Package 'ccrepe'

October 7, 2014

Type Package

Title ccrepe_and_nc.score

Version 1.0.0

Imports infotheo (>= 1.1)

Date 2014-01-24

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Description The CCREPE (Compositionality Corrected by REnormalizaion and PErmutation) package is designed to assess the significance of general similarity measures in compositional datasets. In microbial abundance data, for example, the total abundances of all microbes sum to one; CCREPE is designed to take this constraint into account when assigning p-values to similarity measures between the microbes. The package has two functions: ccrepe: Calculates similarity measures, p-values and q-values for relative abundances of bugs in one or two body sites using bootstrap and permutation matrices of the data. nc.score: Calculates species-level co-variation and co-exclusion patterns based on an extension of the checkerboard score to ordinal data.

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VignetteBuilder knitr

Suggests knitr, BiocStyle, BiocGenerics

biocViews StatisticalMethod, Metagenomics, Software

R topics documented:

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Description

ccrepe was developed for use with microbial relative abundance data data, which is both sparse and compositional in nature.

Details

Package: ccrepe Type: Package Version: 1.0

Date: 2013-04-18 License: MIT

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ccrepe Calculates compositionality-corrected p-values and q-values for compositional data using an arbitrary distance metric.

Description

ccrepe calculates compositionality-corrected p-values and q-values for compositional data by generating first a null distribution of the distance metric generated by permutation and renormalization of the data, and then by generating an alternative distribution of the distance metric by bootstrap resampling of the data. For greater detail, see References

The two distributions are compared using a pooled-variance Z-test to give a compositionality-corrected p-value. The p-values can be calculated for all appropriate (passing certain quality-control measures) pairwise comparisons, or for a subset of user-specified ones.

Q-values are additionally calculated using the Benjamin-Hochberg-Yekutieli procedure (see References)

Usage

```
ccrepe(
x = NA
y = NA,
sim.score = cor,
sim.score.args = list(),
min.subj = 20,
iterations = 1000,
subset.cols.x = NULL,
subset.cols.y = NULL,
errthresh = 1e-04,
verbose = FALSE,
iterations.gap = 100,
distributions = NA,
compare.within.x = TRUE,
concurrent.output = NA,
make.output.table = FALSE)
```

Arguments

x First dataframe or matrix containing the relative abundances in cavity1: columns are bugs, rows are samples. (Rows should therefore sum to a constant.)

The subjectIDs, if present, are assumed to be the row names and NOT the first column of data.

Second dataframe or matrix (optional) containing the relative abundances in cavity2: columns are bugs, rows are samples.

The subjectIDs, if present, are assumed to be the row names. If both x and y are specified, they will be merged by row names. If no row names are specified for either or both datasets, the default is to use the row numbers as subject IDs.

A function defining a similarity measure, such as cor or nc.score. This similarity measure can be a pre-defined R function or user-defined. If the latter, certain properties should be satisfied as detailed below (also see examples). The default similarity measure is Spearman correlation.

A user-defined similarity measure should:

У

sim.score

> 1.Be able to take either two inputs which are vectors or one input which is either a matrix or a dataframe

2.In the case of two inputs, return a single number

3.In the case of one input, return a matrix in which the (i,j)th entry is the similarity score for column i and column j in the original matrix

4. Resulting matrix (in the case of one input) must be symmetric

5. The inputs must be named x and y

sim.score.args A list of arguments for the measurement function. For example: In the case of cor, the following would be acceptable: sim.score.args = list(method='spearman', use='complete.obs'

Note that this is the default behavior.

min.subj Minimum number of subjects that must be measured in a bug/feature/column

in order to apply the similarity measure to that bug/feature/column. This is to ensure that there are sufficient subjects to perform a bootstrap (default: 20).

iterations The number of iterations of bootstrap and permutation (default: 1000).

subset.cols.x Subset of columns from x to work on. (Default: NULL - meaning: Use all columns of x. All the columns of x are used for normalization but calculations

are performed only with the requested subset (by default, all columns).

Subset of columns from y to work on. (Default: NULL - meaning: Use all subset.cols.y

columns of y. If applicable (y present), all the columns of y are used for normalization, but calculations are performed only with the requested subset (by

default, all columns).

errthresh A numeric value representing the probability of getting all 0's in a given boot-

strapped column for the first dataset If a bug/feature/column has a number of zeros that makes the probability of obtaining all zeros when sampling with replacement > errthresh, that bug/feature/column will be excluded from the subsequent analysis. This is to ensure that the standard deviation of the bootstrap

sample is non-zero. (default= 0.0001)

Logical: an indicator whether the user requested verbose output, which prints verhose

periodic progress of the algorithm through the dataset(s), as well as including

more detailed output. (default:FALSE)

iterations.gap If output is verbose - number of iterations after issue a status message (De-

fault=100 - displayed only if verbose=TRUE).

distributions Output Distribution file (default:NA).

compare.within.x

A boolean value indicating whether to do comparisons given by taking all subsets of size 2 from subset.cols.x or to do comparisons given by taking all possible combinations of subset.cols.x or subset.cols.y. If TRUE but subset.cols.y=NA,

returns all comparisons involving any features in subset.cols.x

concurrent.output

Optional output file to which each comparison will be written as it is calculated.

make.output.table

A boolean value indicating whether to include table-formatted output.

