

# Package ‘dualKS’

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**Type** Package

**Title** Dual KS Discriminant Analysis and Classification

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**Description** This package implements a Kolmogorov Smirnov rank-sum based algorithm for training (i.e. discriminant analysis--identification of genes that discriminate between classes) and classification of gene expression data sets. One of the chief strengths of this approach is that it is amenable to the ``multiclass" problem. That is, it can discriminate between more than 2 classes.

**Collate** AllClasses.R summary-methods.R plot-methods.R show-methods.R  
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dksToGeneSet.R dksTrain.R dksWeights.R KS.R makeClassifier.R  
predictClass.R rKS.R rKSScan.R

**License** LGPL (>= 2.0)

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|                |   |
|----------------|---|
| dualKS-package | <i>Dual KS Discriminant Analysis and Classification</i> |
|----------------|---|

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## Description

This package implements a Kolmogorov Smirnov rank-sum based algorithm for training (i.e. discriminant analysis—identification of genes that discriminate between classes) and classification of gene expression data sets. One of the chief strengths of this approach is that it is amenable to the "multiclass" problem. That is, it can discriminate between more than 2 classes.

## Details

Package: dualKS  
 Type: Package  
 Version: 1.0  
 Date: 2008-04-18  
 License: GPL-3

Use of this package requires an ExpressionSet, pre-processed to your liking, containing gene expression data you wish to train on to identify discriminant genes. Optionally, a second ExpressionSet may be employed for subsequent testing or "diagnosis".

If the ExpressionSet does not contain the appropriate phenoData specifying the classes for each sample, a factor containing this information will also be required.

See the examples for further details.

## Author(s)

Eric J. Kort Maintainer: <Eric.Kort@vai.org>

---

|     |   |
|-----|---|
| dks | <i>A demonstration data set for package dualKS.</i> |
|-----|---|

---

## Description

This data set contains an ExpressionSet comprised of a subset of data from the Gene Expression Omnibus (GEO) data set GDS2126 (Rheumatoid arthritis: synovial tissues). The data set has been severely pruned down to allow quick execution of the examples associated with this package.

**Usage**

```
data("dks")
```

**Format**

MIAME formatted data as an ExpressionSet.

**Source**

[http://www.ncbi.nlm.nih.gov/geo/gds/gds\\_browse.cgi?gds=2126](http://www.ncbi.nlm.nih.gov/geo/gds/gds_browse.cgi?gds=2126)

---

|               |  |
|---------------|--|
| DKSClassifier | <i>Class to contain gene set signatures.</i> |
|---------------|--|

---

**Description**

The function `dksSelectGenes` returns an object of class `DKSClassifier` that contains two `DKSGeneSet` objects: one for each signature (upregulated and downregulated), needed for subsequent classification. This way, the gene scoring does not need to be repeated for classifiers based on different numbers of genes (see `DKSGeneScores` and `dksTrain`).

**Author(s)**

Eric J. Kort

**See Also**

[DKSGeneScores-class](#), [dksTrain](#).

---

|                          |  |
|--------------------------|--|
| <code>dksClassify</code> | <i>Predict classes for gene expression sets.</i> |
|--------------------------|--|

---

**Description**

Kolmogorov-Smirnov rank sum scoring will be used to assign one or more samples to one of two or more classes based on previously defined gene signatures (see `dksTrain`).

**Usage**

```
dksClassify(eset, classifier, rescale=FALSE, method="kort")
```

**Arguments**

|            |  |
|------------|--|
| eset       | An ExpressionSet or matrix containing the gene expression data for the samples to be classified.   |
| classifier | An <a href="#">DKSClassifier</a> produced by <a href="#">dksSelectGenes</a> describing the gene expression signature for each class.   |
| rescale    | If TRUE, scores for each class will be mean centered and normalized to remove arbitrary differences in scale and baseline value between signatures for different classes.  |
| method     | Two methods are supported. The 'kort' method returns the maximum of the running sum. The 'yang' method returns the sum of the maximum and the minimum of the running sum, thereby penalizing classes that are highly enriched in a subset of genes of a given signature, but highly down regulated in another subset of that same signature. |

**Value**

An object of class [DKSPredicted](#) containing the class to which each sample in the eset was assigned as well as other information. This object has its own `summary` and `show` functions useful for displaying this information in a user friendly format.

