chr = "chr17",
    start = c(7661778, 41754603),
    end = c(7687538, 41786931),
    gene = c("TP53", "JUP")
  )
)
sex = "female"
purity = 0.9
bamfile <- system.file("extdata", "ZP_example.bam",</pre>
  package = "ZygosityPredictor")
fp <- predict_zygosity(purity = purity, sex = sex,</pre>
  somCna = cnvs,
  somSmallVars = somatic_vars,
  germSmallVars = germline_vars,
  geneModel = reference,
  bamDna = bamfile
)
gene_ov(fp, TP53)
```

GR\_GENE\_MODEL

germline small variant object

# **Description**

germline small variant object

#### Usage

```
data(GR_GENE_MODEL)
```

6 GR\_HAPLOBLOCKS

#### **Format**

```
## 'GR_GENE_MODEL' GRanges object
```

#### Value

Object containing gene model of hg38

GR\_GERM\_SMALL\_VARS

germline small variant object

# Description

germline small variant object

# Usage

```
data(GR_SOM_SMALL_VARS)
```

#### **Format**

```
## 'GR_SOM_SMALL_VARS' GRanges object
```

#### Value

Object containing germline Indels and SNVs of SeqC2 example case

GR\_HAPLOBLOCKS

haploblocks

# Description

haploblocks

# Usage

data(GR\_HAPLOBLOCKS)

# **Format**

```
## 'GR_HAPLOBLOCKS' GRanges object
```

#### Value

Object containing haploblock annotations

GR\_SCNA 7

GR\_SCNA

copynumber object

# Description

copynumber object

# Usage

data(GR\_SCNA)

# **Format**

## 'GR\_SCNA' GRanges object

# Value

Object containing somatic copy number abberations (sCNAs) of SeqC2 example case

GR\_SOM\_SMALL\_VARS

somatic small variant object

# Description

somatic small variant object

# Usage

```
data(GR_GERM_SMALL_VARS)
```

#### **Format**

```
## 'GR_GERM_SMALL_VARS' GRanges object
```

# Value

Object containing somatic Indels and SNVs of SeqC2 example case

8 predict\_per\_variant

#### **Description**

predicts zygosity of a set of variants

#### Usage

```
predict_per_variant(
  purity,
  sex,
  somCna,
  geneModel = NULL,
  somSmallVars = NULL,
  germSmallVars = NULL,
  ploidy = NULL,
  colnameTcn = NULL,
  colnameCnaType = NULL,
  includeHomoDel = TRUE,
  includeIncompleteDel = TRUE,
  assumeSomCnaGaps = FALSE,
  byTcn = TRUE,
  ZP_{env} = NULL
  verbose = FALSE
)
```

#### **Arguments**

purity	nurity of the s	sample (numeri	c value between	0 and 1	indicating the fraction of
purity	purity of the s	sampic (mumeri	c value between	o and i	mulcating the fraction of

relevant sample with control/unrelevant tissue)

sex sex of the sample (character: "male", "female", "m", "f")

somCna GRanges object containing all genomic regions with annotated total copynum-

ber and cna\_type as metadata columns. The total-copynumber column should be named "tcn" but also some other commonly used names. It should contain numeric values or characters that can be converted to numeric values. The cna\_type column must contain the information about loss of heterozygosity (LOH). Therefore the term "LOH" must be explicitly mentioned in the column. If a genomic region is not present in the object, it will be taken as heterozygous

with neutral TCN of 2.

geneModel GRanges object containing the gene-annoattion of the used reference genome

with metadata column of the gene name (gene)

somSmallVars GRanges object containing all somatic small variants (SNV and INDEL). Re-

quired metadata columns are reference base (ref/REF), alternative base (alt/ALT), annotation of the gene name (gene/GENE) and the allele-frequency (af/AF). If the object is not provided the tool assumes there are no somatic small variants.

predict\_per\_variant 9

germSmallVars GRanges object containing all germline small variants (SNV and INDEL). Re-

quired metadata columns are reference base (ref/REF), alternative base (alt/ALT), annotation of the gene name (gene/GENE) and the allele-frequency (af/AF) If the object is not provided the tool assumes there are no germline small variants.

ploidy ploidy of the sample (numeric value)

colnameTcn character indicating the name of the metadata containing the tcn information in

the somCna object. If not provided the tool tries to detect the column according

to default names

colnameCnaType character indicating the name of the metadata containing cna type information in

the somCna object. If not provided the tool tries to detect the column according

to default names

includeHomoDel default = TRUE; if FALSE homozygous deleteions are excluded includeIncompleteDel

default = TRUE; if FALSE heterzygous deleteions are excluded

assumeSomCnaGaps

(logical, default=FALSE) Only required if the somCna object lacks copy number information for genomic segments on which small variants are detected. By default, variants in such regions will be excluded from the analysis as required information about the copy number is missing. These variants will be attached to the final output list in a separate tibble. To include them, this flag must be set TRUE and the ground ploidy must be given as an input. This ground ploidy will then be taken as ten in the missing regions. If no ploidy is given the tool will assume the ground ploidy of 2 when this flag is TRUE.

assume the ground ploidy of 2 when this flag is TRUE.

byTcn logical, default=TRUE; optional if includeHomoDel or includeIncompleteDelS

is TRUE. If FALSE the tool will not use tcn as a criterion to assign large deletions. It will use the cna\_type column and check for indicating strings like HOMDEL/HomoDel/DEL. Some commonly used strings are covered. It is rec-

ommended to leave this flag TRUE

ZP\_env internal variable... not recommened to be changed by user verbose logical, default=FALSE; prints functions that are called

#### Value

A list containing tibbles with all input variants

