

# globaltest

April 19, 2009

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GOstructure

*Class "GOstructure" to serve as input for the function gtGO*

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## Description

The class `GOstructure` stores the Gene Ontology information that can be used by the `gtGO` function to find a GO subgraph significantly associated with a response variable. Objects of type `GOstructure` are typically made with a call to `makeGOstructure`.

## Slots

**ids:** Object of class "character". Vector of GO identifiers.

**offspring:** A named list of vectors of GO identifiers. Lists all offspring terms of each term.

**ancestors:** A named list of vectors of GO identifiers. Lists all ancestor terms of each term.

**genesets:** A named list of vectors of gene identifiers. Lists the genes annotated to each GO id.

## Methods

**show** (`GOstructure`): Summarizes the object.

**length** (`GOstructure`): Gives the number of GO ids in the object.

**genesets** (`GOstructure`): Extracts the genesets slot.

## Author(s)

Jelle Goeman: (j.j.goeman@lumc.nl); Jan Oosting

## See Also

`gtGO`, `makeGOstructure`.

---

```
annotation.vandeVijver
```

*Some selected annotations for the breast cancer data from Rosetta Inpharmatics LLC and Netherlands Cancer Institute, to serve as example data in the globaltest package.*

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### Description

A named list of 64 vectors of probe identifiers appearing in the [vandeVijver](#), linking the probes to annotation terms from the Gene Ontology (GO) database. All 64 GO terms are descendants of the GO term Cell Cycle (GO:0007049).

### Usage

```
data(annotation.vandeVijver)
```

### Format

A named list of vectors.

### See Also

[vandeVijver](#).

---

```
checkerboard
```

*Checkerboard plot for Global Test*

---

### Description

Produces a plot to visualize the test result produced by [globaltest](#), by showing the association between pairs of samples.

### Usage

```
checkerboard(gt, geneset, sort = TRUE, drawlabels = TRUE,
  labelsize = 0.6, ...)
```

### Arguments

gt	The output of a call to <a href="#">globaltest</a> .
geneset	The name or number of the geneset to be plotted (only necessary if multiple genesets were tested)
sort	A logical flag to indicate whether the samples should be sorted by the clinical outcome to give a clearer picture.
drawlabels	Logical value to control drawing of the samplenames on the x- and y-axis of the plot.
labelsize	Relative size of the labels on the x- and y-axis. If it is NULL, the current value for <code>HYPERLINK(par("cex.axis"))(par("cex.axis"))</code> is used
...	Any extra arguments will be forwarded to the plotting function.

## Details

The checkerboard shows the pairs of samples which have high covariance in white and the pairs with low covariance in black. This can be used to visualize the data and to search for outlying arrays.

The left and bottom margins are adjusted to allow enough space for the longest samplename.

## Value

A matrix giving the old and the new sample numbers.

## Author(s)

Jelle Goeman: <j.j.goeman@lumc.nl>; Jan Oosting

## References

J. J. Goeman, S. A. van de Geer, F. de Kort and J. C. van Houwelingen, 2004, *A global test for groups of genes: testing association with a clinical outcome*, *Bioinformatics* 20 (1) 93–99. See also the *How To Globaltest.pdf* included with this package.

## See Also

[globaltest](#), [sampleplot](#), [geneplot](#).

## Examples

```
# Breast cancer data (ExpressionSet) from the Netherlands Cancer
# Institute with annotation:
data(vandeVijver)
data(annotation.vandeVijver)
aPathway <- annotation.vandeVijver[1]

gt <- globaltest(vandeVijver, "ESR1", aPathway)

if (interactive()){
  checkerboard(gt[1])
}
```

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geneplot

*Gene Plot for Global Test*

---

## Description

Produces a plot to show the influence of individual genes on the test result produced by [globaltest](#).

## Usage

```
geneplot(gt, geneset, genesubset, scale = FALSE, drawlabels = TRUE,
  labelsizesize = 0.6, plot = TRUE, addlegend = TRUE, ...)
```

**Arguments**

<code>gt</code>	The output of a call to <code>globaltest</code> .
<code>geneset</code>	The name or number of the geneset to be plotted (only necessary if multiple genesets were tested).
<code>genesubset</code>	A vector of names or numbers of genes to be plotted (default: plot all genes)
<code>scale</code>	Logical: should the bars be scaled to unit standard deviation?
<code>drawlabels</code>	Logical value to control drawing of gene ids (rownames) on the x-axis of the plot.
<code>labelsize</code>	Relative size of the labels on the x-axis. If it is <code>NULL</code> , the current value for <code>HYPERLINK(par("cex.axis"))(par("cex.axis"))</code> is used
<code>plot</code>	If <code>FALSE</code> : does not plot, but only returns a <code>gt.barplot</code> object.
<code>addlegend</code>	If <code>FALSE</code> : does not add a legend to the plot or to the <code>gt.barplot</code> object.
<code>...</code>	Any extra arguments will be forwarded to the plotting function.

