Package 'idr2d'

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Title Irreproducible Discovery Rate for Genomic Interactions Data

Version 1.18.0

Description A tool to measure reproducibility between genomic experiments
that produce two-dimensional peaks (interactions between peaks), such
as ChIA-PET, HiChIP, and HiC. idr2d is an extension of the original
idr package, which is intended for (one-dimensional) ChIP-seq peaks.

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URL https://idr2d.mit.edu

Depends R (>= 3.6)

Imports dplyr (>= 0.7.6), futile.logger (>= 1.4.3), GenomeInfoDb (>=
1.14.0), GenomicRanges (>= 1.30), ggplot2 (>= 3.1.1),
grDevices, grid, idr (>= 1.2), IRanges (>= 2.18.0), magrittr
(>= 1.5), methods, reticulate (>= 1.13), scales (>= 1.0.0),
stats, stringr (>= 1.3.1), utils

Suggests DT (>= 0.4), htmltools (>= 0.3.6), knitr (>= 1.20), rmarkdown
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Contents

calculate_midpoint_distance1d

Distance between Midpoints of two Peaks

Description

Calculates the distance in nucleotides between the midpoints of two peaks.

Note: peaks must be on the same chromosome; start coordinate is always less than end coordinate

Usage

```
calculate_midpoint_distance1d(peak1_start, peak1_end, peak2_start, peak2_end)
```

Arguments

`peak1_start`	integer vector; genomic start coordinate(s) of peak in replicate 1
`peak1_end`	integer vector; genomic end coordinate(s) of peak in replicate 1
`peak2_start`	integer vector; genomic start coordinate(s) of peak in replicate 2
`peak2_end`	integer vector; genomic end coordinate(s) of peak in replicate 2

Value

positive integer vector; distances between peak pairs

Examples

```
# identical, zero distance
calculate_midpoint_distance1d(100, 120,
                              100, 120)

# centered, zero distance
calculate_midpoint_distance1d(100, 120,
                              90, 130)

# off by 10 per anchor
calculate_midpoint_distance1d(100, 120,
                              110, 130)

# vectorized example
calculate_midpoint_distance1d(c(100, 100, 100),
                              c(120, 120, 120),
                              c(100, 90, 110),
                              c(120, 130, 130))
```

---

calculate_midpoint_distance2d
Distance between Anchor Midpoints of two Interactions

Description

Calculates the distance in nucleotides between the anchor midpoints of two interactions, which is the sum of the distance between midpoints of anchor A in interaction 1 and anchor A in interaction 2, and the distance between midpoints of anchor B in interaction 1 and anchor B in interaction 2.

Note: all anchors must be on the same chromosome; start coordinate is always less than end coordinate

Usage

```
calculate_midpoint_distance2d(
  int1_anchor_a_start,
  int1_anchor_a_end,
  int1_anchor_b_start,
  int1_anchor_b_end,
  int2_anchor_a_start,
  int2_anchor_a_end,
  int2_anchor_b_start,
  int2_anchor_b_end
)
```

Arguments

```
int1_anchor_a_start
```
                    integer vector; genomic start coordinate(s) of anchor A in replicate 1 interaction

```
int1_anchor_a_end
```
                    integer vector; genomic end coordinate(s) of anchor A in replicate 1 interaction

```
int1_anchor_b_start
```
                    integer vector; genomic start coordinate(s) of anchor B in replicate 1 interaction

```
int1_anchor_b_end
```
                    integer vector; genomic end coordinate(s) of anchor B in replicate 1 interaction

```
int2_anchor_a_start
```
                    integer vector; genomic start coordinate(s) of anchor A in replicate 2 interaction

```
int2_anchor_a_end
```
                    integer vector; genomic end coordinate(s) of anchor A in replicate 2 interaction

```
int2_anchor_b_start
```
                    integer vector; genomic start coordinate(s) of anchor B in replicate 2 interaction

```
int2_anchor_b_end
```
                    integer vector; genomic end coordinate(s) of anchor B in replicate 2 interaction

Value

positive integer vector; distances between interaction pairs

Examples

```
# identical, zero distance
calculate_midpoint_distance2d(100, 120, 240, 260,
                              100, 120, 240, 260)

# centered, zero distance
calculate_midpoint_distance2d(100, 120, 240, 260,
                              90, 130, 230, 270)

# off by 10 per anchor
calculate_midpoint_distance2d(100, 120, 240, 250,
                              110, 130, 230, 240)
```

```
# off by 10 (anchor B only)
calculate_midpoint_distance2d(100, 120, 240, 250,
                               90, 130, 250, 260)

# vectorized example
calculate_midpoint_distance2d(c(100, 100, 100, 100),
                               c(120, 120, 120, 120),
                               c(240, 240, 240, 240),
                               c(260, 260, 250, 250),
                               c(100, 90, 110, 90),
                               c(120, 130, 130, 130),
                               c(240, 230, 230, 250),
                               c(260, 270, 240, 260))
```

---

calculate_relative_overlap1d

Relative Anchor Overlap of two Peaks

Description

Calculates the overlap between anchor A of interaction 1 and anchor A of interaction 2, as well as anchor B of interaction 1 and anchor B of interaction 2. The overlap (in nucleotides) is then normalized by the length of the anchors.

Usage

```
calculate_relative_overlap1d(peak1_start, peak1_end, peak2_start, peak2_end)
```

Arguments

peak1_start   integer vector; genomic start coordinate(s) of peak in replicate 1

peak1_end   integer vector; genomic end coordinate(s) of peak in replicate 1

peak2_start   integer vector; genomic start coordinate(s) of peak in replicate 2

peak2_end   integer vector; genomic end coordinate(s) of peak in replicate 2

Value

numeric vector; relative overlaps between peak pairs

Examples

```
# 100% overlap
calculate_relative_overlap1d(100, 120,
                             100, 120)

# 50% overlap
```

```
calculate_relative_overlap1d(100, 120,
                             100, 110)

# negative overlap
calculate_relative_overlap1d(100, 120,
                             130, 140)

# larger negative overlap
calculate_relative_overlap1d(100, 120,
                             200, 220)

# vectorized example
calculate_relative_overlap1d(c(100, 100, 100, 100),
                             c(120, 120, 120, 120),
                             c(100, 100, 130, 200),
                             c(120, 110, 140, 220))
```

---

calculate_relative_overlap2d

Relative Anchor Overlap of two Interactions

Description

Calculates the overlap between anchor A of interaction 1 and anchor A of interaction 2, as well as anchor B of interaction 1 and anchor B of interaction 2. The overlap (in nucleotides) is then normalized by the length of the anchors.

