

# *GenomicFeatures* and *BSgenome*

Patrick Aboyoun

Fred Hutchinson Cancer Research Center

7-9 June, 2010

# Outline

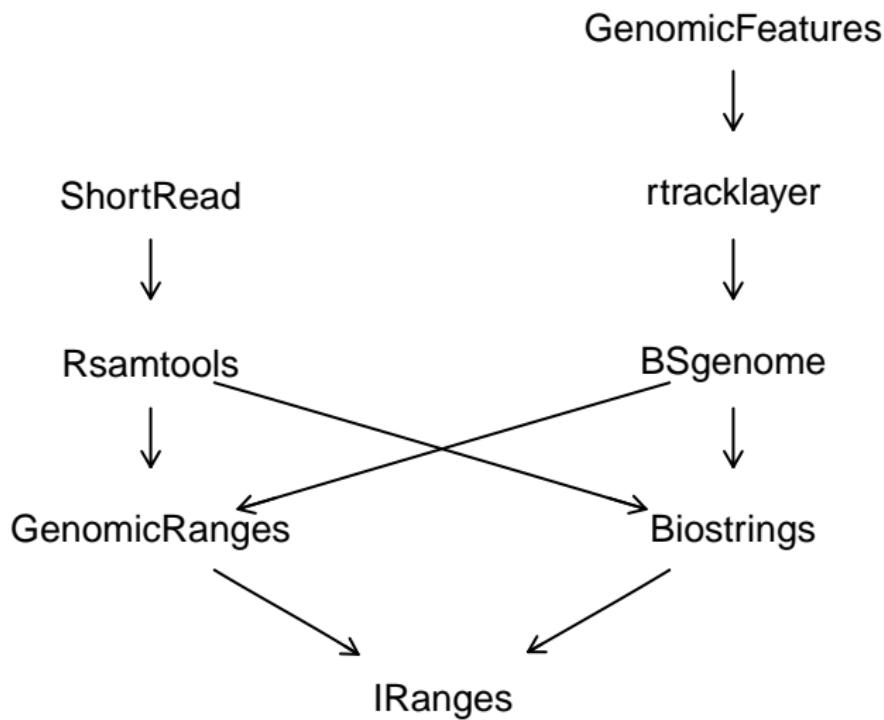
Introduction

Genomic Features

B\$genome

Resources

# Bioconductor Sequence Packages



# *Bioconductor Sequence Annotation Packages*

## *GenomicFeatures*

- ▶ Management of transcript information using *GenomicRanges* infrastructure
- ▶ Transcripts stored in separate SQLite databases

## *BSgenome*

- ▶ Management of whole genomes using *Biostrings* infrastructure
- ▶ Tools for operating on those genomes
- ▶ Genomes stored in separate  
*BSgenome.Organism.Provider.BuildVersion* packages
- ▶ Support for pre-build SNP packages for human

# Outline

Introduction

Genomic Features

B<sub>S</sub> genome

Resources

# *GenomicFeatures* transcript sources

## Constructors

`makeTranscriptDbFromBiomart`, `makeTranscriptDbFromUCSC`

```
> library( GenomicFeatures )
> nrow(supportedUCSCtables())
```

```
[1] 24
```

```
> head(supportedUCSCtables(), 10)
```

	track	subtrack
knownGene	UCSC Genes	<NA>
knownGeneOld3	Old UCSC Genes	<NA>
wgEncodeGencodeManualRel2	Gencode Genes	Genecode Manual
wgEncodeGencodeAutoRel2	Gencode Genes	Genecode Auto
wgEncodeGencodePolyARel2	Gencode Genes	Genecode PolyA
ccdsGene	Consensus CDS	<NA>
refGene	RefSeq Genes	<NA>
xenoRefGene	Other RefSeq	<NA>
vegaGene	Vega Genes	Vega Protein Genes
vegaPseudoGene	Vega Genes	Vega Pseudogenes

# *TranscriptDb* basics

## Making a *TranscriptDb* object

```
> mm9KG <-  
+   makeTranscriptDbFromUCSC(genome = "mm9",  
+                             tablename = "knownGene")
```