**Details**

The geneplot shows a bar and a reference line for each gene. The bar shows the influence of each gene on the test result: the test statistic  $Q$  is the average of the bars of the genes. The reference line shows the expected influence if the gene was not associated with the outcome. The marks on the bars show the standard deviation of the bar under the null hypothesis. The color of the bar indicates positive or negative correlation of the gene with the clinical outcome, to distinguish between up and downregulation.

The bottom margin is adjusted to allow enough space for the longest gene id to draw under the axis.

**Value**

An object of type `gt.barplot`.

**Author(s)**

Jelle Goeman: `<j.j.goeman@lumc.nl>`; Jan Oosting

**References**

For references, type: `citation("globaltest")`. See also the vignette `GlobalTest.pdf` included with this package.

**See Also**

`globaltest`, `sampleplot`.

**Examples**

```
# Breast cancer data (ExpressionSet) from the Netherlands Cancer
# Institute with annotation:
data(vandevijver)
data(annotation.vandevijver)

gt <- globaltest(vandevijver, "StGallen", annotation.vandevijver)

if (interactive()){
```

```

    geneplot(gt["cell cycle checkpoint"])
  }

gp <- geneplot(gt["cell cycle checkpoint"], plot = FALSE)
if (interactive()){
  plot(sort(gp))
}

```

---

globaltest

*Global Test*


---

## Description

In microarray data, tests a (list of) group(s) of genes for significant association with a given clinical variable.

## Usage

```

globaltest(X, Y, genesets, model,
           levels, d, event = 1, adjust,
           method = c("auto", "asymptotic", "permutations", "gamma"),
           nperm = 10^4, scaleX = TRUE, accuracy = 50, ...)

```

## Arguments

- |          |   |
|----------|---|
| X        | Either a matrix of gene expression data, where columns correspond to samples and rows to genes or a Bioconductor <a href="#">ExpressionSet</a> . The data should be properly normalized beforehand (and log- or otherwise transformed), but missing values are allowed (coded as NA). Gene and sample names can be included as the row and column names of X.   |
| Y        | A vector with the clinical outcome of interest, having one value for each sample. If X is an <a href="#">ExpressionSet</a> it can also be the name of a covariate in the <a href="#">phenoData</a> from the <a href="#">ExpressionSet</a> , or a <a href="#">formula</a> object using these names. If the clinical outcome is survival, Y should contain the survival times.  |
| genesets | Either a vector or a list of vectors. Indicates the group(s) of genes to be tested. Each vector in <code>genesets</code> can be given in three formats. Either it can be a vector with 1 (TRUE) or 0 (FALSE) for each gene in X, with 1 indicating that the gene belongs to the group. Or it can be a vector containing the column numbers (in X) of the genes belonging to the group. Or it can be a subset of the rownames or <a href="#">featureNames</a> for X. |
| model    | Globaltest will try to determine the correct model from the input of Y and d. To override the automatic choice, use <code>model = "logistic"</code> for a two-valued outcome Y, <code>model = "linear"</code> for a continuous outcome and <code>model = "survival"</code> for a survival outcome.  |
| levels   | If Y is a factor (or a category in the <a href="#">PhenoData</a> slot of X) and contains more than 2 levels: <code>levels</code> is a vector of levels of Y to test. If <code>levels</code> is length 2: test these 2 groups against each other. If <code>levels</code> is length 1: test that level against the others.  |