Note: anchors A and B of the same interaction have to be on the same chromosome; start coordinate is always less than end coordinate

Usage

```
calculate_relative_overlap2d(
  int1_anchor_a_start,
  int1_anchor_a_end,
  int1_anchor_b_start,
  int1_anchor_b_end,
  int2_anchor_a_start,
  int2_anchor_a_end,
  int2_anchor_b_start,
  int2_anchor_b_end
)
```

Arguments

int1_anchor_a_start
                integer vector; genomic start coordinate(s) of anchor A in replicate 1 interaction

int1_anchor_a_end
                integer vector; genomic end coordinate(s) of anchor A in replicate 1 interaction

int1_anchor_b_start

> integer vector; genomic start coordinate(s) of anchor B in replicate 1 interaction

int1_anchor_b_end

> integer vector; genomic end coordinate(s) of anchor B in replicate 1 interaction

int2_anchor_a_start

> integer vector; genomic start coordinate(s) of anchor A in replicate 2 interaction

int2_anchor_a_end

> integer vector; genomic end coordinate(s) of anchor A in replicate 2 interaction

int2_anchor_b_start

> integer vector; genomic start coordinate(s) of anchor B in replicate 2 interaction

int2_anchor_b_end

> integer vector; genomic end coordinate(s) of anchor B in replicate 2 interaction

Value

numeric vector; relative overlaps between interaction pairs

Examples

```
# 100% overlap
calculate_relative_overlap2d(100, 120, 240, 260,
                             100, 120, 240, 260)

# 50% overlap
calculate_relative_overlap2d(100, 120, 240, 250,
                             100, 110, 240, 260)

# negative overlap
calculate_relative_overlap2d(100, 120, 240, 250,
                             130, 140, 260, 280)

# larger negative overlap
calculate_relative_overlap2d(100, 120, 240, 250,
                             200, 220, 340, 350)

# vectorized example
calculate_relative_overlap2d(c(100, 100, 100, 100),
                             c(120, 120, 120, 120),
                             c(240, 240, 240, 240),
                             c(260, 250, 250, 250),
                             c(100, 100, 130, 200),
                             c(120, 110, 140, 220),
                             c(240, 240, 260, 340),
                             c(260, 260, 280, 350))
```

---

chiapet   *Example Genomic Interaction Data Set*

Description

This object contains genomic interactions on chromosomes 1 to 5, which could be the results of
Hi-C or ChIA-PET experiments, done in duplicates.

Usage

```
chiapet
```

Format

A list with two components, the data frames `rep1_df` and `rep2_df`, which have the following seven
columns:

column 1:	chr_a	character; genomic location of anchor A - chromosome (e.g., "chr3")
column 2:	start_a	integer; genomic location of anchor A - start coordinate
column 3:	end_a	integer; genomic location of anchor A - end coordinate
column 4:	chr_b	character; genomic location of anchor B - chromosome (e.g., "chr3")
column 5:	start_b	integer; genomic location of anchor B - start coordinate
column 6:	end_b	integer; genomic location of anchor B - end coordinate
column 7:	fdr	numeric; False Discovery Rate - significance of interaction

---

chipseq   *Example Genomic Peak Data Set*

Description

This object contains genomic peaks from two replicate ChIP-seq experiments.

Usage

```
chipseq
```

Format

A list with two components, the data frames `rep1_df` and `rep2_df`, which have the following four
columns:

column 1:	chr	character; genomic location of peak - chromosome (e.g., "chr3")
column 2:	start	integer; genomic location of peak - start coordinate
column 3:	end	integer; genomic location of peak - end coordinate
column 4:	value	numeric; heuristic used to rank the peaks

---

determine_anchor_overlap

Identifies Overlapping Anchors

Description

Identifies all overlapping anchor pairs (m:n mapping).

Usage

```
determine_anchor_overlap(rep1_anchor, rep2_anchor, max_gap = -1L)
```

Arguments

rep1_anchor   data frame with the following columns:

column 1:	chr	character; genomic location of anchor in replicate 1 - chromosome (e.g., ″chr3″)
column 2:	start	integer; genomic location of anchor in replicate 1 - start coordinate
column 3:	end	integer; genomic location of anchor in replicate 1 - end coordinate

rep2_anchor   data frame with the following columns:

column 1:	chr	character; genomic location of anchor in replicate 2 - chromosome (e.g., ″chr3″)
column 2:	start	integer; genomic location of anchor in replicate 2 - start coordinate
column 3:	end	integer; genomic location of anchor in replicate 2 - end coordinate

max_gap   integer; maximum gap in nucleotides allowed between two anchors for them to be considered as overlapping (defaults to -1, i.e., overlapping anchors)

Value

A data frame containing overlapping anchor pairs with the following columns:

column 1:	rep1_idx	anchor index in data frame rep1_anchor
column 2:	rep2_idx	anchor index in data frame rep2_anchor

Examples

```
rep1_df <- idr2d:::chiapet$rep1_df
rep2_df <- idr2d:::chiapet$rep2_df

rep1_anchor_a <- data.frame(chr = rep1_df[, 1],
                            start = rep1_df[, 2],
                            end = rep1_df[, 3])
rep2_anchor_a <- data.frame(chr = rep2_df[, 1],
```

```
                            start = rep2_df[, 2],
                            end = rep2_df[, 3])

anchor_a_overlap <- determine_anchor_overlap(rep1_anchor_a, rep2_anchor_a)
```

---

draw_hic_contact_map   *Create Hi-C contact map*

Description

Creates Hi-C contact maps to visualize the results of estimate_idr2d_hic.

Usage

```
draw_hic_contact_map(
  df,
  idr_cutoff = NULL,
  chromosome = NULL,
  start_coordinate = NULL,
  end_coordinate = NULL,
  title = NULL,
  values_normalized = FALSE,
  log_values = TRUE
)
```

Arguments

df   output of estimate_idr2d_hic, a data frame with the following columns:

column 1:	interaction	character; genomic location of interaction block (e.g., "chr1:204940000-204940000")
column 2:	value	numeric; p-value, FDR, or heuristic used to rank the interactions
column 3:	"rep_value"	numeric; value of corresponding replicate interaction
column 4:	"rank"	integer; rank of the interaction, established by value column, ascending order
column 5:	"rep_rank"	integer; rank of corresponding replicate interaction
column 6:	"idr"	integer; IDR of the block and the corresponding block in the other replicate

idr_cutoff   numeric; only show blocks with IDR < idr_cutoff, shows all blocks by default

chromosome   character; chromsome name or list of chromosome names to be analyzed, e.g.,
                    UCSC chromosome 1, "chr1", defaults to all chromosomes (chromosome =
                    NULL)

start_coordinate
                    integer; only show contact map window between "start_coordinate" and
                    "end_coordinate", by default shows entire chromosome

end_coordinate   integer; only show contact map window between "start_coordinate" and
                    "end_coordinate", by default shows entire chromosome

```
title            character; plot title
values_normalized
                 logical; are read counts in value column raw or normalized? Defaults to FALSE
log_values       logical; log-transform value column? Defaults to TRUE
```

Value

ggplot2 object; Hi-C contact map

Examples

```
idr_results_df <- estimate_idr2d_hic(idr2d:::hic$rep1_df,
                                     idr2d:::hic$rep2_df)
draw_hic_contact_map(idr_results_df, idr_cutoff = 0.05, chromosome = "chr1")
```

---

draw_idr_distribution_histogram

Create histogram of IDR values

Description

Creates diagnostic plots to visualize the results of estimate_idr.

Usage

```
draw_idr_distribution_histogram(
  df,
  remove_na = TRUE,
  xlab = "IDR",
  ylab = "density",
  title = "IDR value distribution"
)
```

Arguments

df   part of output of estimate_idr, a data frame with at least the following named
               columns:

        idr   IDR of the peak and the corresponding peak in the other replicate.

remove_na   logical; should NA values be removed?

xlab   character; x axis label

ylab   character; y axis label

title   character; plot title

Value

ggplot2 object; IDR distribution histogram

Examples

```
idr_results <- estimate_idr1d(idr2d:::chipseq$rep1_df,
                              idr2d:::chipseq$rep2_df,
                              value_transformation = "log")
draw_idr_distribution_histogram(idr_results$rep1_df)
```

---

draw_rank_idr_scatterplot

Create scatterplot of IDR values

Description

Creates diagnostic plots to visualize the results of estimate_idr.