## Saving and Loading

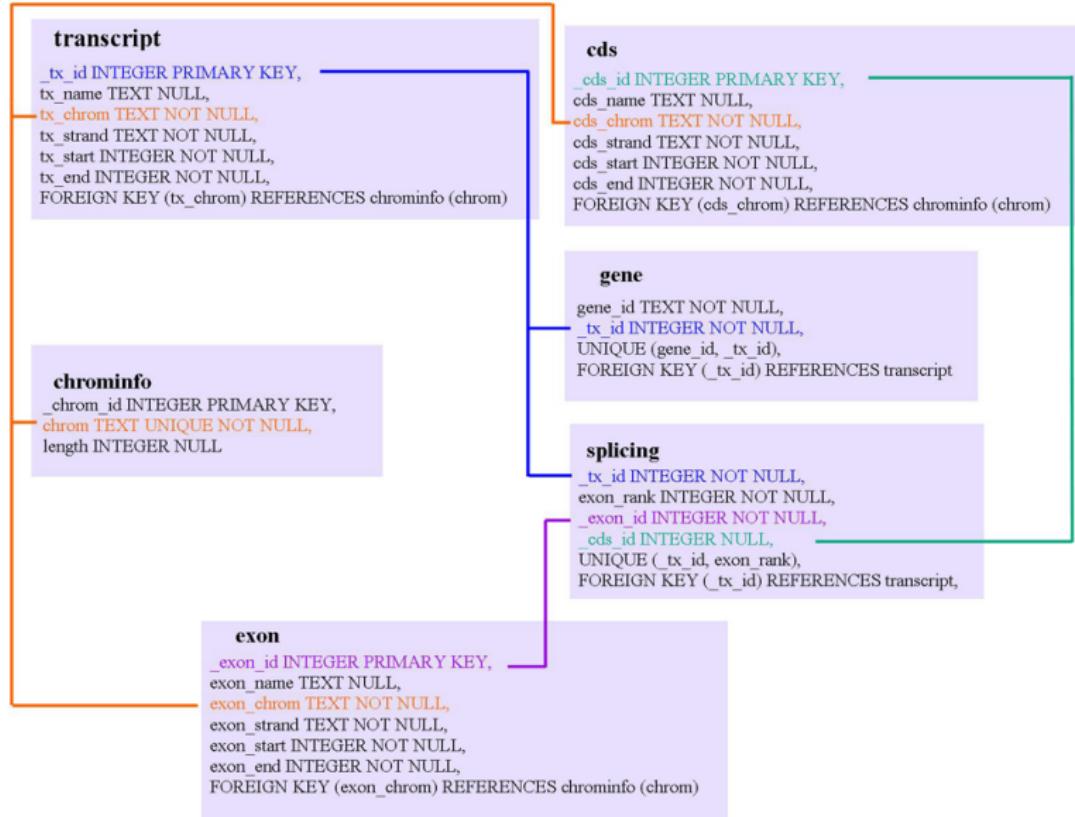
```
> saveFeatures(mm9KG, file="mm9KG.sqlite")  
  
> mm9KGChr9 <-  
+   loadFeatures(system.file("extdata", "mm9KGChr9.sqlite",  
+                           package = "EMBL2010"))
```

## *TranscriptDb* class

```
> mm9KGChr9
```

```
TranscriptDb object:  
| Db type: TranscriptDb  
| Data source: UCSC  
| Genome: mm9  
| UCSC Table: knownGene  
| Type of Gene ID: Entrez Gene ID  
| Full dataset: yes  
| transcript_nrow: 49409  
| exon_nrow: 237551  
| cds_nrow: 204831  
| Db created by: GenomicFeatures package from Bioconductor  
| Creation time: 2010-05-13 17:02:30 -0700 (Thu, 13 May 2010)  
| GenomicFeatures version at creation time: 1.1.1  
| RSQLite version at creation time: 0.8-3
```

# TranscriptDb schema



# Ungrouped transcript-related information

## Extractors

transcripts, exons, cds

```
> tx <- transcripts(mm9KGChr9)
```

```
> length(tx)
```

```
[1] 2910
```

```
> head(tx, 5)
```

GRanges with 5 ranges and 2 elementMetadata values

	seqnames	ranges	strand	tx_id	tx_name
	<Rle>	<IRanges>	<Rle>	<integer>	<character>
[1]	chr9	[3215314, 3215339]	+	24312	uc009oas.1
[2]	chr9	[3335231, 3385846]	+	24315	uc009oat.1
[3]	chr9	[3335473, 3343608]	+	24313	uc009oau.1
[4]	chr9	[3335473, 3380423]	+	24314	uc009oav.1
[5]	chr9	[3335478, 3385846]	+	24316	uc009oaw.1

seqlengths

	chr1	chr2	...	chrX_random	chrY_random
197195432	181748087	...	1785075	58682461	

# Grouped transcript-related information

## Extractors

```
transcriptsBy, exonsBy, cdsBy, intronsByTranscript,  
fiveUTRsByTranscript, threeUTRsByTranscript
```

```
> txExons <- exonsBy(mm9KGChr9)  
> txIntrons <- intronsByTranscript(mm9KGChr9)  
> txExons[6]
```