d	A vector or the name of a covariate in the <code>phenoData</code> from the <code>ExpressionSet</code> X, to indicate which samples experienced an event. Providing a value for d automatically sets <code>model = "survival"</code>
event	The value or values of d that indicates that there was an event.
adjust	Confounders or risk factors for which the test must be adjusted. Must be either a data frame or (if X is an <code>ExpressionSet</code> ) the names of covariates in the <code>phenoData</code> from X or a <code>formula</code> object using these names. Default: no adjustment.
method	The method for calculation the p-value. Use <code>codemethod = "asymptotic"</code> for the full asymptotic distribution of the test statistic; <code>method = "gamma"</code> for the gamma (= scaled chi-squared) approximation to that distribution and <code>method = "permutations"</code> for a permutation p-value. The default: <code>method = "auto"</code> chooses the permutations method if the number of possible permutations does not exceed 10,000 and the asymptotic otherwise. Note that <code>method = "gamma"</code> was the default option prior to version 4.0.0.
nperm	A number of permutations. This gives the (maximum) number of permutations to be used if <code>method = "permutations"</code> or <code>method = "auto"</code> .
scaleX	If true, rescales the expression matrix to get pleasant value for all test statistics. The expression matrix X is multiplied by a constant in such a way that the expected value EQ of the test statistic for the global test becomes exactly 10. This rescaling has no effect on the p-values.
accuracy	Numerical tuning parameter useable only with the asymptotic method and a non-survival response. Determines how much small eigenvalues of the R matrix are smoothed away to increase computation speed. Choose smaller values for quicker computations but conservative p-values; choose larger values for slower calculations but more accuracy.
...	Captures deprecated input for compatibility with older versions of globaltest.

## Details

The Global Test tests whether a group of genes (of any size from one single gene to all genes on the array) is significantly associated with a clinical variable. The group could be for example a known pathway, an area on the genome or the set of all genes. The test investigates whether samples with similar clinical outcomes tend to have similar gene expression patterns. For a significant result it is not necessary that the genes in the group have similar expression patterns, only that many of them are correlated with the outcome.

## Value

The function returns an object of class `gt.result`.

## Note

The options `globaltest` options `sampling` and `permutation` have been replaced by separate functions from version 3.0. See [sampling](#) and [permutations](#).

## Author(s)

Jelle Goeman: [j.j.goeman@lumc.nl](mailto:j.j.goeman@lumc.nl); Jan Oosting

## References

For references, type: `citation("globaltest")`. See also the vignette `GlobalTest.pdf` included with this package.

## See Also

Many more examples in the vignette! [geneplot](#), [sampleplot](#), [sampling](#), [gt.multtest](#), [permutations](#), [checkerboard](#), [regressionplot](#).

## Examples

```
# Breast cancer data (ExpressionSet) from the Netherlands Cancer
# Institute with annotation:
data(vandeVijver)
data(annotation.vandeVijver)

# Many possible calls. See the vignette for more examples and explanation.
globaltest(vandeVijver, "StGallen")
globaltest(vandeVijver, "StGallen", annotation.vandeVijver)
globaltest(vandeVijver, "Surv(TIMEsurvival, EVENTdeath)", annotation.vandeVijver)
globaltest(vandeVijver, StGallen ~ Posnodes + StGallen, annotation.vandeVijver)
globaltest(vandeVijver, "StGallen", method = "p")

# Store the test result
# See help(gt.result) for more options
gt <- globaltest(vandeVijver, "StGallen", annotation.vandeVijver)
gt[1:2]
sort(gt)
p.value(gt)

# Also with simple vector/matrix input
X <- matrix(rnorm(3000), 100, 30) # random expression data
Y <- 1:30 # a response variable
pathway <- 1:40 # a pathway

globaltest(X, Y)
globaltest(X, Y, pathway)
```

---

gostructure.vandeVijver

*A GOstructure object for the breast cancer data from Rosetta Inpharmatics LLC and Netherlands Cancer Institute, to serve as example data in the globaltest package.*

---

## Description

A object with information on 64 annotation terms from the Gene Ontology (GO) database, all descendants of the GO term Cell Cycle (GO:0007049). The object contains information on ancestors, descendants and associated genes.

## Usage

```
data(gostructure.vandeVijver)
```

**Format**

A `GOstructure` object.

**See Also**

`GOstructure`, `vandeVijver`.

---

`gt.barplot-class`    *Class "gt.barplot" for results of the functions `geneplot` and `sampleplot`*

---

**Description**

The class `gt.barplot` stores the output of a call to `geneplot` or `sampleplot`.

