

# iChip

October 5, 2010

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`enrichreg`                      *Call and merge enriched probes to enriched regions.*

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## Description

A function used to call and merge enriched probes to enriched regions using the posterior probability calculated by `iChip2` or `iChip1` functions at certain posterior probability and false discovery rate (FDR) cutoffs.

## Usage

```
enrichreg(pos, enrich, pp, cutoff, method=c("ppcut", "fdrcut"), maxgap=500)
```

## Arguments

|                     |  |
|---------------------|--|
| <code>pos</code>    | A n by 2 matrix or data frame. Rows correspond to probes. The first column of the matrix contains chromosome IDs; the second column contains the genomic positions.  |
| <code>enrich</code> | A vector containing the probe enrichment measurements.   |
| <code>pp</code>     | A vector containing the posterior probabilities returned by <code>iChip2</code> or <code>iChip1</code> .   |
| <code>cutoff</code> | The cutoff value (a scalar) used to call enriched probes. If use posterior probability as a criterion ( <code>method="ppcut"</code> ), a probe is said to be enriched if its <code>pp</code> is greater than the cutoff. If use FDR as a criterion ( <code>method="fdrcut"</code> ), probes are said to be enriched if the probe-based FDR is less than the cutoff. The FDR is calculated using a direct posterior probability approach (Newton et al., 2004). |
| <code>method</code> | 'ppcut' or 'fdrcut'.   |
| <code>maxgap</code> | The criterion used to merge enriched probes. If the genomic distance of adjacent probes is less than <code>maxgap</code> , the probes will be merged into the same enriched regions.   |

## Value

A data frame with rows corresponding to enriched regions and columns corresponding to the following:

|                     |   |
|---------------------|---|
| <code>chr</code>    | Chromosome IDs. For human genome, 23 and 24 denote X and Y, respectively. |
| <code>gstart</code> | The start genomic position of the enriched region.                        |

|         |  |
|---------|--|
| gend    | The end genomic position of the enriched region.   |
| rstart  | The row number for gstart in the position matrix.  |
| rend    | The row number for gend in the position matrix.  |
| peakpos | The peak genomic position of the enriched region where the probe has the largest enrichment value. |
| meanpp  | The mean posterior probability of the probes in the enriched region.                               |
| maxpp   | The maximum posterior probability of the probes in the enriched region.                            |
| nprobe  | The number of probes in the enriched regions. nprobe = rend - rstart + 1                           |

### Author(s)

Qianxing Mo <moq@mskcc.org>

### References

Qianxing Mo, Faming Liang. (2010). Bayesian modeling of ChIP-chip data through a high-order Ising model. *Biometrics*, 2010 Jan 29 [Epub ahead of print]. DOI: 10.1111/j.1541-0420.2009.01379.x

Qianxing Mo, Faming Liang. (2010). A hidden Ising model for ChIP-chip data analysis. *Bioinformatics* 26(6), 777-783. doi:10.1093/bioinformatics/btq032

Newton, M., Noueiry, A., Sarkar, D., Ahlquist, P. (2004). Detecting differential gene expression with a semiparametric hierarchical mixture method. *Biostatistics* 5 , 155-176.

### See Also

[iChip2](#), [iChip1](#), [lmtstat](#)

### Examples

```
library(iChip)
library(limma)

#Analyze the p53 data (average resolution is about 35 bps)
#uncommenting the following code for running

#data(p53)
#p53lmt = lmtstat(p53[,9:14],p53[,3:8])
#p53Y = cbind(p53[,1],p53lmt)
#p53res=iChip2(Y=p53Y,burnin=2000,sampling=10000,winsize=2,sdcut=2,beta=2.5)
#enrichreg(pos=p53[,1:2],enrich=p53lmt,pp=p53res$pp,cutoff=0.9,
#          method="ppcut",maxgap=500)
#enrichreg(pos=p53[,1:2],enrich=p53lmt,pp=p53res$pp,cutoff=0.01,
#          method="fdrcut",maxgap=500)
```

iChip1

*Bayesian modeling of ChIP-chip data through hidden Ising models***Description**

Function iChip1 implements an algorithm of modeling ChIP-chip data through a standard hidden Ising model.