Usage

```
draw_rank_idr_scatterplot(
  df,
  remove_na = TRUE,
  xlab = "rank in replicate 1",
  ylab = "rank in replicate 2",
  log_idr = FALSE,
  title = "rank - IDR dependence",
  color_gradient = c("rainbow", "default"),
  alpha = 1,
  max_points_shown = 2500
)
```

Arguments

df	part of output of estimate_idr, a data frame with at least the following named columns:

	rank	integer; rank of the peak, established by value column, ascending order
	rep_rank	integer; rank of corresponding replicate peak.
	idr	IDR of the peak and the corresponding peak in the other replicate.

remove_na	logical; should NA values be removed?
xlab	character; x axis label
ylab	character; y axis label
log_idr	logical; use logarithmized IDRs for colors to better distinguish highly significant IDRs

title	character; plot title
color_gradient	character; either "rainbow" or "default"
alpha	numeric; transparency of dots, from 0.0 - 1.0, where 1.0 is completely opaque; default is 1.0
max_points_shown	
	integer; default is 2500

Value

ggplot2 object; IDR rank scatterplot

Examples

```
idr_results <- estimate_idr1d(idr2d:::chipseq$rep1_df,
                              idr2d:::chipseq$rep2_df,
                              value_transformation = "log")
draw_rank_idr_scatterplot(idr_results$rep1_df)
```

---

draw_value_idr_scatterplot

Create scatterplot of IDR values

Description

Creates diagnostic plots to visualize the results of [estimate_idr](#).

Usage

```
draw_value_idr_scatterplot(
  df,
  remove_na = TRUE,
  remove_outliers = TRUE,
  xlab = "transformed value in replicate 1",
  ylab = "transformed value in replicate 2",
  log_axes = FALSE,
  log_idr = FALSE,
  title = "value - IDR dependence",
  color_gradient = c("rainbow", "default"),
  alpha = 1,
  max_points_shown = 2500
)
```

Arguments

df
: part of output of [estimate_idr](), a data frame with at least the following named columns:

value	numeric; p-value, FDR, or heuristic used to rank the peaks
rep_value	numeric; value of corresponding replicate peak
idr	IDR of the peak and the corresponding peak in the other replicate.

remove_na
: logical; should NA values be removed?

remove_outliers
: logical; removes extreme data points

xlab
: character; x axis label

ylab
: character; y axis label

log_axes
: logical; show logarithmized values from replicate 1 and 2 (default value is FALSE)

log_idr
: logical; use logarithmized IDRs for colors to better distinguish highly significant IDRs (default value is FALSE)

title
: character; plot title

color_gradient
: character; either "rainbow" or "default"

alpha
: numeric; transparency of dots, from 0.0 - 1.0, where 1.0 is completely opaque; default is 1.0

max_points_shown
: integer; default is 2500

Value

ggplot2 object; IDR value scatterplot

Examples

```
idr_results <- estimate_idr1d(idr2d:::chipseq$rep1_df,
                              idr2d:::chipseq$rep2_df,
                              value_transformation = "log")
draw_value_idr_scatterplot(idr_results$rep1_df)
```

---

establish_bijection	*Finds One-to-One Correspondence between Peaks or interactions from Replicate 1 and 2*

Description

This method establishes a bijective assignment between observations (genomic peaks in case of ChIP-seq, genomic interactions in case of ChIA-PET, HiChIP, and Hi-C) from replicate 1 and 2. An observation in replicate 1 is assigned to an observation in replicate 2 if and only if (1) the observation loci in both replicates overlap (or the gap between them is less than or equal to max_gap), and (2) there is no other observation in replicate 2 that overlaps with the observation in replicate 1 and has a lower *ambiguity resolution value*.

Usage

```
establish_bijection(
  rep1_df,
  rep2_df,
  analysis_type = c("IDR1D", "IDR2D"),
  ambiguity_resolution_method = c("overlap", "midpoint", "value"),
  max_gap = -1L
)
```

Arguments

rep1_df	data frame of observations (i.e., genomic peaks or genomic interactions) of replicate 1. If analysis_type is IDR1D, the columns of rep1_df are described in establish_bijection1d, otherwise in establish_bijection2d
rep2_df	data frame of observations (i.e., genomic peaks or genomic interactions) of replicate 2. Same columns as rep1_df.
analysis_type	"IDR2D" for genomic interaction data sets, "IDR1D" for genomic peak data sets
ambiguity_resolution_method	
	defines how ambiguous assignments (when one interaction or peak in replicate 1 overlaps with multiple interactions or peaks in replicate 2 or vice versa) are resolved. For available methods, see establish_overlap1d or establish_overlap2d, respectively.
max_gap	integer; maximum gap in nucleotides allowed between two anchors for them to be considered as overlapping (defaults to -1, i.e., overlapping anchors)

Value

See establish_bijection1d or establish_bijection2d, respectively.

Examples

```
rep1_df <- idr2d:::chipseq$rep1_df
rep1_df$value <- preprocess(rep1_df$value, "log")

rep2_df <- idr2d:::chipseq$rep2_df
rep2_df$value <- preprocess(rep2_df$value, "log")

mapping <- establish_bijection(rep1_df, rep2_df, analysis_type = "IDR1D")
```

---

establish_bijection1d   *Finds One-to-One Correspondence between Peaks from Replicate 1*
                        and 2

Description

This method establishes a bijective assignment between peaks from replicate 1 and 2. A peak in
replicate 1 is assigned to a peak in replicate 2 if and only if (1) they overlap (or the gap between the
peaks is less than or equal to max_gap), and (2) there is no other peak in replicate 2 that overlaps
with the peak in replicate 1 and has a lower *ambiguity resolution value*.

Usage

```
establish_bijection1d(
  rep1_df,
  rep2_df,
  ambiguity_resolution_method = c("overlap", "midpoint", "value"),
  max_gap = -1L
)
```

Arguments

rep1_df		data frame of observations (i.e., genomic peaks) of replicate 1, with at least the following columns (position of columns matter, column names are irrelevant):

column 1:	chr	character; genomic location of peak - chromosome (e.g., "chr3")
column 2:	start	integer; genomic location of peak - start coordinate
column 3:	end	integer; genomic location of peak - end coordinate
column 4:	value	numeric; p-value, FDR, or heuristic used to rank the interactions

rep2_df		data frame of observations (i.e., genomic peaks) of replicate 2, with the following columns (position of columns matter, column names are irrelevant):

column 1:	chr	character; genomic location of peak - chromosome (e.g., "chr3")
column 2:	start	integer; genomic location of peak - start coordinate
column 3:	end	integer; genomic location of peak - end coordinate
column 4:	value	numeric; p-value, FDR, or heuristic used to rank the interactions

ambiguity_resolution_method

defines how ambiguous assignments (when one interaction in replicate 1 overlaps with multiple interactions in replicate 2 or vice versa) are resolved. Available methods:

"value"	interactions are prioritized by ascending or descending value column (see sorting_direction), e.g., if two in
"overlap"	the interaction pair is chosen which has the highest relative overlap, i.e., overlap in nucleotides of replicate 1 in
"midpoint"	the interaction pair is chosen which has the smallest distance between their anchor midpoints, i.e., distance from

max_gap	integer; maximum gap in nucleotides allowed between two anchors for them to be considered as overlapping (defaults to -1, i.e., overlapping anchors)

Value

Data frames `rep1_df` and `rep2_df` with the following columns:

column 1:	chr	character; genomic location of peak - chromosome (e.g., `"chr3"`)
column 2:	start	integer; genomic location of peak - start coordinate
column 3:	end	integer; genomic location of peak - end coordinate
column 4:	value	numeric; p-value, FDR, or heuristic used to rank the peaks
column 5:	rep_value	numeric; value of corresponding replicate peak. If no corresponding peak was found, rep_value is
column 6:	rank	integer; rank of the peak, established by value column, ascending order
column 7:	rep_rank	integer; rank of corresponding replicate peak. If no corresponding peak was found, rep_rank is set
column 8:	idx	integer; peak index, primary key
column 9:	rep_idx	integer; specifies the index of the corresponding peak in the other replicate (foreign key). If no corr