**Slots**

**res:** Object of class "matrix". The plotted values, with a row for each gene or sample, and the following columns: `influence`: the height of the bar, `Einf`: expected influence under the null hypothesis, `sd.inf`: standard deviation of the influence under the null hypothesis, `up`: the colour of the plotted bar (1=green; 0=red)

**drawlabels:** Object of class "logical". Whether gene or sample labels should be printed at the x-axis.

**labelsize:** Object of class "numeric". The size of the gene/sample labels.

**legend:** The meaning of the colours of the bars, to be used in the legend.

**colour:** The numbers of the colours used in the plot.

**colourCode:** The meaning of the colours used (shorter version of `legend`).

**Methods**

**show** (`gt.barplot`): Summarizes plot in numbers.

**plot** (`gt.barplot`): Redraws the plot. Possible extra arguments: `labelsize` and `drawlabels`.

**"["** (`gt.barplot`): Extracts a subset of the genes or samples.

**result** (`gt.barplot`): Extracts the results matrix as a `data.frame`.

**length** (`gt.barplot`): The number of bars (genes or samples) in the object.

**scale** (`gt.barplot`): Scales the bars to unit standard deviation.

**z.score** (`gt.barplot`): Calculates the z.scores for each bar:  $(\text{influence} - \text{expected inf}) / \text{sd}(\text{inf})$ .

**sort** (`gt.barplot`): Sorts the bars to a decreasing z.score.

**names** (`gt.barplot`): The names of the genes or samples.

**names<-** (`gt.result`): Changes the names of the genes or samples.

**Author(s)**

Jelle Goeman: `{j,j.goeman@lumc.nl}`; Jan Oosting

**See Also**

`sampleplot`, `geneplot`, `globaltest`.



---

gt.result-class      *Class "gt.result" for results of the function globaltest*

---

## Description

The class `gt.result` is the output of a call to `globaltest` and the input of various plotting functions to visualize the test result.

## Slots

- res:** Object of class "matrix". Test results summary.
- ex:** Object of class "matrix". The transformed data matrix.
- genesets:** A list of vectors indicating the tested genes.
- IminH:** Object of class "matrix". Needed when drawing the various diagnostic plots.
- fit:** Object of class "list". This list contains one element which is of class "lm", "glm", "coxph" or "coxph.null". This latter object contains the fitted model of the null hypothesis.
- PermQs:** Object of class "matrix". Stores the permuted test statistics generated by a call to `permutations`.
- SamplingZs:** Object of class "list". Stores the standardized test statistics of random gene sets generated by a call to `sampling`.

## Methods

- show** (gt.result): Summarizes the test result.
- "["** (gt.result): Extracts results of one or more pathways if multiple pathways were tested.
- length** (gt.result): The number of pathways tested.
- size** (gt.result): Extracts a vector with the number of genes tested for each pathway.
- p.value** (gt.result): Extracts the p-values.
- z.score** (gt.result): Extracts z-score  $(Q - EQ) / sd(Q)$ .
- sort** (gt.result): Sorts the pathways to increasing p-values.
- result** (gt.result): Extracts the results matrix.
- names** (gt.result): Extracts the pathway names.
- names<-** (gt.result): Changes the pathway names.
- combine** (gt.result): Combines two `gt.result` objects to one, provided the data and model match.
- fit** (gt.result): Extracts the fitted (adjust)model.
- hist** (gt.result): Produces a histogram to visualize the permutations generated by `permutations`.
- geneplot** (gt.result): The `geneplot` produces a plot to show the influence of individual genes on the test result produced by `globaltest`.
- sampleplot** (gt.result): The `sampleplot` produces a plot to show the influence of individual samples on the test result produced by `globaltest`.
- permutations** (gt.result): The function `permutations` recalculates the p-values using permutations of the outcome Y.
- sampling** (gt.result): The function `sampling` compares the p-values with p-values of randomly generated pathways.

**checkerboard** (gt.result): Produces a plot to visualize the test result produced by `globaltest` by showing the association between pairs of samples.

**regressionplot** (gt.result): Produces a plot which can be used to visualize the effect of specific samples on the test result produced by `globaltest`.

#### Author(s)

Jelle Goeman: (j.j.goeman@lumc.nl); Jan Oosting

#### See Also

`globaltest`, `sampleplot`, `genepplot`, `permutations`.

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Multiple testing on the GO graph

*Multiple testing on the GO graph*

---

#### Description

Three function to test (part of) the GO graph for association of the gene expression profile of GO terms with a response variable. Used together, these functions return multiplicity-adjusted p-values calculated using the Focus Level procedure that preserves the structure of the GO graph.

