

# Package ‘FCBF’

October 14, 2021

**Type** Package

**Title** Fast Correlation Based Filter for Feature Selection

**Version** 2.0.0

**Description** This package provides a simple R implementation for the Fast Correlation Based Filter described in Yu, L. and Liu, H.; Feature Selection for High-Dimensional Data: A Fast Correlation Based Filter Solution, Proc. 20th Intl. Conf. Mach. Learn. (ICML-2003), Washington DC, 2003

The current package is an intent to make easier for bioinformaticians to use FCBF for feature selection, especially regarding transcriptomic data. This implies discretizing expression (function `discretize_exprs`) before calculating the features that explain the class, but are not predictable by other features.

The functions are implemented based on the algorithm of Yu and Liu, 2003 and Rajarshi Guha's implementation from 13/05/2005 available (as of 26/08/2018) at <http://www.rguha.net/code/R/fcbf.R>.

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**Imports** ggplot2, gridExtra, pbapply, parallel, SummarizedExperiment, stats, mclust

**Suggests** caret, mlbench, SingleCellExperiment, knitr, rmarkdown, testthat, BiocManager

**biocViews** GeneTarget, FeatureExtraction, Classification, GeneExpression, SingleCell, ImmunoOncology

**VignetteBuilder** knitr

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discretize_exprs	<i>discretize_exprs Simple discretizing of gene expression</i>
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### Description

This function takes the range of values for each gene in a previously normalized expression table (genes/variables in rows, samples/ observations in columns) and uses it for a width-based discretization. Each feature is divide into "n" bins of equal width. The first bin is attributed the class 'low' and the next bins are assigned to "high". It transposes the original expression table.

### Usage

```
discretize_exprs(
  expression_table,
  number_of_bins = 3,
  method = "varying_width",
  alpha = 1,
  centers = 3,
  min_max_cutoff = 0.25,
  progress_bar = TRUE
)
```

### Arguments

expression\_table

A previously normalized expression table Note: this might drastically change the number of selected features.

number_of_bins	Number of equal-width bins for discretization. Note: it is a binary discretization, with the first bin becoming one class ('low') and the other bins, another class ('high'). Defaults to 3.
method	Method applied to all genes for discretization. Methods available: "varying_width" (Varying width binarization, default, described in function description. Modulated by the number_of_bins param), "mean" (Split in ON/OFF by each gene mean expression), "median" (Split in ON/OFF by each gene median expression), "mean_sd"(Split in low/medium/high by each assigning "medium" to the interval between mean +- standard_deviation. Modulated by the alpha param, which enlarges (>1) or shrinks (<1) the "medium" interval. ), ), "kmeans"(Split in different groups by the kmeans algorithm. As many groups as specified by the centers param) and "min_max_percent" (Similar to the "varying width", a binarization threshold in a percent of the min-max range is set. (minmaxpercent param)), "GMM" (A Gaussian Mixture Model as implemented by the package mclust, trying to fit 2:5 Gaussians)
alpha	Modulator for the "mean_sd" method.Enlarges (>1) or shrinks (<1) the "medium" interval. Defaults to 1.
centers	Modulator for the "kmeans" method. Defaults to 3.
min_max_cutoff	<- Modulator for the "min_max_percent" method. Defaults to 0.25.
progress_bar	Enables a progress bar for the discretization. Defaults to TRUE.

**Value**

A data frame with the discretized features in the same order as previously

**Examples**

```
data(scDengue)
exprs <- SummarizedExperiment::assay(scDengue, 'logcounts')
discrete_expression <- as.data.frame(discretize_exprs(exprs))
head(discrete_expression[, 1:4])
```

---

discretize\_exprs\_supervised  
*supervised\_disc\_df*

---

**Description**

Uses several discretizations and selects the one that is best for a given variable (gene) in comparison to a target class by equivocation

**Usage**

```
discretize_exprs_supervised(expression_table, target, parallel = FALSE)
```

**Arguments**

expression_table	A previously normalized expression table
target	A series of labels matching each of the values in the gene vector (genes in rows, cells/samples in columns)
parallel	Set calculations in parallel. May be worth it if the number of rows and columns is really large. Do watchout for memory overload.