**Author(s)**

Eric J. Kort, Yarong Yang

**See Also**

[dksTrain](#), [dksSelectGenes](#), [dksClassify](#), [DKSGeneScores](#), [DKSPredicted](#), [DKSClassifier](#)

**Examples**

```
data("dks")
tr <- dksTrain(eset, 1, "up")
cl <- dksSelectGenes(tr, 100)
pr <- dksClassify(eset, cl, rescale=FALSE)
summary(pr, pData(eset)[,1])
show(pr)
plot(pr, actual=pData(eset)[,1])
```

---

dksCustomClass

*Create a classification object from predefined gene signature.*

---

**Description**

This utility function will build a [DKSClassifier](#) object from your own list of gene ids for use by [dksClassify](#). This is useful if you want to use the classification functionality of this package, but already have gene signatures you want to use (as opposed to generating them with [dksTrain](#)).

**Usage**

```
dksCustomClass(upgenes=NULL, upclass=NULL, downgenes=NULL, downclass=NULL)
```

**Arguments**

|           |  |
|-----------|--|
| upgenes   | A vector of gene ids that are upregulated in the classes of interest. Note that upgenes and downgenes need not both be specified, but at least one must be.  |
| upclass   | A vector or factor of classes specifying which signature each gene in upgenes belongs to. (Note that upgenes can describe an arbitrary number of classes, and the order of upgenes is not important so long as the position of ids in upgenes and the position of classes in upclass correspond to one another).           |
| downgenes | A vector of gene ids that are downregulated in the classes of interest. Note that upgenes and downgenes need not both be specified, but at least one must be.  |
| downclass | A vector or factor of classes specifying which signature each gene in downgenes belongs to. (Note that downgenes can describe an arbitrary number of classes, and the order of downgenes is not important so long as the position of ids in downgenes and the position of classes in downclass correspond to one another). |

**Value**

An object of class `DKSClassifier`.

**Author(s)**

Eric J. Kort, Yarong Yang

**See Also**

[dksTrain](#), [dksSelectGenes](#), [dksClassify](#), [DKSGeneScores](#), [DKSPredicted](#), [DKSClassifier](#)

**Examples**

```
data("dks")
up <- rownames(exprs(eset))[1:300]
cls <- factor(rep(c(1,2,3), 100))
classifier <- dksCustomClass(upgenes=up, upclass=cls)
```

---

DKSGeneScores

*Class to contain dual Kolmogorov Smirnov rank sum discriminant analysis results for gene expression data.*

---

**Description**

The function `dksTrain` returns an object of class `DKSGeneScore` to hold the analysis results for subsequent classifier extraction and classification. This way, the gene scoring does not need to be repeated for classifiers based on different numbers of genes.

**Creating Objects**

```
new("DKSGeneScore", gscores.up=new("matrix"), gscores.down=new("matrix"))
```

**Slots**

`gscores.up`: A matrix of gene scores, one column per class

`gscores.down`: A matrix of gene scores, one column per class

**Author(s)**

Eric J. Kort

**See Also**[DKSPredicted-class](#), [dksTrain](#).

---

**DKSGeneSet***Class to contain gene signatures.*

---

**Description**

This class contains a vector of genes, and a corresponding factor of classes that indicates to which signature each gene belongs. The function [dksSelectGenes](#) extracts gene signatures of a specified size from an object of class [DKSGeneScores](#).

calculates a score for each possible class (as determined by the [DKSGeneSet](#) passed to it) for each sample in the test set passed to the function. It then determines which class each sample most likely belongs to based on which of those scores is the largest. All this information is useful after the analysis, and so it is retained in the return object of class [DKSPredicted](#).