#### Usage

```
gtGO(..., GO, focus, maxalpha = 0.05, stopafter = 100, verbose = FALSE)
makeGOstructure(data, annotation, top, only.ids, ontology = c("BP", "CC", "MF")
getFocus(GOstructure, maxatoms = 10)
```

#### Arguments

...	Arguments describing the tests to be performed are passed on to <code>globaltest</code> .
GO	An object of class <code>GOstructure</code> describing the structure of the GO graph. This object should be created using <code>makeGOstructure</code>
focus	A vector of GO ids to describe te focus level. Typically made using <code>getFocus</code> .
maxalpha	The maximum multiplicity-adjusted p-value. The algorithm will stop when this value is exceeded.
stopafter	The maximum number of significant GO terms to be found. The algorithm will stop when this value is exceeded.
verbose	If set to TRUE, prints much more extensive progress information.
data	The data set for which the <code>GOstructure</code> object is to be made. Can be either an <code>ExpressionSet</code> or a vector of probe indicators.
annotation	The name of the metaData annotation package to be used. If a metaData package is not available, the name of the organism package (e.g. <code>org.HS.eg.db</code> ) can be given instead, and the mapping from probe identifiers to entrez can be supplied with the <code>entrez</code> argument.
top	The node to be used as top node of the <code>GOstructure</code> . Defaults to one of "GO:0008150", "GO:0005575", "GO:0003674", depending on he ontology chosen. The <code>GOstructure</code> object will only contain offspring of the chosen top node.

<code>only.ids</code>	A vector of GO ids. If this is supplied, GO ids not appearing in this list will not appear in the <code>GOstructure</code> object.
<code>ontology</code>	The ontology to be used. One of Biological Process (BP), Cellular Component (CC) or Molecular Function (MF).
<code>entrez</code>	A vector of EntrezGene ids with the same length as the number of genes in <code>data</code> , (and in the same order) giving the mapping from probe identifiers to EntrezGene ids. If this argument is used, the <code>annotation</code> argument should give the name of the organism annotation package.
<code>unreliable</code>	Can be used to designate one or more of the GO evidence codes as unreliable. Value must be a vector containing one or more from "IC", "IDA", "IEA", "IEP", "IGI", "IMP", "IPI", "ISS", "NAS", "ND", "RCA", "TAS", "NR".
<code>GOstructure</code>	An object of class <code>GOstructure</code> .
<code>maxatoms</code>	A tuning parameter that governs the choice of the focus level. <code>getFocus</code> will choose those GO terms in the focus level in such a way that all offspring gene sets of each focus level term can be constructed as unions of no more than <code>maxatoms</code> atom gene sets. Default value of <code>maxatoms</code> is 10. Higher values quickly lead to slower computation. Lower values typically lead to reduced power.

### Details

These functions should be used in the following order. First use `makeGOstructure` to make a GO graph tailored to a specific data set. Then `getFocus` can be used to choose a focus level. Finally `gtGO` performs the focus level procedure.

### Value

The function returns a named vector of multiplicity-adjusted p-values. Adjusted p-values of GO terms not appearing in this vector are larger than the chosen value of `maxalpha`.

### Note

`gtGO` cannot be used in combination with the permutation version of `globaltest`.

### Author(s)

Jelle Goeman: `<j.j.goeman@lumc.nl>`; Jan Oosting

### References

For references, type: `citation("globaltest")`. See also the vignette `GlobalTest.pdf` included with this package.

### See Also

Examples in the vignette! `globaltest`, `GOstructure`, `gt.multtest`.

## Examples

```
# Breast cancer data (ExpressionSet) from the Netherlands Cancer
# Institute with small GO structure (all descendants of Cell Cycle)
# See the vignette for details on making a GO structure object.
data(vandeVijver)
data(gostructure.vandeVijver)

focus <- getFocus(gostructure.vandeVijver)
adjustedP <- gtGO(vandeVijver, "Surv(TIMESurvival, EVENTdeath)",
  GO = gostructure.vandeVijver, focus = focus)

# The significant GO graph can be visualized using
# the GOstats and Rgraphviz packages.
# See the vignette for details. Type: vignette("GlobalTest")
```

---

mlogit

*Multinomial Logistic Regression*

---

## Description

Fits a multinomial logistic regression model to a nominal scale outcome.

## Usage

```
mlogit(formula, data, control = glm.control())
```

## Arguments

formula	An object of class <code>formula</code> containing a symbolic description of the model to be fit. See the documentation of <code>formula</code> for details.
data	An optional data frame containing the variables in the model. If not found in 'data', the variables are taken from the environment from which 'mlogit' is called.
control	A list of parameters for controlling the fitting process. See the documentation of <code>glm.control</code> for details.

## Details

The function `mlogit` fits a multinomial logistic regression model for a multi-valued outcome with nominal scale. The implementation and behaviour are designed to mimic those of `glm`, but the options are (as yet) more limited. Missing values are not allowed in the data.