**Value**

A data frame with the discretized features in the same order as previously

**Examples**

```
data(scDengue)
exprs <- as.data.frame(SummarizedExperiment::assay(scDengue, 'logcounts'))
exprs <- exprs [1:200, 1:120]
infection <- SummarizedExperiment::colData(scDengue)
target <- infection$infection
discrete_expression <- as.data.frame(discretize_exprs_supervised(exprs,target))
fcbf(discrete_expression,target, minimum_su = 0.05, verbose = TRUE)
```

---

discretize\_gene\_supervised

*discretize\_gene\_supervised*

---

**Description**

Uses several discretizations and selects the one that is best for a given variable (gene) in comparison to a target class by equivocation Note that set.seed() should be used for reproducing the results. The inner kmeans #' function would, otherwise, provide different results each time.

**Usage**

```
discretize_gene_supervised(
  gene,
  target,
  output = "discretized_vector",
  discs = c(".split_vector_in_two_by_median", ".split_vector_in_two_by_mean",
    ".split_vector_by_kmeans", ".split_vector_in_three_by_mean_sd",
    ".split_vector_in_two_by_min_max_thresh"),
  vw_params = c(0.25, 0.5, 0.75),
  kmeans_centers = c(2, 3, 4),
  sd_alpha = c(0.75, 1, 1.25)
)
```

**Arguments**

gene	A previously normalized gene expression vector
target	A series of labels matching each of the values in the gene vector
output	If it is equal to 'discretized_vector', the output is the vector. If it is 'su', returns a dataframe. Defaults to 'discretized_vector'
discs	Defaults to c(".split_vector_in_two_by_median", "split_vector_in_two_by_mean", ".split_vector_by_kmeans", ".split_vector_in_three_by_mean_sd", ".split_vector_in_two_by_vw")
vw_params	cutoff parameters for the varying width function. Defaults to 0.25, 0.5 and 0.75
kmeans_centers	Numeric vector with the number of centers to use for kmeans. Defaults to 2, 3 and 4
sd_alpha	Parameter for adjusting the 'medium' level of the mean +/- sd discretization. Defaults to sd_alpha = c(0.75, 1, 1.25))

**Details**

Note that a seed for random values has to be set for reproducibility. Otherwise, the "kmeans" value might vary from iteration to iteration.

**Value**

A data frame with the discretized features in the same order as previously

**Examples**

```
data(scDengue)
exprs <- as.data.frame(SummarizedExperiment::assay(scDengue, 'logcounts'))
gene <- exprs['ENSG00000166825',]
infection <- SummarizedExperiment::colData(scDengue)
target <- infection$infection
set.seed(3)
discrete_expression <- as.data.frame(discretize_gene_supervised(gene, target))
table(discrete_expression)
```

---

fcbf

*Fast Correlation Based Filter function.*


---

**Description**

This function allows selection of variables from a feature table of discrete/categorical variables and a target class. The function is based on the algorithm described in Yu, L. and Liu, H.; Feature Selection for High-Dimensional Data A Fast Correlation Based Filter Solution, Proc. 20th Intl. Conf. Mach. Learn. (ICML-2003), Washington DC, 2003

**Usage**

```
fcbf(
  feature_table,
  target_vector,
  minimum_su = 0.25,
  n_genes_selected_in_first_step = NULL,
  verbose = FALSE,
  samples_in_rows = FALSE,
  balance_classes = FALSE
)
```