```
cnvs = GenomicRanges::GRanges(
    dplyr::tibble(
        chr = "chr17",
        start = c(170060, 34520990),
        end = c(34520990, 83198614),
        tcn = c(2, 1),
        cna_type = c("neutral", "LOH")
    )
)
somatic_vars = GenomicRanges::GRanges(
```

```
dplyr::tibble(
   chr="chr17",
   start = 7675088,
   end = 7675088,
   ref = "C",
   alt = T,
   af = 0.65,
   gene = "TP53"
  )
)
germline_vars = GenomicRanges::GRanges(
  dplyr::tibble(
   chr="chr17",
   start = 41771694,
   end = 41771694,
   ref = "GTGT",
   alt = "G",
   af = 0.95,
   gene = "JUP"
  )
)
reference = GenomicRanges::GRanges(
  dplyr::tibble(
   chr = "chr17"
   start = c(7661778, 41754603),
   end = c(7687538, 41786931),
   gene = c("TP53", "JUP")
  )
)
sex = "female"
purity = 0.9
predict_per_variant(purity = purity, sex = sex,
  somCna = cnvs,
  somSmallVars = somatic_vars,
  germSmallVars = germline_vars,
  geneModel = reference
)
```

predict\_zygosity

predicts zygosity of a set of genes of a sample

# Description

predicts zygosity of a set of genes of a sample

# Usage