The model is fitted without using a reference outcome category; the parameters are made identifiable by the requirement that the sum of corresponding regression coefficients over the outcome categories is zero.

**Value**

An object of (S4) class `mlogit`. The class has slots: `coefficients` (matrix), `standard.err` (matrix), `fitted.values` (matrix), `x` (matrix), `y` (matrix), `formula` (formula), `call` (call), `df.null` (numeric), `df.residual` (numeric), `null.deviance` (numeric), `deviance` (numeric), `iter` (numeric), `converged` (logical).

Methods implemented for the `mlogit` class are `coefficients`, `fitted.values`, `residuals` and which extract the relevant quantities, and `summary`, which gives the same output as with a `glm` object.

**Author(s)**

Jelle Goeman: <j.j.goeman@lumc.nl>; Jan Oosting

**See Also**

`glm`, `multinom`.

**Examples**

```
y <- factor(rep(1:4, 5))
x <- 1:20
fit <- mlogit(y ~ x)
summary(fit)
residuals(fit)
```

---

gt.multtest

*Correct globaltest results for multiple testing*

---

**Description**

Corrects the raw p-values resulting from a call to `globaltest` for multiple testing, using either Benjamini and Hochberg's False Discovery Rate or Holm's procedure for controlling the Family-Wise Error Rate.

**Usage**

```
gt.multtest(gt, proc = c("FDR", "FWER"))
```

**Arguments**

<code>gt</code>	The output of a call to <code>globaltest</code> .
<code>proc</code>	The procedure to be used. Either "FDR" for Benjamini and Hochberg's (1995) False Discovery Rate-controlling procedure or "FWER" for Holm's (1979) Family-Wise Error Rate controlling procedure.

**Details**

This function is completely based on the `mt.rawp2adjp` function from the `multtest` package.

**Value**

An object of class `gt.result`.

**Note**

This function must be called *prior* to any selection of significant genes.

**Author(s)**

Jelle Goeman: <j.j.goeman@lumc.nl>; Jan Oosting

**References**

For references, type: `citation("globaltest")`. See also the vignette `GlobalTest.pdf` included with this package.

**See Also**

`globaltest`, `gtGO`.

**Examples**

```
# Breast cancer data (ExpressionSet) from the Netherlands Cancer
# Institute with annotation:
data(vandeVijver)
data(annotation.vandeVijver)

gt <- globaltest(vandeVijver, "StGallen", annotation.vandeVijver)
sort(gt.multttest(gt))
```

---

permutations

*Permutations version of the Global Test*

---

**Description**

Produces permutations of the `globaltest` test statistic and uses these to recalculate the p-value.

**Usage**

```
permutations(gt, geneset, nperm = 10^4)
```

**Arguments**

<code>gt</code>	The output of a call to <code>globaltest</code> .
<code>geneset</code>	The name or number of the <code>geneset(s)</code> to be used (only necessary if multiple <code>genesets</code> were tested).
<code>nperm</code>	The number of permutations to be used.

**Details**

A permutation p-value is calculated by comparing the value of the test statistic using permutations of the clinical outcome to the value of the test statistic for the true clinical outcome. If the number of possible permutations is smaller than the value of `nperm`, all possible permutations are used, otherwise `nperm` random permutations. The permuted test statistics are stored for later visualisation using `hist`.

**Value**

An object of class `gt.result`.

**Note**

Permutations does not work if the adjusted version of `globaltest` was used.

**Author(s)**

Jelle Goeman: `<j.j.goeman@lumc.nl>`; Jan Oosting

**References**

For references, type: `citation("globaltest")`. See also the vignette `GlobalTest.pdf` included with this package.

**See Also**

`globaltest`, `sampleplot`, `geneplot`.

**Examples**

```
# Breast cancer data (ExpressionSet) from the Netherlands Cancer
# Institute with annotation:
data(vandeVijver)
data(annotation.vandeVijver)
aPathway <- annotation.vandeVijver[1]

gt <- globaltest(vandeVijver, "StGallen", aPathway)
perm.gt <- permutations(gt)

if (interactive()) {
  hist(perm.gt)
}
```

---

regressionplot      *Regression Plot for Global Test*

---

**Description**

Produces a plot which can be used to visualize the effect of specific samples on the test result produced by `globaltest`.