**Usage**

```
iChip1(enrich, burnin=2000, sampling=10000, sdcut=2, beta0=3,
       minbeta=0, maxbeta=100, normsd=0.1, verbose=FALSE)
```

**Arguments**

|          |  |
|----------|--|
| enrich   | A vector containing the probe enrichment measurements. The measurements must be sorted, firstly by chromosome and then by genomic position. The measurements could be log2 ratios of the intensities of IP-enriched and control samples for a single replicate, or summary statistics such as t-like statistics or mean differences for multiple replicates. We suggest to use the empirical Bayesian t-statistics implemented in the limma package for multiple replicates. Note, binding probes must have a larger mean value than non-binding probes. |
| burnin   | The number of MCMC burn-in iterations.   |
| sampling | The number of MCMC sampling iterations. The posterior probability of binding and non-binding state is calculated based on the samples generated in the sampling period.  |
| sdcut    | A value used to set the initial state for each probe. The enrichment measurements of a enriched probe is typically several standard deviations higher than the global mean enrichment measurements.  |
| beta0    | The initial parameter used to control the strength of interaction between probes, which must be a positive value. A larger value of beta represents a stronger interaction between probes. The value for beta0 could not be too small (e.g. < 1.0). Otherwise, the Ising system may not be able to reach a super-paramagnetic state.   |
| minbeta  | The minimum value of beta allowed.   |
| maxbeta  | The maximum value of beta allowed.   |
| normsd   | iChip1 uses a Metropolis random walk proposal for sampling from the posterior distributions of the model parameters. The proposal distribution is a normal distribution with mean 0 and standard deviation specified by normsd.  |
| verbose  | A logical variable. If TRUE, the number of completed MCMC iterations is reported.  |

**Value**

A list with the following elements.

|    |   |
|----|---|
| pp | The posterior probabilities of probes in the binding/enriched state. There is a strong evidence to be a binding/enriched probe if the probe has a posterior probability close to 1. |
|----|---|

|        |  |
|--------|--|
| beta   | The posterior samples of the interaction parameter of the Ising model.                             |
| mu0    | The posterior samples of the mean measurement of the probes in the non-binding/non-enriched state. |
| mu1    | The posterior samples of the mean measurement of the probes in the binding/enriched state.         |
| lambda | The posterior samples of the precision of the enrichment measurements of the probes.               |

**Author(s)**

Qianxing Mo <moq@mskcc.org>

**References**

Qianxing Mo, Faming Liang. (2010). A hidden Ising model for ChIP-chip data analysis. *Bioinformatics* 26(6), 777-783. doi:10.1093/bioinformatics/btq032

**See Also**

[iChip2](#), [enrichreg](#), [lmtstat](#)

**Examples**

```
# oct4 and p53 data are log2 transformed and quantile-normalized intensities
# Analyze the Oct4 data (average resolution is about 280 bps)
data(oct4)

### sort oct4 data, first by chromosome then by genomic position
oct4 = oct4[order(oct4[,1],oct4[,2]),]

# calculate the enrichment measurements --- the limma t-statistics
oct4lmt = lmtstat(oct4[,5:6],oct4[,3:4])

# Apply the standard Ising model to the ChIP-chip data
oct4res = iChip1(enrich=oct4lmt,burnin=2000,sampling=10000,sdcut=2,
  beta0=3,minbeta=0,maxbeta=100,normsd=0.1)

# check the enriched regions detected by the Ising model using
# posterior probability (pp) cutoff at 0.9 or FDR cutoff at 0.01
enrichreg(pos=oct4[,1:2],enrich=oct4lmt,pp=oct4res$pp,cutoff=0.9,
  method="ppcut",maxgap=500)
enrichreg(pos=oct4[,1:2],enrich=oct4lmt,pp=oct4res$pp,cutoff=0.01,
  method="fdrcut",maxgap=500)

# Analyze the p53 data (average resolution is about 35 bps)
# uncommenting the following code for running

# data(p53)
# must sort the data first
```

```
# p53 = p53[order(p53[,1],p53[,2]),]
# p53lmt = lmtstat(p53[,9:14],p53[,3:8])
# p53res = iChip1(p53lmt,burnin=2000,sampling=10000,sdcut=2,beta0=3,
#               minbeta=0,maxbeta=100,normsd=0.1)

# enrichreg(pos=p53[,1:2],enrich=p53lmt,pp=p53res$pp,cutoff=0.9,
#           method="ppcut",maxgap=500)
# enrichreg(pos=p53[,1:2],enrich=p53lmt,pp=p53res$pp,cutoff=0.01,
#           method="fdrcut",maxgap=500)
```

---

iChip2

*Bayesian modeling of ChIP-chip data through hidden Ising models*


---

### Description

Function iChip2 implements a method of modeling ChIP-chip data through a high-order hidden Ising model.