```
predict_zygosity(
  purity,
  sex,
```

```
somCna,
  geneModel,
  bamDna,
  somSmallVars = NULL,
  germSmallVars = NULL,
  bamRna = NULL,
  ploidy = NULL,
  colnameTcn = NULL,
  colnameCnaType = NULL,
  includeHomoDel = TRUE,
  includeIncompleteDel = TRUE,
  showReadDetail = FALSE,
  printLog = FALSE,
  assumeSomCnaGaps = FALSE,
  byTcn = TRUE,
  vcf = NULL,
  haploBlocks = NULL,
  distCutOff = 5000,
  verbose = FALSE,
  debug = FALSE,
  logDir = NULL,
  snpQualityCutOff = 1,
  phasingMode = "fast",
  AllelicImbalancePhasing = FALSE
)
```

# **Arguments**

purity

purity of the sample (numeric value between 0 and 1 indicating the fraction of

relevant sample with control/unrelevant tissue)

sex

sex of the sample (character: "male", "female", "m", "f")

somCna

GRanges object containing all genomic regions with annotated total copynumber and cna\_type as metadata columns. The total-copynumber column should be named "tcn" but also some other commonly used names. It should contain numeric values or characters that can be converted to numeric values. The cna\_type column must contain the information about loss of heterozygosity (LOH). Therefore the term "LOH" must be explicitly mentioned in the column. If a genomic region is not present in the object, it will be taken as heterozygous

with neutral TCN of 2.

geneModel

GRanges object containing the gene-annoattion of the used reference genome with metadata column of the gene name (gene)

bamDna

path to bam-file

somSmallVars

GRanges object containing all somatic small variants (SNV and INDEL). Required metadata columns are reference base (ref/REF), alternative base (alt/ALT), annotation of the gene name (gene/GENE) and the allele-frequency (af/AF). If the object is not provided the tool assumes there are no somatic small variants.

germSmallVars GRanges object containing all germline small variants (SNV and INDEL). Re-

quired metadata columns are reference base (ref/REF), alternative base (alt/ALT), annotation of the gene name (gene/GENE) and the allele-frequency (af/AF) If the object is not provided the tool assumes there are no germline small variants.

bamRna optional; path to rna file (bam format)
ploidy ploidy of the sample (numeric value)

colnameTcn character indicating the name of the metadata containing the tcn information in

the somCna object. If not provided the tool tries to detect the column according

to default names

colnameCnaType character indicating the name of the metadata containing cna type information in

the somCna object. If not provided the tool tries to detect the column according

to default names

includeHomoDel default = TRUE; if FALSE homozygous deleteions are excluded

includeIncompleteDel

default = TRUE; if FALSE heterzygous deleteions are excluded

showReadDetail default = FALSE; if TRUE a table is added to the output, containing all used

reads/rea-pairs with annuated read classification (mut1, mut2, both, none, skipped,

dev\_var)

printLog default = FALSE; if TRUE the gene which is evaluated is printed in console,

containing the query-name of each read which was used to perform haplotype-

phasing and the info into which class it was assigned.

assumeSomCnaGaps

(logical, default=FALSE) Only required if the somCna object lacks copy number information for genomic segments on which small variants are detected. By default, variants in such regions will be excluded from the analysis as required information about the copy number is missing. These variants will be attached to the final output list in a separate tibble. To include them, this flag must be set TRUE and the ground ploidy must be given as an input. This ground ploidy will then be taken as ten in the missing regions. If no ploidy is given the tool will

assume the ground ploidy of 2 when this flag is TRUE.

byTcn logical, default=TRUE; optional if includeHomoDel or includeIncompleteDelS

is TRUE. If FALSE the tool will not use ton as a criterion to assign large deletions. It will use the cna\_type column and check for indicating strings like HOMDEL/HomoDel/DEL. Some commonly used strings are covered. It is rec-

ommended to leave this flag TRUE

vcf character; path to variant call file (.vcf.gz format). Will be used (if provided) for

extended SNP phasing if variants on the same gene are too far away from each

other for direct haplotype phasing

haploBlocks GRanges object containing haploblocks. Haploblocks are defined as genomic

regions in which SNPs are phased to a specific allele. For example a haploblock could be chr1:1000-10000. This would mean that every genotype annotation in the format "110" or "011" of a SNP in this region will be used to phase somatic

variants and define their genotype

distCutOff numeric, default=5000; if input vcf is provided and SNP phasing is performed,

this will limt the distance at which the SNP phasing should not be tried anymore.

As the probability of finding overlapping reads at such a long distance is very

low and the runtime will increase exponentially.

verbose logical, default=FALSE; prints functions that are called debug logical, default=FALSE; prints output for debugging

logDir character; path to directory where logfiles and detailed infos of the run can be

stored, if not given, no details will be stored or printed

snpQualityCutOff

numeric, default=1; Cutoff to filter for SNPS that can be used for phasing

phasingMode character, default="fast"; if set to full. Even if high confidence phasing result

could be achieved, following phasing approaches will be carried out

AllelicImbalancePhasing

logical, default=FALSE. Enables alleleic imbalance phasing if TRUE

#### Value

A list of dataframes. Those are the evaluation per variant, the evaluation per gene and, if performed, the info about the haplotype-phasing.