**Usage**

```
regressionplot(gt, geneset, sampleid, ...)
```

**Arguments**

<code>gt</code>	The output of a call to <code>globaltest</code> .
<code>geneset</code>	The name or number of the geneset to be plotted (only necessary if multiple genesets were tested).
<code>sampleid</code>	An optional vector of names or numbers of the samples of interest.
<code>...</code>	Any extra arguments will be forwarded to the plotting function.

## Details

The regressionplot plots, for all pairs of samples, the covariance between the expression patterns against the covariance between their clinical outcomes. Each point in the plot therefore represents a pair of samples. A regression line is fitted through the samples, which visualizes the test result of the function `globaltest`. A steeply increasing slope indicates a high (possibly significant) value of the test statistic.

An optional argument `sampleid` can be supplied, giving sample numbers of possibly outlying arrays. In this case, all pairs of arrays involving one of the arrays in `sampleid` is marked as a red cross, while the other pairs are marked as a blue dot. The blue line which is fitted through all points can now be compared to a red dotted line which is fitted through only the red crosses.

## Value

NULL (no output).

## Note

Regressionplot does not work if the adjusted version of globaltest was used.

## Author(s)

Jelle Goeman: `<j.j.goeman@lumc.nl>`; Jan Oosting

## References

J. J. Goeman, S. A. van de Geer, F. de Kort and J. C. van Houwelingen, 2004, *A global test for groups of genes: testing association with a clinical outcome*, *Bioinformatics* 20 (1) 93–99. See also the `How To Globaltest.pdf` included with this package.

## See Also

`globaltest`, `sampleplot`, `geneplot`.

## Examples

```
# Breast cancer data (ExpressionSet) from the Netherlands Cancer
# Institute with annotation:
data(vandeVijver)
data(annotation.vandeVijver)

gt <- globaltest(vandeVijver, "ESR1", annotation.vandeVijver)

if (interactive()) {
  regressionplot(gt[1])
  regressionplot(gt[1], sampleid = "122")
}
```



sampleplot

*Sample Plot for Global Test***Description**

Produces a plot to show the influence of individual samples on the test result produced by `globaltest`.

**Usage**

```
sampleplot(gt, geneset, samplesubset, scale = TRUE, drawlabels = TRUE,
           labelsize = 0.6, plot = TRUE, addlegend = TRUE, ...)
```

**Arguments**

<code>gt</code>	The output of a call to <code>globaltest</code> .
<code>geneset</code>	The name or number of the geneset to be plotted (only necessary if multiple genesets were tested).
<code>samplesubset</code>	A vector of names or numbers of samples to be plotted. Default: plot all samples
<code>scale</code>	Logical: should the bars be scaled to unit standard deviation?
<code>drawlabels</code>	Logical value to control drawing of the samplenames on the x-axis of the plot.
<code>labelsize</code>	Relative size of the labels on the x-axis. If it is <code>NULL</code> , the current value for <code>HYPERLINK(par("cex.axis"))(par("cex.axis"))</code> is used
<code>plot</code>	If <code>FALSE</code> : does not plot, but only returns a <code>gt.barplot</code> object.
<code>addlegend</code>	If <code>FALSE</code> : does not add a legend to the plot or to the <code>gt.barplot</code> object.
<code>...</code>	Any extra arguments will be forwarded to the plotting function.

**Details**

The `sampleplot` shows a bar and a reference line for each sample. The bar shows the influence of each gene on the test statistic. Samples with a positive influence carry evidence against the null hypothesis (in favour of a significant p-value), because they are similar in expression profile to samples with a similar clinical outcome. Samples with a negative influence bar supply evidence in favour of the null hypothesis and of a non-significant p-value: they are relatively similar in expression profile to samples with a different clinical outcome.

The influence varies around zero if the tested geneset is not associated with the outcome. Marks on the bars show the standarddeviation of the influence under the null hypothesis for those samples which are more than one standard deviation away from zero.

The color of the bar indicates the sign of the residual of Y. In a logistic model the coloring this distinguishes the original groups.

The bottom margin is adjusted to allow enough space for the longest samplename to draw under the axis.

**Value**

An object of type `gt.barplot`.

**Author(s)**

Jelle Goeman: `<j.j.goeman@lumc.nl>`; Jan Oosting

**See Also**

[globaltest](#), [genepLOT](#), [regressionplot](#), [checkerboard](#).