GRangesList of length 1

\$24313

GRanges with 3 ranges and 3 elementMetadata values

	seqnames	ranges	strand	exon_id	exon_name
	<Rle>	<IRanges>	<Rle>	<integer>	<character>
[1]	chr9	[3335473, 3335594]	+	117005	NA
[2]	chr9	[3338456, 3338591]	+	117006	NA
[3]	chr9	[3343015, 3343608]	+	117007	NA

exon\_rank

<integer>

```
[1]      1  
[2]      2  
[3]      3
```

# Overlapping with transcripts

## Methods

`findOverlaps, countOverlaps, match, %in%, subsetByOverlaps`

## Usage

```
> findOverlaps(query, subject, maxgap = 0L, minoverlap = 1L,
+                 type = c("any", "start", "end"),
+                 select = c("all", "first"))
> help("findOverlaps, GRanges, GRangesList-method")

> grngs <- GRanges("chr9", gaps(ranges(txExons[[6]]))), "+")
> countOverlaps(grngs, tx)

[1] 4 4

> rbind(countOverlaps(grngs, txExons), countOverlaps(grngs, txIntrons))

 [,1] [,2]
[1,]    1    0
[2,]    4    4
```

# Outline

Introduction

Genomic Features

B\$genome

Resources

# *BSgenome* packages

```
> library(BSgenome)
> available.genomes()

[1] "BSgenome.Amellifera.BeeBase.assembly4"
[2] "BSgenome.Amellifera.UCSC.apiMel2"
[3] "BSgenome.Athaliana.TAIR.01222004"
[4] "BSgenome.Athaliana.TAIR.04232008"
[5] "BSgenome.Btaurus.UCSC.bosTau3"
[6] "BSgenome.Btaurus.UCSC.bosTau4"
[7] "BSgenome.Celegans.UCSC.ce2"
[8] "BSgenome.Cfamiliaris.UCSC.canFam2"
[9] "BSgenome.Dmelanogaster.UCSC.dm2"
[10] "BSgenome.Dmelanogaster.UCSC.dm3"
[11] "BSgenome.Drerio.UCSC.danRer5"
[12] "BSgenome.Ecoli.NCBI.20080805"
[13] "BSgenome.Ggallus.UCSC.galGal3"
[14] "BSgenome.Hsapiens.UCSC.hg17"
[15] "BSgenome.Hsapiens.UCSC.hg18"
[16] "BSgenome.Hsapiens.UCSC.hg19"
[17] "BSgenome.Mmusculus.UCSC.mm8"
[18] "BSgenome.Mmusculus.UCSC.mm9"
[19] "BSgenome.Ptroglobutes.UCSC.panTro2"
[20] "BSgenome.Rnorvegicus.UCSC.rn4"
[21] "BSgenome.Scerevisiae.UCSC.sacCer1"
[22] "BSgenome.Scerevisiae.UCSC.sacCer2"
```

# *BSgenome* class decomposition

## *BSgenome* slots

```
> getSlots("BSgenome")
```

source_url	seqnames	seqlengths
"character"	"character"	"integer"
mseqnames	seqs_pkgnname	seqs_dir
"character"	"character"	"character"
nmask_per_seq	masks_pkgnname	masks_dir
"integer"	"character"	"character"
injectSNPs_handler	.seqs_cache	.link_counts
"InjectSNPsHandler"	"environment"	"environment"
organism	species	provider
"character"	"character"	"character"
provider_version	release_date	release_name
"character"	"character"	"character"

## Notes

- ▶ .seqs\_cache and .link\_counts slots manage memory.
- ▶ seqs\_dir and masks\_dir slots specify storage location.