```
cnvs = GenomicRanges::GRanges(
  dplyr::tibble(
   chr = "chr17",
    start = c(170060, 34520990),
    end = c(34520990, 83198614),
    tcn = c(2, 1),
    cna_type = c("neutral", "LOH")
  )
)
somatic_vars = GenomicRanges::GRanges(
  dplyr::tibble(
    chr="chr17",
    start = 7675088,
    end = 7675088,
    ref = "C",
   alt = T,
   af = 0.65,
    gene = "TP53"
  )
germline_vars = GenomicRanges::GRanges(
  dplyr::tibble(
   chr="chr17",
    start = 41771694,
    end = 41771694,
    ref = "GTGT",
   alt = "G",
   af = 0.95,
    gene = "JUP"
  )
```

 $ZP_{ov}$ 

```
reference = GenomicRanges::GRanges(
  dplyr::tibble(
   chr = "chr17",
   start = c(7661778, 41754603),
   end = c(7687538, 41786931),
   gene = c("TP53", "JUP")
  )
)
sex = "female"
purity = 0.9
bamfile <- system.file("extdata", "ZP_example.bam",</pre>
  package = "ZygosityPredictor")
predict_zygosity(purity = purity, sex = sex,
  somCna = cnvs,
  somSmallVars = somatic_vars,
  germSmallVars = germline_vars,
  geneModel = reference,
  bamDna = bamfile
)
```

ZP\_ov

accesor for ZygoistyPredictor runs. Prints an overview about the run

# **Description**

accesor for ZygoistyPredictor runs. Prints an overview about the run

### Usage

```
ZP_ov(fp)
```

#### **Arguments**

fp

full prediction (output of predict\_zygoisty())

# Value

prints overview about run from function predict\_zygoisty()

```
cnvs = GenomicRanges::GRanges(
    dplyr::tibble(
        chr = "chr17",
        start = c(170060, 34520990),
        end = c(34520990, 83198614),
        tcn = c(2, 1),
        cna_type = c("neutral", "LOH")
    )
```

ZP\_ov 15

```
somatic_vars = GenomicRanges::GRanges(
  dplyr::tibble(
   chr="chr17",
   start = 7675088,
   end = 7675088,
   ref = "C",
   alt = T,
   af = 0.65,
   gene = "TP53"
  )
)
germline_vars = GenomicRanges::GRanges(
  dplyr::tibble(
   chr="chr17",
   start = 41771694,
   end = 41771694,
   ref = "GTGT",
   alt = "G",
   af = 0.95,
   gene = "JUP"
  )
)
reference = GenomicRanges::GRanges(
  dplyr::tibble(
   chr = "chr17",
   start = c(7661778, 41754603),
   end = c(7687538, 41786931),
   gene = c("TP53", "JUP")
  )
)
sex = "female"
purity = 0.9
bamfile <- system.file("extdata", "ZP_example.bam",</pre>
  package = "ZygosityPredictor")
fp <- predict_zygosity(purity = purity, sex = sex,</pre>
  somCna = cnvs,
  somSmallVars = somatic_vars,
  germSmallVars = germline_vars,
  geneModel = reference,
  bamDna = bamfile
ZP_ov(fp)
```

# **Index**

```
\ast datasets
    GR_GENE_MODEL, 5
    GR_GERM_SMALL_VARS, 6
    GR_HAPLOBLOCKS, 6
    GR_SCNA, 7
    GR_SOM_SMALL_VARS, 7
aff_germ_copies, 2
aff\_som\_copies, 3
gene_ov, 4
GR_GENE_MODEL, 5
GR_GERM_SMALL_VARS, 6
GR_HAPLOBLOCKS, 6
GR_SCNA, 7
GR_SOM_SMALL_VARS, 7
predict_per_variant, 8
predict_zygosity, 10
ZP\_ov, 14
```