Value

Returns a list containing the calculation results and the parameters used. Default parameters shown:

min.subj	Description above
errThresh	Description same as errthresh above
sim.score	A matrix of the simliarity scores for all the requested comparisons. The (i,j) th element of sim.score correponds to the similarity score of column i (or the i th column of subset.cols.1) and column j (or the j th column of subset.cols.1) in one dataset, or to the similarity score of column i (or the i th column of subset.cols.1) in dataset x and column y (or the y th column of subset.cols.2) in dataset y in the case of two datasets.
p.values	A matrix of the p-values for all the requested comparisons. The (i,j)th element of p.values corresponds to the p-value of the (i,j)th element of sim.score.
q.values	A matrix of the Benjamini-Hochberg-Yekutieli FDR corrected p-values. The (i,j) th element of q.values corresponds to the q-value fo the (i,j) th element of sim.score.
z.stat	A matrix of the z-statistics for all the requested comparisons. The (i,j)th element corresponds to the z-statistic which gave rise to the (i,j)th p-value.
output.table	(Only if make.output.table=TRUE) A table where each row is one comparision. Each row contains the features being compared with their similarity scores, z-statistics, p-values and q-values

Additional parameters if verbose=TRUE:

iterations Description Above subset.cols.x Description Above subset.cols.y Description Above iterations.gap Description Above sim.score.parameters Description Above compare.within.x Description Above make.output.table Description Above

Author(s)

Emma Schwager <emma.schwager@gmail.com>

References

Emma Schwager and Colleagues. Detecting statistically significant associtations between sparse and high dimensional compositioanl data. In Progress.

Benjamini and Yekutieli (2001). "The control of the false discovery rate in multiple testing under dependency." The Annals of Statistics. Vol. 19, No. 4. pp. 1165-1188.

Examples

```
data <- matrix(rlnorm(40,meanlog=0,sdlog=1),nrow=10)
data.rowsum <- apply(data,1,sum)
data.norm <- data/data.rowsum
testdata <- data.norm
dimnames(testdata) <- list(paste("Sample",seq(1,10)),paste("Feature",seq(1,4)))
ccrepe.results <-ccrepe (x=testdata, iterations=20, min.subj=10)
ccrepe.results.nc.score <- ccrepe(x=testdata,iterations=20,min.subj=10,sim.score=nc.score)
ccrepe.results
ccrepe.results.nc.score</pre>
```

ccrepeSampleTestFunction

ccrepeSampleTestFunction - Simple example of a test measurent function to be used with ccrepe

Description

This simple example of a test measurent function to be used with ccrepe used in the same fashion that cor would be used

Some properties of the function:

- 1. Be able to take either two inputs which are vectors or one input which is either a matrix or a data frame
- 3.In the case of one input, return a matrix in which the (i,j)th entry is the similarity score for column i and column j in the original matrix
- 4. Resulting matrix must be symmetric
- 5. The inputs must be named x and y

Usage

```
ccrepeSampleTestFunction(x, y = NA)
```

Arguments

x x is a vector or a matrix

y y is a vector.

if y selected then x must be a vector too

Value

If x and y are vectors it returns a number: 0.5 If x is a matrix it returns a matrix of all 0.5

Author(s)

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Description

nc.score calculates species-level co-variation and co-exclusion patterns based on an extension of the checkerboard score to ordinal data.

It is an extension to Diamond's checkerboard score (See references below) to ordinal data and implements a framework for robust detection of species-level association patterns in metagenomic data. It performs quality control and transforms relative abundances to ordinal values and computes an association measure, the NC-score (n-dimensional checkerboard score), for each species pair.

Usage

```
nc.score(x = NA,
  y = NA,
bins = NA,
verbose = FALSE,
min.abundance = 1e-04,
min.samples = 0.1)
```

Arguments

X	A numeric vector, data frame, or matrix. The first entity to be processed. Columns are bugs, rows are samples.
у	Numeric vector. If selected, x must be a numeric vector as well.
bins	Either a single integer specifying the number of bins to use or a numeric vector specifying the cutoffs. If a single number is given, this is used as the number of bins in the discretize function of the package infotheo. If a vector is specified, the function findInterval is used to discretize the data according to the cutoffs given. The default behavior is to use the defaults for the discretize function.

verbose Logical flag to request verbose output

min. abundance A numeric value specifying the minimum abundance threshold: Assures selec-

tion of species with abundance >= min.abundance in more than min.samples

percent of samples.

min.samples An fraction specifying the minimum samples threshold: Assures selection of

species with abundance >= min.abundance in more than min.samples percent of

samples. Default=0.1

Value

Matrix or vector of normalized scores.

Author(s)

Craig BielskiCraig Bielski<a href="mailto:Craig.bielski@gmailto

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References

Emma Schwager and Colleagues. Detecting statistically significant associtations between sparse and high dimensional compositioanl data. In Progress.

Examples

```
data <- matrix(rlnorm(40,meanlog=0,sdlog=1),nrow=10)
data.rowsum <- apply(data,1,sum)
data.norm <- data/data.rowsum
testdata <- data.norm
dimnames(testdata) <- list(paste("Sample",seq(1,10)),paste("Feature",seq(1,4)))
nc.score.results <- nc.score( x=testdata )
nc.score.results.bins <- nc.score( x=testdata, bins=5 )
nc.score.results.bin.cutoffs <- nc.score( x=testdata, bins = c(.001,.25,.5) )
nc.score.results.bins
nc.score.results.bins
nc.score.results.bin.cutoffs</pre>
```

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