

Additional plots for: Independent filtering increases power for detecting differentially expressed genes, Bourgon et al., PNAS (2010)

Richard Bourgon

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1 Introduction

This vignette illustrates use of some functions in the *genefilter* package that provide useful diagnostics for independent filtering [1]:

- `kappa_p` and `kappa_t`
- `filtered_p` and `filtered_R`
- `filter_volcano`
- `rejection_plot`

2 Data preparation

Load the ALL data set and the *genefilter* package:

```
library("genefilter")
library("ALL")
data("ALL")
```

Reduce to just two conditions, then take a small subset of arrays from these, with 3 arrays per condition:

```
bcell <- grep("^B", as.character(ALL$BT))
moltyp <- which(as.character(ALL$mol.biol) %in%
               c("NEG", "BCR/ABL"))
ALL_bcrneg <- ALL[, intersect(bcell, moltyp)]
ALL_bcrneg$mol.biol <- factor(ALL_bcrneg$mol.biol)
n1 <- n2 <- 3
set.seed(1969)
use <- unlist(tapply(1:ncol(ALL_bcrneg),
                    ALL_bcrneg$mol.biol, sample, n1))
subsample <- ALL_bcrneg[,use]
```

We now use functions from *genefilter* to compute overall standard deviation filter statistics as well as standard two-sample *t* and related statistics.

```
S <- rowSds( exprs( subsample ) )
temp <- rowttests( subsample, subsample$mol.biol )
d <- temp$dm
p <- temp$p.value
t <- temp$statistic
```

3 Filtering volcano plot

Filtering on overall standard deviation and then using a standard *t*-statistic induces a lower bound of fold change, albeit one which varies somewhat with the significance of the *t*-statistic. The `filter_volcano` function allows you to visualize this effect.

The output is shown in the left panel of Fig. 1.

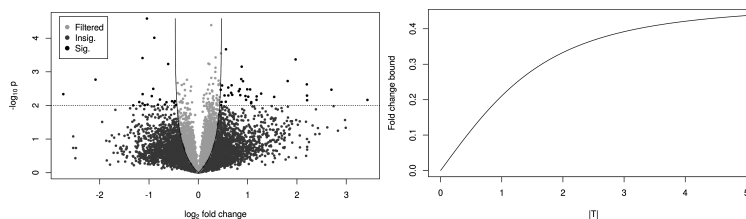


Figure 1: Left panel: plot produced by the `filter_volcano` function. Right panel: graph of the `kappa_t` function.

4.1

The `kappa_p` and `kappa_t` functions, used to make the volcano plot, compute the fold change bound multiplier as a function of either a t -test p -value or the t -statistic itself. The actual induced bound on the fold change is κ times the filter's cutoff on the overall standard deviation. Note that fold change bounds for values of $|T|$ which are close to 0 are not of practical interest because we will not reject the null hypothesis with test statistics in this range.

The plot is shown in the right panel of Fig. 1.

4 Rejection count plots

4.1 Across p -value cutoffs

The `filtered_p` function permits easy simultaneous calculation of unadjusted or adjusted p -values over a range of filtering thresholds (θ). Here, we return to the full "BCR/ABL" versus "NEG" data set, and compute adjusted p -values using the method of Benjamini and Hochberg, for a range of different filter stringencies.

```
table(ALL_bcrneg$mol.biol)
```

```
##  
## BCR/ABL    NEG  
##      37     42
```

```
S2 <- rowVars(exprs(ALL_bcrneg))  
p2 <- rowttests(ALL_bcrneg, "mol.biol")$p.value  
theta <- seq(0, .5, .1)  
p_bh <- filtered_p(S2, p2, theta, method="BH")
```

```
head(p_bh)
```

```
##           0%    10%    20%    30%    40%    50%  
## [1,] 0.919 0.894 0.862 0.828    NA    NA  
## [2,] 0.959 0.946 0.930 0.906 0.887 0.871  
## [3,] 0.702    NA    NA    NA    NA    NA
```

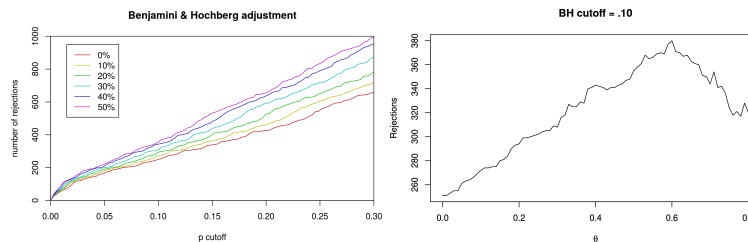


Figure 2: Left panel: plot produced by the `rejection_plot` function. Right panel: graph of `theta`.

```
## [4,] 0.981 0.975 0.968 0.957    NA    NA
## [5,] 0.951 0.935 0.912 0.884    NA    NA
## [6,] 0.634 0.590 0.544 0.495 0.450 0.410
```

The `rejection_plot` function takes sets of p -values corresponding to different filtering choices — in the columns of a matrix or in a list — and shows how rejection count (R) relates to the choice of cutoff for the p -values. For these data, over a reasonable range of FDR cutoffs, increased filtering corresponds to increased rejections.

```
rejection_plot(p_bh, at="sample",
               xlim=c(0,.3), ylim=c(0,1000),
               main="Benjamini & Hochberg adjustment")
```

The plot is shown in the left panel of Fig. 2.

4.2 Across filtering fractions

If we select a fixed cutoff for the adjusted p -values, we can also look more closely at the relationship between the fraction of null hypotheses filtered and the total number of discoveries. The `filtered_R` function wraps `filtered_p` and just returns rejection counts. It requires a p -value cutoff.

```
theta <- seq(0, .80, .01)
R_BH <- filtered_R(alpha=.10, S2, p2, theta, method="BH")
```

```
head(R_BH)
## 0% 1% 2% 3% 4% 5%
## 251 251 253 255 255 261
```

Because overfiltering (or use of a filter which is inappropriate for the application domain) discards both false and true null hypotheses, very large values of θ reduce power in this example:

```
plot(theta, R_BH, type="l",
      xlab=expression(theta), ylab="Rejections",
      main="BH cutoff = .10"
      )
```

The plot is shown in the right panel of Fig. 2.

Session information

- R version 3.3.0 (2016-05-03), x86_64-pc-linux-gnu
- Base packages: base, datasets, grDevices, graphics, methods, parallel, stats, utils
- Other packages: ALL 1.14.0, Biobase 2.32.0, BiocGenerics 0.18.0, DESeq 1.24.0, RColorBrewer 1.1-2, genefilter 1.54.2, knitr 1.13, lattice 0.20-33, locfit 1.5-9.1, pasilla 0.12.0
- Loaded via a namespace (and not attached): AnnotationDbi 1.34.2, BiocStyle 2.0.2, DBI 0.4-1, IRanges 2.6.0, Matrix 1.2-6, RSQLite 1.0.0, S4Vectors 0.10.0, XML 3.98-1.4, annotate 1.50.0, codetools 0.2-14, digest 0.6.9, evaluate 0.9, formatR 1.4, geneplotter 1.50.0, grid 3.3.0, highr 0.6, magrittr 1.5, splines 3.3.0, stats4 3.3.0, stringi 1.0-1, stringr 1.0.0, survival 2.39-4, tools 3.3.0, xtable 1.8-2

References

- [1] Richard Bourgon, Robert Gentleman and Wolfgang Huber. Independent filtering increases power for detecting differentially expressed genes.