Examples

```
rep1_df <- idr2d:::chipseq$rep1_df
rep1_df$value <- preprocess(rep1_df$value, "log")

rep2_df <- idr2d:::chipseq$rep2_df
rep2_df$value <- preprocess(rep2_df$value, "log")

mapping <- establish_bijection1d(rep1_df, rep2_df)
```

---

establish_bijection2d *Finds One-to-One Correspondence between Interactions from Replicate 1 and 2*

Description

This method establishes a bijective assignment between interactions from replicate 1 and 2. An interaction in replicate 1 is assigned to an interaction in replicate 2 if and only if (1) both anchors of the interactions overlap (or the gap between anchor A/B in replicate 1 and 2 is less than or equal to `max_gap`), and (2) there is no other interaction in replicate 2 that overlaps with the interaction in replicate 1 and has a lower *ambiguity resolution value*.

Usage

```
establish_bijection2d(
  rep1_df,
  rep2_df,
  ambiguity_resolution_method = c("overlap", "midpoint", "value"),
  max_gap = -1L
)
```

Arguments

rep1_df		data frame of observations (i.e., genomic interactions) of replicate 1, with at least the following columns (position of columns matter, column names are irrelevant):

column 1:	chr_a	character; genomic location of anchor A - chromosome (e.g., "chr3")
column 2:	start_a	integer; genomic location of anchor A - start coordinate
column 3:	end_a	integer; genomic location of anchor A - end coordinate
column 4:	chr_b	character; genomic location of anchor B - chromosome (e.g., "chr3")
column 5:	start_b	integer; genomic location of anchor B - start coordinate
column 6:	end_b	integer; genomic location of anchor B - end coordinate
column 7:	value	numeric; p-value, FDR, or heuristic used to rank the interactions

rep2_df		data frame of observations (i.e., genomic interactions) of replicate 2, with the following columns (position of columns matter, column names are irrelevant):

column 1:	chr_a	character; genomic location of anchor A - chromosome (e.g., "chr3")
column 2:	start_a	integer; genomic location of anchor A - start coordinate
column 3:	end_a	integer; genomic location of anchor A - end coordinate
column 4:	chr_b	character; genomic location of anchor B - chromosome (e.g., "chr3")
column 5:	start_b	integer; genomic location of anchor B - start coordinate
column 6:	end_b	integer; genomic location of anchor B - end coordinate
column 7:	value	numeric; p-value, FDR, or heuristic used to rank the interactions

ambiguity_resolution_method

       defines how ambiguous assignments (when one interaction in replicate 1 overlaps with multiple interactions in replicate 2 or vice versa) are resolved. Available methods:

"value"	interactions are prioritized by ascending or descending value column (see sorting_direction), e.g., if two in
"overlap"	the interaction pair is chosen which has the highest relative overlap, i.e., overlap in nucleotides of replicate 1 in
"midpoint"	the interaction pair is chosen which has the smallest distance between their anchor midpoints, i.e., distance from

max_gap	integer; maximum gap in nucleotides allowed between two anchors for them to be considered as overlapping (defaults to -1, i.e., overlapping anchors)

Value

Data frames rep1_df and rep2_df with the following columns:

column 1:	chr_a	character; genomic location of anchor A - chromosome (e.g., "chr3")
column 2:	start_a	integer; genomic location of anchor A - start coordinate
column 3:	end_a	integer; genomic location of anchor A - end coordinate
column 4:	chr_b	character; genomic location of anchor B - chromosome (e.g., "chr3")
column 5:	start_b	integer; genomic location of anchor B - start coordinate
column 6:	end_b	integer; genomic location of anchor B - end coordinate
column 7:	value	numeric; p-value, FDR, or heuristic used to rank the interactions
column 8:	"rep_value"	numeric; value of corresponding replicate interaction. If no corresponding interaction was foun

column 9: "rank" integer; rank of the interaction, established by value column, ascending order
column 10: "rep_rank" integer; rank of corresponding replicate interaction. If no corresponding interaction was found,
column 11: "idx" integer; interaction index, primary key
column 12: "rep_idx" integer; specifies the index of the corresponding interaction in the other replicate (foreign key).

Examples

```
rep1_df <- idr2d:::chiapet$rep1_df
rep1_df$fdr <- preprocess(rep1_df$fdr, "log_additive_inverse")

rep2_df <- idr2d:::chiapet$rep2_df
rep2_df$fdr <- preprocess(rep2_df$fdr, "log_additive_inverse")

mapping <- establish_bijection2d(rep1_df, rep2_df)
```

---

establish_overlap1d *Establish m:n Mapping Between Peaks from Replicate 1 and 2*

Description

This method returns all overlapping interactions between two replicates. For each pair of overlapping interactions, the *ambiguity resolution value* (ARV) is calculated, which helps to reduce the m:n mapping to a 1:1 mapping. The semantics of the ARV depend on the specified ambiguity_resolution_method, but in general interaction pairs with lower ARVs have priority over interaction pairs with higher ARVs when the bijective mapping is established.

Usage

```
establish_overlap1d(
  rep1_df,
  rep2_df,
  ambiguity_resolution_method = c("overlap", "midpoint", "value"),
  max_gap = -1L
)
```

Arguments

rep1_df data frame of observations (i.e., genomic peaks) of replicate 1, with at least the following columns (position of columns matter, column names are irrelevant):

column 1: chr character; genomic location of peak - chromosome (e.g., "chr3")
column 2: start integer; genomic location of peak - start coordinate
column 3: end integer; genomic location of peak - end coordinate
column 4: value numeric; p-value, FDR, or heuristic used to rank the interactions

rep2_df   data frame of observations (i.e., genomic peaks) of replicate 2, with the follow-
                 ing columns (position of columns matter, column names are irrelevant):

    column 1:   chr   character; genomic location of peak - chromosome (e.g., "chr3")
    column 2:   start   integer; genomic location of peak - start coordinate
    column 3:   end   integer; genomic location of peak - end coordinate
    column 4:   value   numeric; p-value, FDR, or heuristic used to rank the interactions

ambiguity_resolution_method
                 defines how ambiguous assignments (when one interaction in replicate 1 over-
                 laps with multiple interactions in replicate 2 or vice versa) are resolved. Avail-
                 able methods:

   "value"   interactions are prioritized by ascending or descending value column (see sorting_direction), e.g., if two i
 "overlap"   the interaction pair is chosen which has the highest relative overlap, i.e., overlap in nucleotides of replicate 1 in
"midpoint"   the interaction pair is chosen which has the smallest distance between their anchor midpoints, i.e., distance fro

max_gap   integer; maximum gap in nucleotides allowed between two anchors for them to
                 be considered as overlapping (defaults to -1, i.e., overlapping anchors)

Value

data frame with the following columns:

column 1:   rep1_idx   index of interaction in replicate 1 (i.e., row index in rep1_df)
column 2:   rep2_idx   index of interaction in replicate 2 (i.e., row index in rep2_df)
column 3:   arv   ambiguity resolution value used turn m:n mapping into 1:1 mapping. Interaction pairs with lower ar

Examples

```
rep1_df <- idr2d:::chipseq$rep1_df
rep1_df$value <- preprocess(rep1_df$value, "log_additive_inverse")

rep2_df <- idr2d:::chipseq$rep2_df
rep2_df$value <- preprocess(rep2_df$value, "log_additive_inverse")

# shuffle to break preexisting order
rep1_df <- rep1_df[sample.int(nrow(rep1_df)), ]
rep2_df <- rep2_df[sample.int(nrow(rep2_df)), ]

# sort by value column
rep1_df <- dplyr::arrange(rep1_df, value)
rep2_df <- dplyr::arrange(rep2_df, value)

pairs_df <- establish_overlap1d(rep1_df, rep2_df)
```

establish_overlap2d	*Establish m:n mapping between interactions from replicate 1 and 2*

Description

This method returns all overlapping interactions between two replicates. For each pair of overlapping interactions, the *ambiguity resolution value* (ARV) is calculated, which helps to reduce the m:n mapping to a 1:1 mapping. The semantics of the ARV depend on the specified `ambiguity_resolution_method`, but in general interaction pairs with lower ARVs have priority over interaction pairs with higher ARVs when the bijective mapping is established.