### Usage

```
iChip2(Y,burnin=2000,sampling=10000,winsize=2,sdcut=2,beta=2.5,verbose=FALSE)
```

### Arguments

|          |  |
|----------|--|
| Y        | A n by 2 matrix or data frame. The first column of Y contains the chromosome IDs; the second column of Y contains the probe enrichment measurements. Y must be sorted, firstly by chromosome and then by genomic position. The probe enrichment measurements could be log <sub>2</sub> ratios of the intensities of IP-enriched and control samples for a single replicate, or summary statistics such as t-like statistics or mean differences for multiple replicates. We suggest to use the empirical Bayesian t-statistics implemented in the limma package for multiple replicates. Note, binding probes must have a larger mean value than non-binding probes. |
| burnin   | The number of MCMC burn-in iterations.   |
| sampling | The number of MCMC sampling iterations. The posterior probability of binding and non-binding state is calculated based on the samples generated in the sampling period.  |
| winsize  | The parameter to control the order of interactions between probes. For example, winsize = 2, means that probe i interacts with probes i-2,i-1,i+1 and i+2. A balance between high sensitivity and low FDR could be achieved by setting winsize = 2.  |
| sdcut    | A value used to set the initial state for each probe. The enrichment measurements of a enriched probe is typically several standard deviations higher than the global mean enrichment measurements.  |
| beta     | The parameter used to control the strength of interaction between probes, which must be a positive value. A larger value of beta represents a stronger interaction between probes. In general, high resolution array such as Affymetrix tiling arrays have relatively stronger probe interactions than low resolution array such as Agilent tiling arrays. For the second order Ising model (winsize = 2), the   |

critical value of beta is around 1.0. For low resolution array data (e.g. 280 bp resolution), beta could be set to close to the critical value; For high resolution array data (e.g. 35 bp resolution), beta could be set to a value between 2 to 4. In general, choosing a large value of beta amounts to using a more stringent criterion for detecting enriched regions in ChIP-chip experiments.

`verbose` A logical variable. If TRUE, the number of completed MCMC iterations is reported.

### Value

A list with the following elements.

|                      |   |
|----------------------|---|
| <code>pp</code>      | The posterior probabilities of probes in the binding/enriched state. There is a strong evidence to be a binding/enriched probe if the probe has a posterior probability close to 1. |
| <code>mu0</code>     | The posterior samples of the mean measurement of the probes in the non-binding/non-enriched state.  |
| <code>mu1</code>     | The posterior samples of the mean measurement of the probes in the binding/enriched state.  |
| <code>lambda0</code> | The posterior samples of the precision of the enrichment measurements of the probes in the non-binding/non-enriched state.  |
| <code>lambda1</code> | The posterior samples of the precision of the enrichment measurements of the probes in the binding/enriched state.  |

### Author(s)

Qianxing Mo <moq@mskcc.org>

### References

Qianxing Mo, Faming Liang. (2010). Bayesian modeling of ChIP-chip data through a high-order Ising model. *Biometrics*, 2010 Jan 29 [Epub ahead of print]. DOI: 10.1111/j.1541-0420.2009.01379.x

### See Also

[iChip1](#), [enrichreg](#), [lmtstat](#)