**Examples**

```
# Breast cancer data (ExpressionSet) from the Netherlands Cancer
# Institute with annotation:
data(vandevijver)
data(annotation.vandevijver)

gt <- globaltest(vandevijver, "StGallen", annotation.vandevijver)

if (interactive()){
  sampleplot(gt[1])
}

sp <- sampleplot(gt[1], plot = FALSE)
if (interactive()){
  plot(sort(sp))
}
```

---

sampling

*Sampling random 'pathways' for the Global Test*

---

**Description**

For every pathway in a result of `globaltest`, calculates how many randomly drawn groups of genes of the same size have a smaller or equal p-value.

**Usage**

```
sampling(gt, geneset, ndraws = 10^3)
```

**Arguments**

<code>gt</code>	The output of a call to <code>globaltest</code> .
<code>geneset</code>	The name or number of the geneset(s) to be used (only necessary if multiple genesets were tested).
<code>ndraws</code>	The number of random pathways to be used.

**Details**

For every pathway in `gt[geneset]`, a number `ndraws` random 'pathways' is selected by randomly sampling sets of genes of the same size as the tested pathway. A 'comparative p-value' is calculated by counting what proportion of the random pathways has a larger standardized test statistic  $(Q - EQ) / sd(Q)$  than the tested pathway.

**Value**

An object of class `gt.result`.

**Note**

The function `sampling` cannot be applied to a `gt.result` object resulting from a call to `permutations`.

**Author(s)**

Jelle Goeman: (j.j.goeman@lumc.nl); Jan Oosting

**References**

Jelle J. Goeman, Jan Oosting, Anne-Marie Cleton-Jansen, Jakob, K. Anninga, Hans C. van Houwelingen (2005) Testing association of a pathway with survival. *Bioinformatics* 21, 1950-1957. See also the vignette `Globaltest.pdf` that comes with this package.

**See Also**

`globaltest`, `permutations`, `sampleplot`, `geneplot`.

**Examples**

```
# Breast cancer data (ExpressionSet) from the Netherlands Cancer
# Institute with annotation:
data(vandeVijver)
data(annotation.vandeVijver)

gt <- globaltest(vandeVijver, "StGallen", annotation.vandeVijver)

sampling(gt, ndraws=100)
```

---

vandeVijver	<i>A subset of the Breast cancer data from Rosetta Inpharmatics LLC and Netherlands Cancer Institute.</i>
-------------	---

---

**Description**

An `ExpressionSet` containing expression data of 100 breast cancer patients on 407 selected genes plus the following phenotype data.

**SampleID** Sample ID given by NKI for 295 samples in cohort study (NEJM 347, 01999, 2002)

**FirstSeriesID** Sample ID in first series given by NKI for 78 samples in our first study (Nature 415, p530, 2002)

**Posnodes** Lymph node status from pathology report

**EVENTmeta** Distant metastasis at any time during follow-up, 0 = no, 1 = yes

**EVENTdeath** Death, 0 = no, 1 = yes

**TIMEsurvival** Total survival interval in years between first date of treatment and last date of follow-up (if `EVENTdeath` = 0) or date of death (if `EVENTdeath` = 1)

**TIMErecurrence** Disease free interval in years between first date of treatment and last date of follow-up (if all 'EVENTs' are 0) or date of first EVENT (if one or more 'EVENTs' are 1, i.e., local recurrence, loco-regional recurrence, second primary tumor or distant metastasis)

**TIMEmeta** Metastasis free interval in years between first date of treatment and date of diagnosis of distant metastasis (only if EVENTmeta = 1) (note that interval relates to metastasis at any time during follow-up and not just as first event)

**ESR1** Estrogen receptor alpha expression measurement from microarray (0 = no, 1 = yes)

**NIH** Patient characteristics according to NIH criteria, 1 = low risk, 0 = high risk

**St Gallen** Patient characteristics according to St. Gallen criteria, 1 = low risk, 0 = intermediate/high risk

### Usage

```
data(vandeVijver)
```

### Format

An [ExpressionSet](#). See the Biobase package

### References

M.J. van de Vijver, Y.D. He, L.J. van 't Veer, H. Dai, A.A.M. Hart, D.W. Voskuil, G.J. Schreiber, J.L. Peterse, C. Roberts, M.J. Marton, M. Parrish, D. Atsma, A. Witteveen, A. Glas, L. Delahaye, T. van der Velde, H. Bartelink, S. Rodenhuis, E.T. Rutgers, S.H. Friend, and R. Bernards (2002). A gene-expression signature as a predictor of survival in breast cancer. *New England Journal of Medicine* 347 (25), 1999–2009.

### See Also

[annotation.vandeVijver](#).

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