**Arguments**

<code>feature_table</code>	A table of features (samples in rows, variables in columns, and each observation in each cell)
<code>target_vector</code>	A target vector, factor containing classes of the observations. Note: the observations must be in the same order as the parameter <code>x</code>
<code>minimum_su</code>	A minimum <code>su</code> old for the minimum correlation (as determined by symmetrical uncertainty) between each variable and the class. Defaults to 0.25. Note: this might drastically change the number of selected features.
<code>n_genes_selected_in_first_step</code>	Sets the number of genes to be selected in the first part of the algorithm. The final number of selected genes is related to this parameter, but depends on the correlation structure of the data. It overrides the <code>minimum_su</code> parameter. If left unchanged, it defaults to <code>NULL</code> and the <code>minimum_su</code> parameter is used.
<code>verbose</code>	Adds verbosity. Defaults to <code>FALSE</code> .
<code>samples_in_rows</code>	A flag for the case in which samples are in rows and variables/genes in columns. Defaults to <code>FALSE</code> .
<code>balance_classes</code>	Balances number of instances in the target vector <code>y</code> by sampling the number of instances in the minor class from all others. The number of samplings is controlled by <code>resampling_number</code> . Defaults to <code>FALSE</code> .

**Details**

Obs: For gene expression, you will need to run `discretize_exprs` first

**Value**

Returns a data frame with the selected features index (first row) and their symmetrical uncertainty values regarding the class (second row). Variable names are present in `rownames`

**Examples**

```
data(scDengue)
exprs <- SummarizedExperiment::assay(scDengue, 'logcounts')
```

```
discrete_expression <- as.data.frame(discretize_exprs(exprs))
head(discrete_expression[,1:4])
infection <- SummarizedExperiment::colData(scDengue)
target <- infection$infection
fcbf(discrete_expression,target, minimum_su = 0.05, verbose = TRUE)
fcbf(discrete_expression,target, n_genes_selected_in_first_step = 100)
```

---

get\_ig\_for\_feature\_table\_and\_vector  
*Get information gain*

---

## Description

This functions runs information gain for a feature table and a class, returning the scores of information gain for all features

## Usage

```
get_ig_for_feature_table_and_vector(feature_table, target_vector)
```

## Arguments

**feature\_table** A table of features (observations in rows, variables in columns)

**target\_vector** A target vector, factor containing classes of the observations. Note: the observations must be in the same order as the parameter x.

## Value

A dataframe containing the SU values for each feature

## Examples

```
data(scDengue)
exprs <- SummarizedExperiment::assay(scDengue, 'logcounts')
discrete_expression <- as.data.frame(discretize_exprs(exprs))
infection <- SummarizedExperiment::colData(scDengue)
target <- infection$infection
ig_values <- get_ig_for_feature_table_and_vector(discrete_expression[,],target[])
ig_values[1:10,]
```

get\_IG\_for\_vector\_pair

*Information Gain This functions runs Information Gain for two features, returning the score*

---

### Description

Information Gain This functions runs Information Gain for two features, returning the score

### Usage

```
get_IG_for_vector_pair(x, y, base = 2)
```

### Arguments

x	A vector containing a categorical feature
y	A vector containing other categorical feature
base	The base used for the logarithmic function. The default is $\exp(1)$ (~2.718)

### Value

A numerical value for the Information Gain score

### Examples

```
data(scDengue)
exprs <- SummarizedExperiment::assay(scDengue, 'logcounts')
discrete_expression <- as.data.frame(discretize_exprs(exprs))
discrete_expression_gene_1 <- discrete_expression$V1
discrete_expression_gene_2 <- discrete_expression$V2
get_IG_for_vector_pair(discrete_expression_gene_1,discrete_expression_gene_2)
```

---

get\_su\_for\_feature\_table\_and\_vector

*Symmetrical Uncertainty diagnostic*

---

### Description

This functions runs symmetrical uncertainty for a feature table and a class, returning the scores of symmetrical uncertainty for all features

**Usage**

```
get_su_for_feature_table_and_vector(
  feature_table,
  target_vector,
  samples_in_rows = FALSE,
  bar_of_progress = FALSE
)
```

**Arguments**

`feature_table` A table of features (observations in rows, variables in columns)

`target_vector` A target vector, factor containing classes of the observations. Note: the observations must be in the same order as the parameter `x`.