**Slots**

**genes:** A vector of gene identifiers. Order is not important.

**classes:** A factor specifying which class (signature) each gene belongs to.

**Author(s)**

Eric J. Kort

**See Also**[DKSGeneScores-class](#), [DKSClassifier-class](#), [dksSelectGenes](#).

---

**dksPerm***Estimate significance of signature scores.*

---

**Description**

The distribution of Kolmogorov Rank Sum scores generated by this package depends on a variety of factors including the size of the signature and the total number of genes measured in each sample. For a given classifier, this function bootstraps an approximate distribution for the scores and then identifies optimum parameters for the gamma distribution that best fits the bootstrap distribution. The corresponding gamma probability function is then returned, allowing p-values for one or more scores to be readily computed.

**Usage**

```
dksPerm(eset, class, n=100, samples=100, type="up", rescale=FALSE,
        verbose=FALSE, method="kort")
```

**Arguments**

|         |   |
|---------|---|
| eset    | An ExpressionSet or matrix containing the gene expression data to be used for bootstrapping.  |
| class   | A factor with two or more levels indicating which class each sample in the expression set belongs OR an integer indicating which column of pData(eset) contains this information. |
| n       | The number of genes per class to use in the bootstrap signature.  |
| samples | The number of bootstrap samples to generate. A value of at least 1000 give good results, but may take a a while.  |
| type    | One of "up", "down", or "both". See <a href="#">dksTrain</a> .  |
| rescale | Logical indicating whether scores should be rescaled to range c(0,1).   |
| verbose | Set to TRUE if you want more evidence of progress while data is being processed. Set to FALSE if you want your CPU cycles to be used on analysis and not printing messages.       |
| method  | One of either 'kort' or 'yang'. Should match that used for <a href="#">dksTrain</a> .   |

**Value**

A function (1-pgamma(x, ...)) with the appropriate parameters preset based on log likelihood maximization relative to the bootstrapped distribution.

**Note**

All arguments should match those used by [dksClassify](#), otherwise the estimated p-values will not meaningfully describe the distribution of scores generated by that function.

**Author(s)**

Eric J. Kort

**See Also**

[dksTrain](#), [dksSelectGenes](#), [dksClassify](#), [DKSGeneScores](#), [DKSPredicted](#), [DKSClassifier](#)

**Examples**

```
data("dks")
p.value <- dksPerm(eset, 1, samples=25)
# this is not nearly enough samples, but will suffice for
# the demonstration. See the vignette for more informative
# example.
p.value(250)
p.value(1500)
```

---

DKSPredicted

*Class to contain classification results from dksClassify*

---

### Description

The function `dksClassify` calculates a score for each possible class (as determined by the `DKSGeneSet` passed to it) for each sample in the test set passed to the function. It then determines which class each sample most likely belongs to based on which of those scores is the largest. All this information is useful after the analysis, and so it is retained in the return object of class `DKSPredicted`.

### Creating Objects

```
new("DKSPredicted", samples=new("vector"), predictedClass=new("factor"), predictedScore=new("vector"))
```

### Slots

`samples`: A vector of sample names

`predictedClass`: A factor specifying which class each sample belongs to based on the classification.

`predictedScore`: A vector giving the score corresponding to the assigned class for each sample.

`scoreMatrix`: The complete score matrix for future reference.

### Methods

Standard generic methods:

`plot(DKSPredicted)`: Plot scores for each sample and class sorted, in turn, by each class.

`plot(DKSPredicted, missing)`: Plot scores for each sample and class sorted, in turn, by each class.

`summary(DKSPredicted)`: Display summary information for the classification.

`summary(DKSPredicted, factor)`: Display summary information for the classification including the percent correctly classified (as specified by the factor).

`show(DKSPredicted)`: Displays a table of scores and predicted classes for each sample.

### Author(s)

Eric J. Kort

### See Also

[DKSGeneScores-class](#), [DKSClassifier-class](#), [dksClassify](#).



---

|                |   |
|----------------|---|
| dksSelectGenes | <i>Extract gene signatures from a DKSGeneScores object.</i> |
|----------------|---|

---

### Description

The DKSGeneScores returned by [dksTrain](#) holds the rank data for all the genes in the original ExpressioSet. However, generally only the top n genes for each class are desired for classification. Rather than needing to re-run dksTrain every time a signature of different size (n) is desired, you simply extract that top n genes from this object using dksSelectGenes.