# *BSgenome* methods

## Sequence selection

[[, \$

## Subsequence selection

getSeq

## Accessors

length, names/seqnames, mseqnames, seqlengths, masknames,  
sourceUrl

## Matching

vmatchPattern, vcountPattern, vmatchPDict, vcountPDict,  
matchPWM, countPWM

## SNPs (Human only at this point)

injectSNPs, SNPlocs\_pkgnmae, SNPcount, SNPlocs

## *BSgenome* package without masks

```
> library(BSgenome.Scerevisiae.UCSC.sacCer2)
> Scerevisiae

Yeast genome
|
| organism: Saccharomyces cerevisiae (Yeast)
| provider: UCSC
| provider version: sacCer2
| release date: June 2008
| release name: SGD June 2008 sequence
|
| sequences (see '?seqnames'):
|   chrI      chrII     chrIII    chrIV      chrV      chrVI
|   chrVII    chrVIII   chrIX     chrX       chrXI     chrXII
|   chrXIII   chrXIV    chrXV     chrXVI     chrM      2micron
|
| (use the '$' or '[[' operator to access a given sequence)

> Scerevisiae$chrI

230208-letter "DNAString" instance
seq: CCACACCACACCCACACACACACACC...GGTGTGGTGTGGGTGTGGTGTGTGGG
```

## *BSgenome* package with masks

```
> library(BSgenome.Hsapiens.UCSC.hg19)
> Hsapiens$chr1
 249250621-letter "MaskedDNAString" instance (# for masking)
seq: #####...#####
masks:
  maskedwidth maskedratio active names
1     23970000  0.09616827   TRUE AGAPS
2           0  0.00000000   TRUE   AMB
3    114014472  0.45742904 FALSE    RM
4    1581889  0.00634658 FALSE    TRF
                                desc
1                         assembly gaps
2   intra-contig ambiguities (empty)
3                         RepeatMasker
4 Tandem Repeats Finder [period<=12]
all masks together:
  maskedwidth maskedratio
 138071094  0.5539448
all active masks together:
  maskedwidth maskedratio
 23970000  0.09616827
```

# Sequence information

## Operations that don't load sequences

```
> head(seqnames(Scerevisiae), 6)
[1] "chrI"   "chrII"  "chrIII" "chrIV"  "chrV"   "chrVI"
> head(seqlengths(Scerevisiae), 8)
chrI    chrII   chrIII   chrIV    chrV    chrVI   chrVII  chrVIII
230208  813178  316617  1531919  576869  270148  1090947 562643
```

## Operation that does

```
> sapply(head(seqnames(Scerevisiae), 8), function(i)
+         alphabetFrequency(Scerevisiae[[i]], baseOnly=TRUE))
          chrI   chrII  chrIII   chrIV    chrV    chrVI   chrVII  chrVIII
A      69826  249653  98657  476749  176531  82928  338319  174022
C      44646  157410  62359  289343  109828  52201  207776  109098
G      45765  154397  59639  291356  112313  52435  207451  107488
T      69971  251718  95962  474471  178197  82584  337401  172035
other     0       0       0       0       0       0       0       0
```

## Matches for single pattern across genome

```
> exclude <- setdiff(seqnames(Hsapiens), c("chr1", "chr2"))
> vcountPattern("ACYTANCAGT", Hsapiens,
+                 fixed = c(pattern = FALSE, subject = TRUE),
+                 exclude = exclude)

  seqname strand count
1   chr1      +  1546
2   chr1      -  1545
3   chr2      +  1722
4   chr2      -  1684

> patmatch <-
+ vmatchPattern("ACYTANCAGT", Hsapiens,
+                fixed = c(pattern = FALSE, subject = TRUE),
+                exclude = exclude, asRangedData = FALSE)
> head(patmatch, 3)
```

GRanges with 3 ranges and 0 elementMetadata values

	seqnames	ranges	strand	
	<Rle>	<IRanges>	<Rle>	
[1]	chr1	[ 361581, 361590]	+	
[2]	chr1	[1738000, 1738009]	+	
[3]	chr1	[1814381, 1814390]	+	

## Pattern dictionary (Microarray probes)

```
> library("hgu95av2probe")
> probes <- DNAStringSet(hgu95av2probe$sequence[1:100])
> head(probes, 10)
```

A DNAStringSet instance of length 10

	width	seq
[1]	25	TGGCTCCTGCTGAGGTCCCCTTCC
[2]	25	GGCTGTGAATT CCTGTACATATTTC
[3]	25	GCTTCAATTCCATTATGTTTAATG
[4]	25	GCCGTTTGACAGAGCATGCTCTGCG
[5]	25	TGACAGAGCATGCTCTGC GTTGTG
[6]	25	CTCTGC GTTGGTT ACCAGCT
[7]	25	GGTTTCACCAGCTTCTGCCCTCACA
[8]	25	TTCTGCCCTCACATGCACAGGGATT
[9]	25	CCTCACATGCACAGGGATTAAACAA
[10]	25	TCCTTGGTACTCTGCCCTCCTGTCA