Usage

```
establish_overlap2d(
  rep1_df,
  rep2_df,
  ambiguity_resolution_method = c("overlap", "midpoint", "value"),
  max_gap = -1L
)
```

Arguments

rep1_df		data frame of observations (i.e., genomic interactions) of replicate 1, with at least the following columns (position of columns matter, column names are irrelevant):

column 1:	chr_a	character; genomic location of anchor A - chromosome (e.g., "chr3")
column 2:	start_a	integer; genomic location of anchor A - start coordinate
column 3:	end_a	integer; genomic location of anchor A - end coordinate
column 4:	chr_b	character; genomic location of anchor B - chromosome (e.g., "chr3")
column 5:	start_b	integer; genomic location of anchor B - start coordinate
column 6:	end_b	integer; genomic location of anchor B - end coordinate
column 7:	value	numeric; p-value, FDR, or heuristic used to rank the interactions

rep2_df		data frame of observations (i.e., genomic interactions) of replicate 2, with the following columns (position of columns matter, column names are irrelevant):

column 1:	chr_a	character; genomic location of anchor A - chromosome (e.g., "chr3")
column 2:	start_a	integer; genomic location of anchor A - start coordinate
column 3:	end_a	integer; genomic location of anchor A - end coordinate
column 4:	chr_b	character; genomic location of anchor B - chromosome (e.g., "chr3")
column 5:	start_b	integer; genomic location of anchor B - start coordinate
column 6:	end_b	integer; genomic location of anchor B - end coordinate
column 7:	value	numeric; p-value, FDR, or heuristic used to rank the interactions

ambiguity_resolution_method

defines how ambiguous assignments (when one interaction in replicate 1 over-
laps with multiple interactions in replicate 2 or vice versa) are resolved. Avail-
able methods:

"value"   interactions are prioritized by ascending or descending value column (see `sorting_direction`), e.g., if two i
"overlap" the interaction pair is chosen which has the highest relative overlap, i.e., overlap in nucleotides of replicate 1 in
"midpoint" the interaction pair is chosen which has the smallest distance between their anchor midpoints, i.e., distance from

max_gap   integer; maximum gap in nucleotides allowed between two anchors for them to
                 be considered as overlapping (defaults to -1, i.e., overlapping anchors)

Value

data frame with the following columns:

column 1: rep1_idx index of interaction in replicate 1 (i.e., row index in `rep1_df`)
column 2: rep2_idx index of interaction in replicate 2 (i.e., row index in `rep2_df`)
column 3: arv   ambiguity resolution value used turn m:n mapping into 1:1 mapping. Interaction pairs with lower ar

Examples

```
rep1_df <- idr2d:::chiapet$rep1_df
rep1_df$fdr <- preprocess(rep1_df$fdr, "log_additive_inverse")

rep2_df <- idr2d:::chiapet$rep2_df
rep2_df$fdr <- preprocess(rep2_df$fdr, "log_additive_inverse")

# shuffle to break preexisting order
rep1_df <- rep1_df[sample.int(nrow(rep1_df)), ]
rep2_df <- rep2_df[sample.int(nrow(rep2_df)), ]

# sort by value column
rep1_df <- dplyr::arrange(rep1_df, rep1_df$fdr)
rep2_df <- dplyr::arrange(rep2_df, rep2_df$fdr)

pairs_df <- establish_overlap2d(rep1_df, rep2_df)
```

---

estimate_idr   *Estimates IDR for Genomic Peaks or Genomic Interactions*

Description

Estimates IDR for Genomic Peaks or Genomic Interactions

Usage

```
estimate_idr(
  rep1_df,
  rep2_df,
  analysis_type = "IDR2D",
  value_transformation = c("identity", "additive_inverse", "multiplicative_inverse",
    "log", "log_additive_inverse"),
  ambiguity_resolution_method = c("overlap", "midpoint", "value"),
  remove_nonstandard_chromosomes = TRUE,
  max_factor = 1.5,
  jitter_factor = 1e-04,
  max_gap = -1L,
  mu = 0.1,
  sigma = 1,
  rho = 0.2,
  p = 0.5,
  eps = 0.001,
  max_iteration = 30,
  local_idr = TRUE
)
```

Arguments

rep1_df   data frame of observations (i.e., genomic peaks or genomic interactions) of replicate 1. If analysis_type is IDR1D, the columns of rep1_df are described in [establish_bijection1d](#), otherwise in [establish_bijection2d](#)

rep2_df   data frame of observations (i.e., genomic peaks or genomic interactions) of replicate 2. Same columns as rep1_df.

analysis_type   "IDR2D" for genomic interaction data sets, "IDR1D" for genomic peak data sets

value_transformation

the values in x have to be transformed in a way such that when ordered in descending order, more significant interactions end up on top of the list. If the values in x are p-values, "log_additive_inverse" is recommended. The following transformations are supported:

"identity"	no transformation is performed on x
"additive_inverse"	x. = -x
"multiplicative_inverse"	x. = 1 / x
"log"	x. = log(x). Note: zeros are replaced by .Machine$double.xmin
"log_additive_inverse"	x. = -log(x), recommended if x are p-values. Note: zeros are replaced by .Machine$double

either "ascending" (more significant interactions have lower value in value column) or "descending" (more significant interactions have higher value in value column)

ambiguity_resolution_method

defines how ambiguous assignments (when one interaction or peak in replicate 1 overlaps with multiple interactions or peaks in replicate 2 or vice versa) are re-

solved. For available methods, see `establish_overlap1d` or `establish_overlap2d`, respectively.

remove_nonstandard_chromosomes

      removes peaks and interactions containing genomic locations on non-standard chromosomes using `keepStandardChromosomes` (default is TRUE)

max_factor   numeric; controls the replacement values for Inf and `-Inf`. Inf are replaced by `max(x) * max_factor` and `-Inf` are replaced by `min(x) / max_factor`.

jitter_factor   numeric; controls the magnitude of the noise that is added to x. This is done to break ties in x. Set `jitter_factor = NULL` for no jitter.

max_gap   integer; maximum gap in nucleotides allowed between two anchors for them to be considered as overlapping (defaults to -1, i.e., overlapping anchors)

mu   a starting value for the mean of the reproducible component.

sigma   a starting value for the standard deviation of the reproducible component.

rho   a starting value for the correlation coefficient of the reproducible component.

p   a starting value for the proportion of reproducible component.

eps   Stopping criterion. Iterations stop when the increment of log-likelihood is < eps*log-likelihood, Default=0.001.

max_iteration integer; maximum number of iterations for IDR estimation (defaults to 30)

local_idr   see `est.IDR`

Value

See `estimate_idr1d` or `estimate_idr2d`, respectively.