### Usage

```
dksSelectGenes(data, n)
```

### Arguments

|      |   |
|------|---|
| data | An object of class DKSGeneScores, typically generated by <a href="#">dksTrain</a> |
| n    | The number of genes, per class, to include in the classification signature.       |

### Value

An object of class [DKSGeneScores](#)

### Author(s)

Eric J. Kort, Yarong Yang

### See Also

[dksTrain](#), [dksSelectGenes](#), [dksClassify](#), [DKSGeneScores](#), [DKSPredicted](#), [DKSClassifier](#)

### Examples

```
data("dks")
tr <- dksTrain(eset, 1, "up")
cl <- dksSelectGenes(tr, 100)
pr <- dksClassify(eset, cl)
summary(pr, pData(eset)[,1])
show(pr)
plot(pr, actual=pData(eset)[,1])
```

dksTrain

*Perform Dual KS Discriminant Analysis***Description**

This function will perform dual KS discriminant analysis on a training set of gene expression data (in the form of an ExpressionSet) and a vector of classes describing which of (two or more) classes each column of data corresponds to. Genes will be ranked based on the degree to which they are upregulated or downregulated in each class, or both. Discriminant gene signatures are then extracted using dksSelectGenes and applied to new samples with dksClassify.

**Usage**

```
dksTrain(eset, class, type = "up", verbose=FALSE, weights=FALSE, logweights=TRUE, method='kort')
```

**Arguments**

|            |  |
|------------|--|
| eset       | Gene expression data in the form of an ExpressionSet or matrix   |
| class      | A factor with two or more levels indicating which class each sample in the expression set belongs OR an integer indicating which column of pData(eset) contains this information.  |
| type       | One of "up", "down", or "both" indicating whether you want to analyze and classify based on up or down regulated genes, or both (note that classification of samples based on down regulated genes from single color experiments should be expected to work well due to the noise at low expression levels. Therefore, 'down', or 'both' should only be used for two color experiments or one color data that has been converted to ratios based on some reference sample(s).) |
| verbose    | Set to TRUE if you want more evidence of progress while data is being processed. Set to FALSE if you want your CPU cycles to be used on analysis and not printing messages.  |
| weights    | Value determines whether and how genes are weighted when building the signatures. See details.   |
| logweights | Should the weights be log10 transformed prior to applying?   |
| method     | Two methods are supported. The 'kort' method returns the maximum of the running sum. The 'yang' method returns the sum of the maximum and the minimum of the running sum, thereby penalizing genes that are highly enriched in a subset of samples of a given class, but highly down regulated in another subset of that same class.   |

**Details**

This function calculates the Kolmogorov-Smirnov rank sum statistic for each gene and each level of 'class'. The highest scoring genes can then be extracted for use in classification.

If weights=FALSE, signatures are defined based on the ranks of members of each class when sorted on each gene. Those genes for which a given class has the highest rank when sorting samples by those genes will be included in the classifier, with no regard to the absolute expression level of those genes. This is the classic KS statistic.

Very discriminant genes identified in this way may or may not be the highest expressed genes. The result is that signatures identified in this way have arbitrary "baseline" values. This may lead to

misclassification when comparing two signatures (using, for example, [dksClassify](#)). Therefore, one may wish to weight genes based on absolute expression level, or some other metric.

Setting `weights = TRUE` causes the genes to be weighted according to the log (base 10) of the relative rank of the mean expression of each gene in each class. Alternatively, you may provide your own weight matrix as the argument to `weights`. This matrix must have one column for each possible value of `class`, and one row for each gene in `eset`. Note that for `type='down'` or the down component of `type='both'`, the weight matrix will be inverted as `1-matrix`, so the range of weights should be 0 - 1 for each class. NAs are handled "gracefully" by discarding any genes for which any column of the corresponding row of `weights` is NA. Our experience has been that weights that are a linear function of some feature of the gene expression (like mean) can be too subtle. The effect of the weights can be increased by setting `logweights=TRUE` (which is the default).

### Value

An object of class [DKSGeneScores](#).