## Count matches for multiple patterns across genome

```
> counts <- vcountPDict(probes, Hsapiens, exclude = exclude)
> head(counts, 5)

DataFrame with 5 rows and 4 columns
  seqname strand    index count
  <Rle>   <Rle> <integer> <Rle>
1 chr1      +        1      0
2 chr1      +        2      0
3 chr1      +        3      0
4 chr1      +        4      0
5 chr1      +        5      0

> dim(counts)
[1] 400   4

> whichMatch <- seqselect(counts$index, counts$count > 0)
> length(whichMatch)

[1] 15

> whichMatch
[1]  1  2  3  4  5  6  7  8  9 10 11 12 13 14 16

> matchedProbes <- probes[whichMatch]
```

## Find match locations for probes

	start	end	width	
[1]	113420812	113420836	25	[TGGCTCCTGCTGAGGTCCCCTTCC]
[2]	113420842	113420866	25	[GGCTGTGAATTCCCTGTACATATTC]
[3]	113420884	113420908	25	[GCTTCAATTCCATTATGTTTAATG]
[4]	113420962	113420986	25	[GCCGTTTGACAGAGCATGCTCTGCG]
[5]	113420968	113420992	25	[TGACAGAGCATGCTCTGCCTTGTG]
[6]	113420980	113421004	25	[CTCTGCCTTGTGGTTTACCAAGCT]
[7]	113420992	113421016	25	[GGTTCACCAAGCTTCTGCCCTCACAC]
[8]	113421004	113421028	25	[TTCTGCCCTCACATGCACAGGGATT]
[9]	113421010	113421034	25	[CCTCACATGCACAGGGATTAAACAA]
[10]	113421082	113421106	25	[TCCTTGGTACTCTGCCCTCCTGTCA]
[11]	113421094	113421118	25	[TGCCCTCCTGTCACTAGTAGTGGCAGGA]
[12]	113421118	113421142	25	[ATCTATTGGCATATTGGGAGCTTC]
[13]	113421130	113421154	25	[ATTGGGAGCTTCTTAGAGGGATGA]
[14]	113421274	113421298	25	[AAGATTCTGGCAGTGTGGGATGGA]
[15]	113421340	113421364	25	[CAGCCTCCATGTTCAATTGTCTAC]

## SNP packages

```
> available.SNPs()

[1] "SNPlocs.Hsapiens.dbSNP.20071016"
[2] "SNPlocs.Hsapiens.dbSNP.20080617"
[3] "SNPlocs.Hsapiens.dbSNP.20090506"
[4] "SNPlocs.Hsapiens.dbSNP.20100427"

> SNPlocs_pkgname(Hsapiens)

NULL

> HsWithSNPs <-
+   injectSNPs(Hsapiens, "SNPlocs.Hsapiens.dbSNP.20090506")
> class(HsWithSNPs)

[1] "BSgenome"
attr(,"package")
[1] "BSgenome"

> SNPlocs_pkgname(HsWithSNPs)

[1] "SNPlocs.Hsapiens.dbSNP.20090506"
```

# SNP exploration

```
> SNPcount(HsWithSNPs)
```

```
chr1    chr2    chr3    chr4    chr5    chr6    chr7    chr8    chr9  
920233 933616 789121 798603 706109 760249 655873 612367 496064  
chr10   chr11   chr12   chr13   chr14   chr15   chr16   chr17   chr18  
583240 577300 558759 427010 365742 331501 354239 316396 322866  
chr19   chr20   chr21   chr22   chrX    chrY  
268235 323041 160580 187392 391414 6539
```

```
> alphabetFrequency(Hsapiens$chr1)
```

A	C	G	T	M	R	W
65570891	47024412	47016562	65668756	0	0	0
S	Y	K	V	H	D	B
O	O	O	O	O	O	O
N	-	+				
O	O	O				

```
> alphabetFrequency(HsWithSNPs$chr1)
```

A	C	G	T	M	R	W
65306157	46833464	46825359	65403357	40477	150327	40710
S	Y	K	V	H	D	B
31997	150117	41304	102527	125770	126323	102322
N	-	+				
410	O	O				

# Outline

Introduction

Genomic Features

BSgenome

Resources

# Resources

## *Bioconductor* Web site

- ▶ ‘*GenomicFeatures*’ and ‘*BSgenome*’ links.
- ▶ <http://bioconductor.org>
- ▶ ‘Installation’, ‘Software’, and ‘Mailing lists’ links.

## Help in *R*

- ▶ `help.start()` to view a help browser.
- ▶ `help(package = "BSgenome")`
- ▶ `?transcriptsBy`
- ▶ `browseVignettes("GenomicFeatures")`