### Examples

```
# oct4 and p53 data are log2 transformed and quantile-normalized intensities
# Analyze the Oct4 data (average resolution is about 280 bps)

data(oct4)

### sort oct4 data, first by chromosome then by genomic position
oct4 = oct4[order(oct4[,1],oct4[,2]),]

# calculate the enrichment measurements --- the limma t-statistics

oct4lmt = lmtstat(oct4[,5:6],oct4[,3:4])

# prepare the data used for the Ising model
```

```

oct4Y = cbind(oct4[,1],oct4lmt)

# Apply the second-order Ising model to the ChIP-chip data

oct4res=iChip2(Y=oct4Y,burnin=2000,sampling=10000,winsize=2,sdcut=2,beta=1.25)

# check the enriched regions detected by the Ising model using
# posterior probability (pp) cutoff at 0.9 or FDR cutoff at 0.01

enrichreg(pos=oct4[,1:2],enrich=oct4lmt,pp=oct4res$pp,cutoff=0.9,
          method="ppcut",maxgap=500)
enrichreg(pos=oct4[,1:2],enrich=oct4lmt,pp=oct4res$pp,cutoff=0.01,
          method="fdrcut",maxgap=500)

# Analyze the p53 data (average resolution is about 35 bps)
# uncommenting the following code for running

# data(p53)
# must sort the data first
# p53 = p53[order(p53[,1],p53[,2]),]
# p53lmt = lmtstat(p53[,9:14],p53[,3:8])
# p53Y = cbind(p53[,1],p53lmt)
# p53res=iChip2(Y=p53Y,burnin=2000,sampling=10000,winsize=2,sdcut=2,beta=2.5)

# enrichreg(pos=p53[,1:2],enrich=p53lmt,pp=p53res$pp,cutoff=0.9,
#           method="ppcut",maxgap=500)
# enrichreg(pos=p53[,1:2],enrich=p53lmt,pp=p53res$pp,cutoff=0.01,
#           method="fdrcut",maxgap=500)

```

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lmtstat

*A wrapper function used to calculate the limma t-statistics*


---

## Description

A wrapper function used to calculate the empirical Bayes t-statistics (limma t-statistics) using functions in the limma package.

## Usage

```
lmtstat(IP, CON)
```

## Arguments

IP Data matrix for IP-enriched samples, where the rows and columns correspond to the probes and sample replicates, respectively. The number of replicates must be greater than one. If CON is missing, IP is assumed to be in log-ratio format (e.g.  $\log_2(\text{IP-enriched}/\text{control})$ ). In this case, paired t-statistics are calculated. If CON is NOT missing, IP and CON are assumed to be the normalized intensities for the IP-enriched and control samples, respectively. In this case, two-sample t-statistics are calculated.

CON Data matrix for control samples, where the rows and columns correspond to the probes and sample replicates, respectively. The number of replicates must be greater than one.

**Value**

Empirical Bayes t-statistics calculated using functions in the limma package.

**Author(s)**

Qianxing Mo <moq@mskcc.org>

**References**

Smyth, G. K. (2004). Linear models and empirical Bayes methods for assessing differential expression in microarray experiments. *Statistical Applications in Genetics and Molecular Biology* 3, No. 1, Article 3.

**See Also**

[enrichreg](#), [iChip2](#), [iChip1](#)

**Examples**

```
library(limma)

# load the log2 transformed and quantile-normalized Oct4 data
data(oct4)
oct4[1:3,]

# calculate the enrichment measurements --- two-sample limma t-statistics
oct4lmt1 = lmtstat(oct4[,5:6],oct4[,3:4])

# calculate paired limma t-statistics for the data that are in
# the log-ratio format (e.g., log2(IP-enriched/control))

oct4log2r = oct4[,5:6] - oct4[,3:4]
oct4lmt2 = lmtstat(oct4log2r)
```

---

oct4

*Oct4 data*

---

**Description**

This is a subset of the Oct4 data containing 12584 probes on chromosome 20. The data were log2 transformed and quantile-normalized.

**Usage**

```
data(oct4)
```

**Source**

[http://jura.wi.mit.edu/young\\\_public/hESregulation/Data\\\_download.html](http://jura.wi.mit.edu/young\_public/hESregulation/Data\_download.html)

**References**

Boyer LA, Lee TI, Cole MF, et al. (2005). Core transcriptional regulatory circuitry in human embryonic stem cells. *Cell*, 122(6), 828-30.

---

p53

*p53 data*

---

**Description**

This is a subset of the p53 data containing 10000 probes on chromosome 22. The data were log2 transformed and quantile-normalized.

**Usage**

data (p53)

**Source**

[http://www.gingeras.org/affy\\\_archive\\\_data/publication/tfbs/](http://www.gingeras.org/affy\_archive\_data/publication/tfbs/)

**References**

Cawley S, Bekiranov S, Ng HH, et al. (2004). Unbiased mapping of transcription factor binding sites along human chromosomes 21 and 22 points to widespread regulation of noncoding RNAs. *Cell*, 116(4), 499-509.

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