### Author(s)

Eric J. Kort, Yarong Yang

### See Also

[dksTrain](#), [dksSelectGenes](#), [dksClassify](#), [DKSGeneScores](#), [DKSPredicted](#), [DKSClassifier](#)

### Examples

```
data("dks")
tr <- dksTrain(eset, 1, "up")
cl <- dksSelectGenes(tr, 100)
pr <- dksClassify(eset, cl)
summary(pr, pData(eset)[,1])
show(pr)
plot(pr, actual=pData(eset)[,1])
```

---

`dksWeights`

*Calculate gene weights based on average expression.*

---

### Description

Prior to selecting genes it may be desirable to calculate weights for each genes so that some genes are more likely than others to be included in the gene signature all other things being equal. This function will calculate an  $N \times M$  weight matrix for  $N$  genes in `data` and  $M$  unique classes in `class`. The weights are based on mean expression of each gene in each class such that genes that are highly expressed on average in a given class will be weighted more highly when scoring genes for that class.

The resulting weight matrix can be passed to [dksTrain](#) as the `weights` argument.

### Usage

```
dksWeights(eset, class)
```

**Arguments**

|                    |  |
|--------------------|--|
| <code>eset</code>  | An ExpressionSet or matrix containing the gene expression data to be used for bootstrapping.   |
| <code>class</code> | A factor with two or more levels indicating which class each sample in the expression set belongs OR an integer indicating which column of <code>pData(eset)</code> contains this information. |

**Value**

An N x M matrix containing the weights for each gene and each class.

**Note**

There are many metrics the user might want to use for weighting. This convenience function just implements one of the most obvious ones. The user can provide his/her own N x M weight matrix to `dksTrain`. The weight matrix calculated by this function will be calculated on the fly if the `weights` is set to TRUE when calling `dksTrain`. However, if multiple calls to `dksTrain` are being made (for example when performing some type of optimization or validation), it will save a lot of time if the weight matrix is pre-calculated by a call to this function and the resulting matrix supplied directly to `dksTrain` rather than having it re-calculate the weight matrix every time.

**Author(s)**

Eric J. Kort

**See Also**

[dksTrain](#), [dksSelectGenes](#), [dksClassify](#), [DKSGeneScores](#), [DKSPredicted](#), [DKSClassifier](#)

**Examples**

```
data("dks")
wt <- dksWeights(eset, 1)
str(wt)
```

---

KS

*Calculate Kolmogorov Smirnov rank sum scores.*

---

**Description**

This function calculates the degree to which a subset of genes (i.e. a "signature") is biased in the ordered list of all genes. The function is typically used internally by `dksClassify`, but the user may want to call it directly to inspect the running sums.

**Usage**

```
KS(data, geneset, decreasing=TRUE, method="kort")
```

**Arguments**

|            |  |
|------------|--|
| data       | A vector of gene expression data. The data need not be sorted, as the function will sort it itself.  |
| geneset    | A <a href="#">DKSGeneSet</a> object, such as one of the slots of the <a href="#">DKSClassifier</a> returned by <code>link{dksClassify}</code> .  |
| decreasing | Indicates which way data should be sorted. If TRUE, the degree of upregulation will be scored. If FALSE, the degree of down regulation will be scored.   |
| method     | Two methods are supported. The 'kort' method returns the maximum of the running sum. The 'yang' method returns the sum of the maximum and the minimum of the running sum, thereby penalizing classes that are highly enriched in a subset of genes of a given signature, but highly down regulated in another subset of that same signature. |

**Value**

|             |   |
|-------------|---|
| runningSums | A matrix with 1 row per gene and 1 column per signature. The value is the running sum of the KS metric at each point along the sorted list of genes. The maximum of this column vector corresponds to the KS score for the corresponding signature. |
| ksScores    | A named vector giving the KS score for each signature.  |

**Author(s)**

Eric J. Kort, Yarong Yang

**See Also**

[dksTrain](#), [dksSelectGenes](#), [dksClassify](#), [DKSGeneScores](#), [DKSPredicted](#), [DKSClassifier](#)

**Examples**

```
data("dks")
tr <- dksTrain(eset, 1, "both")
cl <- dksSelectGenes(tr, 100)
sc <- KS(exprs(eset)[,1], cl@genes.up)
plot(sc$runningSums[,1], type='l')
```

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