`samples_in_rows` A flag for the case in which samples are in rows and variables/genes in columns. Defaults to FALSE.

`bar_of_progress` A flag to show progress. Defaults to FALSE.

**Value**

A dataframe containing the SU values for each feature

**Examples**

```
data(scDengue)
exprs <- SummarizedExperiment::assay(scDengue, 'logcounts')
discrete_expression <- as.data.frame(discretize_exprs(exprs))
infection <- SummarizedExperiment::colData(scDengue)
target <- infection$infection
su_values <- get_su_for_feature_table_and_vector(discrete_expression[,], target[])
su_values[1:10,]
```

---

get\_SU\_for\_vector\_pair

*get\_SU\_for\_vector\_pair* Formula for symmetrical uncertainty as described in Yu, L. and Liu, H. , 2003. This functions runs symmetrical uncertainty for two features, returning the score

---

**Description**

get\_SU\_for\_vector\_pair Formula for symmetrical uncertainty as described in Yu, L. and Liu, H. , 2003. This functions runs symmetrical uncertainty for two features, returning the score

**Usage**

```
get_SU_for_vector_pair(x, y, base = 2)
```

**Arguments**

x	A vector containing a categorical feature
y	A vector containing other categorical feature
base	The base used for the logarithmic function. The default is $\exp(1)$ (~2.718)

**Value**

A numerical value for the Symmetrical Uncertainty score

**Examples**

```
data(scDengue)
exprs <- SummarizedExperiment::assay(scDengue, 'logcounts')
discrete_expression <- as.data.frame(discretize_exprs(exprs))
discrete_expression_gene_1 <- discrete_expression$V1
discrete_expression_gene_2 <- discrete_expression$V2
get_SU_for_vector_pair(discrete_expression_gene_1, discrete_expression_gene_2)
```

---

scDengue	<i>Dengue infected macrophages; gene expression data from GEO study GSE110496</i>
----------	---

---

**Description**

Expression data from single cells, from adengue virus infection study by Zanini et al, #' 2018. The expression was filtered to get cells 12 hours after infection with #' a multiplicity of infection (moi) of 1 (dengue) or uninfected(ctrl). Gene counts were normalized via Bioconductor package "SCNorm".

**Usage**

```
data(scDengue)
```

**Format**

An object of class SingleCellExperiment

**Details**

Gene expression has to be discretized for use in FCBF.

**Source**

[GEO](#)

**References**

Zanini, F., Pu, S. Y., Bekerman, E., Einav, S., & Quake, S. R. (2018). Single-cell transcriptional dynamics of flavivirus infection. *Elife*, 7, e32942. [PubMed](#)

## Examples

```
data(scDengue)
exprs <- SummarizedExperiment::assay(scDengue, 'logcounts')
infection <- SummarizedExperiment::colData(scDengue)
target <- infection$infection
# Discretize gene expression
discrete_expression <- as.data.frame(discretize_exprs(exprs))
fcbf_features <- fcbf(discrete_expression,
                     target,
                     minimum_su = 0.05,
                     verbose = TRUE)
```

---

su\_plot

*Symmetrical Uncertainty diagnostic*

---

## Description

This functions runs symmetrical uncertainty for a feature table and a class, returning an histogram of the scores

## Usage

```
su_plot(feature_table, target_vector)
```

## Arguments

**feature\_table** A table of features (observations in rows, variables in columns)

**target\_vector** A target vector, factor containing classes of the observations. Note: the observations must be in the same order as the parameter x.

## Value

Plots an histogram of symmetrical uncertainty values regarding the class.

## Examples

```
data(scDengue)
exprs <- SummarizedExperiment::assay(scDengue, 'logcounts')
discrete_expression <- as.data.frame(discretize_exprs(exprs))
infection <- SummarizedExperiment::colData(scDengue)
target <- infection$infection
su_plot(discrete_expression, target)
```

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