References

Q. Li, J. B. Brown, H. Huang and P. J. Bickel. (2011) Measuring reproducibility of high-throughput experiments. Annals of Applied Statistics, Vol. 5, No. 3, 1752-1779.

Examples

```
idr_results <- estimate_idr(idr2d:::chiapet$rep1_df,
                            idr2d:::chiapet$rep2_df,
                            analysis_type = "IDR2D",
                            value_transformation = "log_additive_inverse")
summary(idr_results)
```

estimate_idr1d *Estimates IDR for Genomic Peak Data*

Description

This method estimates Irreproducible Discovery Rates (IDR) for peaks in replicated ChIP-seq experiments.

Usage

```
estimate_idr1d(
  rep1_df,
  rep2_df,
  value_transformation = c("identity", "additive_inverse", "multiplicative_inverse",
    "log", "log_additive_inverse"),
  ambiguity_resolution_method = c("overlap", "midpoint", "value"),
  remove_nonstandard_chromosomes = TRUE,
  max_factor = 1.5,
  jitter_factor = 1e-04,
  max_gap = -1L,
  mu = 0.1,
  sigma = 1,
  rho = 0.2,
  p = 0.5,
  eps = 0.001,
  max_iteration = 30,
  local_idr = TRUE
)
```

Arguments

rep1_df   data frame of observations (i.e., genomic peaks) of replicate 1, with at least the
               following columns (position of columns matter, column names are irrelevant):

column 1:	chr	character; genomic location of peak - chromosome (e.g., "chr3")
column 2:	start	integer; genomic location of peak - start coordinate
column 3:	end	integer; genomic location of peak - end coordinate
column 4:	value	numeric; p-value, FDR, or heuristic used to rank the interactions

rep2_df   data frame of observations (i.e., genomic peaks) of replicate 2, with the following columns (position of columns matter, column names are irrelevant):

column 1:	chr	character; genomic location of peak - chromosome (e.g., "chr3")
column 2:	start	integer; genomic location of peak - start coordinate
column 3:	end	integer; genomic location of peak - end coordinate
column 4:	value	numeric; p-value, FDR, or heuristic used to rank the interactions

value_transformation

    the values in x have to be transformed in a way such that when ordered in descending order, more significant interactions end up on top of the list. If the values in x are p-values, "log_additive_inverse" is recommended. The following transformations are supported:

"identity"	no transformation is performed on x
"additive_inverse"	x. = -x
"multiplicative_inverse"	x. = 1 / x
"log"	x. = log(x). Note: zeros are replaced by .Machine$double.xmin
"log_additive_inverse"	x. = -log(x), recommended if x are p-values. Note: zeros are replaced by .Machine$doubl

    either "ascending" (more significant interactions have lower value in value column) or "descending" (more significant interactions have higher value in value column)

ambiguity_resolution_method

    defines how ambiguous assignments (when one interaction in replicate 1 overlaps with multiple interactions in replicate 2 or vice versa) are resolved. Available methods:

"value"	interactions are prioritized by ascending or descending value column (see sorting_direction), e.g., if two i
"overlap"	the interaction pair is chosen which has the highest relative overlap, i.e., overlap in nucleotides of replicate 1 in
"midpoint"	the interaction pair is chosen which has the smallest distance between their anchor midpoints, i.e., distance from

remove_nonstandard_chromosomes

    removes peaks containing genomic locations on non-standard chromosomes using [keepStandardChromosomes](#) (default is TRUE)

max_factor   numeric; controls the replacement values for Inf and -Inf. Inf are replaced by max(x) * max_factor and -Inf are replaced by min(x) / max_factor.

jitter_factor   numeric; controls the magnitude of the noise that is added to x. This is done to break ties in x. Set jitter_factor = NULL for no jitter.

max_gap   integer; maximum gap in nucleotides allowed between two anchors for them to be considered as overlapping (defaults to -1, i.e., overlapping anchors)

mu   a starting value for the mean of the reproducible component.

sigma   a starting value for the standard deviation of the reproducible component.

rho   a starting value for the correlation coefficient of the reproducible component.

p   a starting value for the proportion of reproducible component.

eps   Stopping criterion. Iterations stop when the increment of log-likelihood is < eps*log-likelihood, Default=0.001.

max_iteration   integer; maximum number of iterations for IDR estimation (defaults to 30)

local_idr   see [est.IDR](#)

Value

List with three components, (rep1_df, rep2_df, and analysis_type) containing the interactions
from input data frames rep1_df and rep2_df with the following additional columns:

column 1:	chr	character; genomic location of peak - chromosome (e.g., "chr3")
column 2:	start	integer; genomic location of peak - start coordinate
column 3:	end	integer; genomic location of peak - end coordinate
column 4:	value	numeric; p-value, FDR, or heuristic used to rank the peaks
column 5:	rep_value	numeric; value of corresponding replicate peak. If no corresponding peak was found, rep_value
column 6:	rank	integer; rank of the peak, established by value column, ascending order
column 7:	rep_rank	integer; rank of corresponding replicate peak. If no corresponding peak was found, rep_rank is s
column 8:	idx	integer; peak index, primary key
column 9:	rep_idx	integer; specifies the index of the corresponding peak in the other replicate (foreign key). If no co
column 10:	idr	IDR of the peak and the corresponding peak in the other replicate. If no corresponding peak was f

References

Q. Li, J. B. Brown, H. Huang and P. J. Bickel. (2011) Measuring reproducibility of high-throughput
experiments. Annals of Applied Statistics, Vol. 5, No. 3, 1752-1779.

Examples

```
idr_results <- estimate_idr1d(idr2d:::chipseq$rep1_df,
                              idr2d:::chipseq$rep2_df,
                              value_transformation = "log")
summary(idr_results)
```

---

estimate_idr2d	*Estimates IDR for Genomic Interaction Data*

Description

This method estimates Irreproducible Discovery Rates (IDR) between two replicates of experiments
identifying genomic interactions, such as Hi-C, ChIA-PET, and HiChIP.

Usage

```
estimate_idr2d(
  rep1_df,
  rep2_df,
  value_transformation = c("identity", "additive_inverse", "multiplicative_inverse",
    "log", "log_additive_inverse"),
  ambiguity_resolution_method = c("overlap", "midpoint", "value"),
  remove_nonstandard_chromosomes = TRUE,
  max_factor = 1.5,
  jitter_factor = 1e-04,
```

```
    max_gap = -1L,
    mu = 0.1,
    sigma = 1,
    rho = 0.2,
    p = 0.5,
    eps = 0.001,
    max_iteration = 30,
    local_idr = TRUE
)
```

Arguments

rep1_df   data frame of observations (i.e., genomic interactions) of replicate 1, with at
               least the following columns (position of columns matter, column names are ir-
               relevant):

column 1:	chr_a	character; genomic location of anchor A - chromosome (e.g., "chr3")
column 2:	start_a	integer; genomic location of anchor A - start coordinate
column 3:	end_a	integer; genomic location of anchor A - end coordinate
column 4:	chr_b	character; genomic location of anchor B - chromosome (e.g., "chr3")
column 5:	start_b	integer; genomic location of anchor B - start coordinate
column 6:	end_b	integer; genomic location of anchor B - end coordinate
column 7:	value	numeric; p-value, FDR, or heuristic used to rank the interactions

rep2_df   data frame of observations (i.e., genomic interactions) of replicate 2, with the
               following columns (position of columns matter, column names are irrelevant):

column 1:	chr_a	character; genomic location of anchor A - chromosome (e.g., "chr3")
column 2:	start_a	integer; genomic location of anchor A - start coordinate
column 3:	end_a	integer; genomic location of anchor A - end coordinate
column 4:	chr_b	character; genomic location of anchor B - chromosome (e.g., "chr3")
column 5:	start_b	integer; genomic location of anchor B - start coordinate
column 6:	end_b	integer; genomic location of anchor B - end coordinate
column 7:	value	numeric; p-value, FDR, or heuristic used to rank the interactions

value_transformation

               the values in x have to be transformed in a way such that when ordered in de-
               scending order, more significant interactions end up on top of the list. If the
               values in x are p-values, "log_additive_inverse" is recommended. The fol-
               lowing transformations are supported:

"identity"	no transformation is performed on x
"additive_inverse"	x. = -x
"multiplicative_inverse"	x. = 1 / x
"log"	x. = log(x). Note: zeros are replaced by .Machine$double.xmin
"log_additive_inverse"	x. = -log(x), recommended if x are p-values. Note: zeros are replaced by .Machine$double

               either "ascending" (more significant interactions have lower value in value
               column) or "descending" (more significant interactions have higher value in
               value column)

ambiguity_resolution_method

defines how ambiguous assignments (when one interaction in replicate 1 over-
laps with multiple interactions in replicate 2 or vice versa) are resolved. Avail-
able methods:

"value"	interactions are prioritized by ascending or descending value column (see sorting_direction), e.g., if two i
"overlap"	the interaction pair is chosen which has the highest relative overlap, i.e., overlap in nucleotides of replicate 1 in
"midpoint"	the interaction pair is chosen which has the smallest distance between their anchor midpoints, i.e., distance fror

remove_nonstandard_chromosomes

removes interactions containing genomic locations on non-standard chromo-
somes using [keepStandardChromosomes](#) (default is TRUE)

max_factor	numeric; controls the replacement values for Inf and -Inf. Inf are replaced by max(x) * max_factor and -Inf are replaced by min(x) / max_factor.
jitter_factor	numeric; controls the magnitude of the noise that is added to x. This is done to break ties in x. Set jitter_factor = NULL for no jitter.
max_gap	integer; maximum gap in nucleotides allowed between two anchors for them to be considered as overlapping (defaults to -1, i.e., overlapping anchors)
mu	a starting value for the mean of the reproducible component.
sigma	a starting value for the standard deviation of the reproducible component.
rho	a starting value for the correlation coefficient of the reproducible component.
p	a starting value for the proportion of reproducible component.
eps	Stopping criterion. Iterations stop when the increment of log-likelihood is < eps*log-likelihood, Default=0.001.
max_iteration	integer; maximum number of iterations for IDR estimation (defaults to 30)
local_idr	see [est.IDR](#)

Value

List with three components, (rep1_df, rep2_df, and analysis_type) containing the interactions
from input data frames rep1_df and rep2_df with the following additional columns:

column 1:	chr_a
column 2:	start_a
column 3:	end_a
column 4:	chr_b
column 5:	start_b
column 6:	end_b
column 7:	value
column 8:	"rep_value"
column 9:	"rank"
column 10:	"rep_rank"
column 11:	"idx"
column 12:	"rep_idx"
idr	IDR of the interaction and the corresponding interaction in the other replicate. If no corresponding interaction w

References

Q. Li, J. B. Brown, H. Huang and P. J. Bickel. (2011) Measuring reproducibility of high-throughput experiments. Annals of Applied Statistics, Vol. 5, No. 3, 1752-1779.

Examples

```
idr_results <- estimate_idr2d(idr2d:::chiapet$rep1_df,
                              idr2d:::chiapet$rep2_df,
                              value_transformation = "log_additive_inverse")
summary(idr_results)
```

---

estimate_idr2d_hic   *Estimates IDR for Genomic Interactions measured by Hi-C experiments*

Description

This method estimates Irreproducible Discovery Rates (IDR) of genomic interactions between two replicates of Hi-C experiments.

Before calling this method, call Juicer .hic contact matrix c

The contact matrix is subdivided into blocks, where the block size is determined by `resolution`. The reads per block are used to rank blocks and replicate blocks are easily matched by genomic location.

Usage

```
estimate_idr2d_hic(
  rep1_df,
  rep2_df,
  combined_min_value = 30,
  combined_max_value = Inf,
  min_value = -Inf,
  max_value = Inf,
  max_factor = 1.5,
  jitter_factor = 1e-04,
  mu = 0.1,
  sigma = 1,
  rho = 0.2,
  p = 0.5,
  eps = 0.001,
  max_iteration = 30,
  local_idr = TRUE
)
```

Arguments

rep1_df	data frame of either parsed .hic file from Juicer (output of [parse_juicer_matrix](#)) or parsed .matrix and .bed files from HiC-Pro (output of [parse_hic_pro_matrix](#)) for replicate 1
rep2_df	data frame of either parsed .hic file from Juicer (output of [parse_juicer_matrix](#)) or parsed .matrix and .bed files from HiC-Pro (output of [parse_hic_pro_matrix](#)) for replicate 2
combined_min_value	exclude blocks with a combined (replicate 1 + replicate 2) read count or normalized read count of less than combined_min_value (default is 20 reads, set combined_min_value = -Inf to disable)
combined_max_value	exclude blocks with a combined (replicate 1 + replicate 2) read count or normalized read count of more than combined_max_value (disabled by default, set combined_max_value = Inf to disable)
min_value	exclude blocks with a read count or normalized read count of less than min_value in one replicate (disabled by default, set min_value = -Inf to disable)
max_value	exclude blocks with a read count or normalized read count of more than max_value in one replicate (disabled by default, set max_value = Inf to disable)
max_factor	numeric; controls the replacement values for Inf and -Inf. Inf are replaced by max(x) * max_factor and -Inf are replaced by min(x) / max_factor.
jitter_factor	numeric; controls the magnitude of the noise that is added to x. This is done to break ties in x. Set jitter_factor = NULL for no jitter.
mu	a starting value for the mean of the reproducible component.
sigma	a starting value for the standard deviation of the reproducible component.
rho	a starting value for the correlation coefficient of the reproducible component.
p	a starting value for the proportion of reproducible component.
eps	Stopping criterion. Iterations stop when the increment of log-likelihood is < eps*log-likelihood, Default=0.001.
max_iteration	integer; maximum number of iterations for IDR estimation (defaults to 30)
local_idr	see [est.IDR](#)

Value

Data frame with the following columns:

column 1:	interaction	character; genomic location of interaction block (e.g., "chr1:204940000-204940000")
column 2:	value	numeric; p-value, FDR, or heuristic used to rank the interactions
column 3:	"rep_value"	numeric; value of corresponding replicate interaction
column 4:	"rank"	integer; rank of the interaction, established by value column, ascending order
column 5:	"rep_rank"	integer; rank of corresponding replicate interaction
column 6:	"idr"	integer; IDR of the block and the corresponding block in the other replicate

References

Q. Li, J. B. Brown, H. Huang and P. J. Bickel. (2011) Measuring reproducibility of high-throughput experiments. Annals of Applied Statistics, Vol. 5, No. 3, 1752-1779.

Examples

```
idr_results_df <- estimate_idr2d_hic(idr2d:::hic$rep1_df,
                                     idr2d:::hic$rep2_df)
summary(idr_results_df)
```

---

hic   *Example Hi-C data set*

Description

This object contains data from a Hi-C contact map of human chromosome 1 and a resolution of 2.5 * 10^6, extracted from GEO series GSE71831.

Usage

```
hic
```

Format

A list with two components, the data frames `rep1_df` and `rep2_df`, which have the following four columns:

column 1:	chr	character; genomic location of block - chromosome (e.g., "chr3")
column 2:	region1	integer; genomic location of block - coordinate A
column 3:	region2	integer; genomic location of block - coordinate B
column 4:	value	numeric; heuristic used to rank blocks, in this case: number of reads

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parse_hic_pro_matrix   *Parse .matrix and .bed files from HiC-Pro for IDR2D analysis*

Description

This function is used to convert the contact matrix from a HiC-Pro pipeline analysis run into an IDR2D compatible format. It takes one .matrix and one .bed file per replicate from HiC-Pro and returns the contact matrix for a specific chromosome for IDR2D analysis (see estimate_idr2d_hic)

Usage

```
parse_hic_pro_matrix(matrix_file, bed_file, chromosome = "chr1")
```

Arguments

`matrix_file`	path to .matrix file from HiC-Pro analysis run
`bed_file`	path to .bed file from HiC-Pro analysis run
`chromosome`	chromsome name to be analyzed, defaults to UCSC chromosome 1 (`"chr1"`)

Value

Data frame with the following columns:

column 1:	`chr`	character; chromosome of block (e.g., `"chr3"`)
column 2:	`region1`	integer; genomic location of side A of block in Hi-C contact matrix
column 3:	`region2`	integer; genomic location of side B of block in Hi-C contact matrix
column 4:	`value`	numeric; (normalized) read count in block

References

Servant, N., Varoquaux, N., Lajoie, B.R. et al. HiC-Pro: an optimized and flexible pipeline for Hi-C data processing. Genome Biol 16, 259 (2015) doi:10.1186/s13059-015-0831-x

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`parse_juicer_matrix`	*Parse .hic files from Juicer for IDR2D analysis*

Description

`parse_juicer_matrix` uses the Python package `hic-straw` internally to read .hic contact matrix files (see hic-straw on PyPI or the Aiden lab GitHub repository for more information).

The contact matrix is subdivided into blocks, where the block size is determined by `resolution`. The reads per block are used to rank blocks and replicate blocks are easily matched by genomic location.

Usage

```
parse_juicer_matrix(
  hic_file,
  resolution = 1e+06,
  normalization = c("NONE", "VC", "VC_SQRT", "KR"),
  chromosome = "chr1",
  use_python = NULL,
  use_virtualenv = NULL,
  use_condaenv = NULL
)
```

Arguments

`hic_file`	path to .hic file (either local file path or URL).
`resolution`	block resolution of Hi-C contact matrix in base pairs, defaults to 1,000,000 bp (usually one of the following: 2500000, 1000000, 500000, 250000, 100000, 50000, 25000, 10000, 5000)
`normalization`	normalization step performed by Python package `hic-straw`, one of the following: `"NONE"`, `"VC"`, `"VC_SQRT"`, `"KR"`.
`chromosome`	chromsome name to be analyzed, defaults to UCSC chromosome 1 (`"chr1"`)
`use_python`	if Python is not on PATH, specify path to Python binary here (see `use_python`)
`use_virtualenv`	if Python package `hic-straw` is not in base virtualenv environment, specify environment here (see `use_virtualenv`)
`use_condaenv`	if Python package `hic-straw` is not in base conda environment, specify environment here (see `use_condaenv`)

Value

Data frame with the following columns:

column 1:	`chr`	character; chromosome of block (e.g., `"chr3"`)
column 2:	`region1`	integer; genomic location of side A of block in Hi-C contact matrix
column 3:	`region2`	integer; genomic location of side B of block in Hi-C contact matrix
column 4:	`value`	numeric; (normalized) read count in block

References

Neva C. Durand, James T. Robinson, Muhammad S. Shamim, Ido Machol, Jill P. Mesirov, Eric S. Lander, and Erez Lieberman Aiden. "Juicebox provides a visualization system for Hi-C contact maps with unlimited zoom." Cell Systems 3(1), 2016.

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preprocess	*Prepares Data for IDR Analysis*

Description

This method removes invalid values, establishes the correct ranking, and breaks ties prior to IDR analysis.

`Inf` and `-Inf` are replaced by `max(x) * max_factor` and `min(x) / max_factor`, respectively.

`NA` values in `x` are replaced by `mean(x)`.

All values in `x` are transformed using the transformation specified in `value_transformation`.

Lastly, a small amount of noise is added to `x` to break ties. The magnitude of the noise is controlled by `jitter_factor`.

Usage

```
preprocess(
  x,
  value_transformation = c("identity", "additive_inverse", "multiplicative_inverse",
    "log", "log_additive_inverse"),
  max_factor = 1.5,
  jitter_factor = 1e-04
)
```

Arguments

x   numeric vector of values

value_transformation

the values in x have to be transformed in a way such that when ordered in descending order, more significant interactions end up on top of the list. If the values in x are p-values, `"log_additive_inverse"` is recommended. The following transformations are supported:

`"identity"`	no transformation is performed on x
`"additive_inverse"`	`x. = -x`
`"multiplicative_inverse"`	`x. = 1 / x`
`"log"`	`x. = log(x)`. Note: zeros are replaced by `.Machine$double.xmin`
`"log_additive_inverse"`	`x. = -log(x)`, recommended if x are p-values. Note: zeros are replaced by `.Machine$double`

either `"ascending"` (more significant interactions have lower value in value column) or `"descending"` (more significant interactions have higher value in value column)

max_factor   numeric; controls the replacement values for `Inf` and `-Inf`. `Inf` are replaced by `max(x) * max_factor` and `-Inf` are replaced by `min(x) / max_factor`.

jitter_factor   numeric; controls the magnitude of the noise that is added to x. This is done to break ties in x. Set `jitter_factor = NULL` for no jitter.

Value

numeric vector; transformed and stripped values of x, ready for IDR analysis

Examples

```
rep1_df <- idr2d:::chiapet$rep1_df
rep1_df$fdr <- preprocess(rep1_df$fdr, "log_additive_inverse")
```

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remove_nonstandard_chromosomes1d

Removes Peaks on Non-standard Chromosomes

Description

Removes Peaks on Non-standard Chromosomes

Usage

```
remove_nonstandard_chromosomes1d(x)
```

Arguments

x   data frame of genomic peaks, with the following columns (position of columns matter, column names are irrelevant):

column 1:	chr	character; genomic location of peak - chromosome (e.g., "chr3")
column 2:	start	integer; genomic location of peak - start coordinate
column 3:	end	integer; genomic location of peak - end coordinate
column 4:	value	numeric; p-value, FDR, or heuristic used to rank the peaks

Value

x without non-standard chromosomes.

Examples

```
rep1_df <- remove_nonstandard_chromosomes1d(idr2d:::chipseq$rep1_df)
```

---

remove_nonstandard_chromosomes2d

Removes Interactions on Non-standard Chromosomes

Description

Removes Interactions on Non-standard Chromosomes

Usage

```
remove_nonstandard_chromosomes2d(x)
```

Arguments

x
: data frame of genomic interactions, with the following columns (position of columns matter, column names are irrelevant):

column 1:	chr_a	character; genomic location of anchor A - chromosome (e.g., "chr3")
column 2:	start_a	integer; genomic location of anchor A - start coordinate
column 3:	end_a	integer; genomic location of anchor A - end coordinate
column 4:	chr_b	character; genomic location of anchor B - chromosome (e.g., "chr3")
column 5:	start_b	integer; genomic location of anchor B - start coordinate
column 6:	end_b	integer; genomic location of anchor B - end coordinate
column 7:	value	numeric; p-value, FDR, or heuristic used to rank the interactions

Value

x without non-standard chromosomes.

Examples

```
rep1_df <- remove_nonstandard_chromosomes2d(idr2d:::chiapet$rep1_df)
```

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